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ANTIBACTERIAL ACTIVITY OF ESSENTIAL OILS EXTRACTED FROM CITRUS FRUITS

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ABSTRACT

A comprehensive literature review was conducted on the antibacterial activity of essential oils extracted from citrus fruits, such as lemon, orange, mandarin and grapefruit. The chemical composition of these oils is complex and can vary due to factors such as temperature, altitude and growing conditions. Essential oils are known for their antimicrobial properties and are used in various industries. The antimicrobial action is specific and it is suggested to combine them with other preservation methods. Citrus peel is rich in essential oils, the extraction of which has been of interest to the pharmaceutical and cosmetic industries. These oils contain compounds such as limonene and terpenes, known for their antibacterial properties. The concentration of essential oils varies in different citrus fruits, and those with thicker skins tend to have higher concentrations. The extraction of essential oils is carried out using emerging, more efficient and environmentally friendly technologies. The evaluation of antimicrobial activity is based on the Minimum Inhibitory Concentration (MIC), which stops bacterial growth. The essential oils of lemon and eucalyptus showed bactericidal effect on various strains, while those of orange and mandarin showed less activity. The relationship between antibacterial activity and the chemical composition of essential oils is crucial, for example compounds such as monoterpenes for Salmonella control are essential for antimicrobial activity. The reviewed literature evidenced that citrus essential oils are a promising alternative to conventional food preservatives due to their antimicrobial properties.

Keywords: antibacterial activity, essential oils, citrus fruits, Minimum Inhibitory Concentration (MIC).

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1 Introduction

Citrus fruits include various species belonging to the genus "Citrus", such as lemon (*Citrus limon L.*), orange (*Citrus sinensis L.*), tangerine (*Citrus reticulata White*) and grapefruit (*Citrus paradisi Macfie.*). The antimicrobial efficacy of citrus fruits can vary due to the complexity of their chemical components. (Bozkurt, Aka, & Gulnaz, 2017)

Throughout history, more efficient and better quality essential oil extraction methods have been sought and their chemical composition is influenced by factors such as temperature, altitude, growing conditions, soil origin, and plant age. . (Jiménez, Zamora, Campoverde, Mariscal, & Walter, 2022) Essential oils (ECs) have natural antimicrobial properties and are obtained from a variety of aromatic plants and fruits. They exhibit a wide range of biological activities, which has made them widely used ingredients in food chemistry, pharmacology, pharmacy, and related fields. (Song, y otros, 2020)

The antimicrobial action of essential oils is highly specific and it is generally recommended to combine essential oils with other preservation methods to achieve biopreservation. (Denkova, y otros, 2020)

The use of emerging technologies for essential oil extraction focuses on sustainable methods that reduce environmental impact. Two notable examples of these technologies are supercritical fluid-assisted extraction, which uses carbon dioxide in a supercritical state to obtain essential oils efficiently and without generating harmful residues, and ultrasonic extraction, which uses sound waves to release aromatic compounds from plants. These innovations play an important role in preserving biodiversity by preventing overexploitation of natural resources. (Wong, Zárate, Veana, & Muñoz, 2021)

Evaluations of oils from different sources show that grapefruit and lemon essential oil are effective against pathogenic bacteria (*E. coli*), but less effective against beneficial bacteria (*L. plantarum* ATCC 8014 and *L. plantarum* ES147) in the gastrointestinal tract, which may be beneficial. (Raspo, Vignola, Andreatta, & Juliani, 2020)

Essential oils offer natural antimicrobial properties and diverse applications in different industries, but their use must be complemented with other preservation methods and factors that affect their chemical composition must be considered. The aim of this article is to investigate the antibacterial activity of essential oils extracted from citrus fruits.

2 Citrus fruits

Citrus fruits are in predominant demand in the food industry, especially in the beverage sector, due to their high juice content that is used in the production of items such as juices and nectars. The various citrus varieties used include oranges (*Citrus sinensis*), tangerines (*C. unshi*, *C. tangerine*, *C. reticulata*, *C. clementine*), lemons (*C. lemon*), limes (*C. aurantifolia/latifolia*), grapefruit (*C. paradisi*) and grapefruits (*C. grandis*). (Wong, Aguilar, Veana, & Muñoz, 2020)

1. Waste

Citrus fruits have different chemical compositions as reported in Table 1.

0,61 Chemical Components of Some Citrus Fruits

Fruit	Energy (kcal)	Water (g)	carbohydrates (g)	Dietary fiber (g)	Fats (g)	Proteins (g)
Citrus sinensis	38	86	8,6	2,3	0,3	0,8
citrus lemon	39	87	9	1	0,3	0,7
Citrus reticulata	39	86	9	Fruit	Energy (kcal)	Water (g)
Carbohydrates (g)	Dietary Fiber (g)	Fat (g)	Protein (g)	Citrus sinensis	38	86

Fountain: (Cazar, 2016)

It is relevant to note that the chemical structure of citrus fruits can change depending on the species, however, citrus fruits are a remarkable source of essential nutrients and beneficial elements for well-being.

3 Essential oils

Essential oils are highly concentrated, aromatic and volatile extracts that can be obtained from the bark, leaves, flowers and fruits of various plants. (López, 2014)

The extraction of essential oils from waste has attracted the attention of the pharmaceutical and cosmetic industry due to its rich chemical composition, which includes alcohols, aldehydes, ketones, phenols, esters, ethers, and terpenes in different proportions. Many of these components are generally considered "safe" for use. (Mitidieri, Barbieri, Brambilla, & Piris, 2021)

It is possible to obtain these essential oils from the peel of the lemon, as it contains components such as monoterpene hydrocarbons, limonene, pinene and gamma-terpinene, which have been shown to have outstanding antibacterial properties. (Alvarez, Rodriguez, & Polo, 2022)

The components of oils are classified into terpenoids and non-terpenoids:

Terpenoids are essential and widely marketed in the industry due to their complex structure and presence in plants, especially citrus species.

Non-terpenoid compounds include aliphatic, aromatic, sulphur and nitrogenous substances, although they are less important in terms of utility. (Sharmeen, Mahomoodally, Zengin, & Maggi, 2021).

Table 2 shows the percentage yield of essential oil from citrus peels.

0,21%2 Yield Percentage of Citrus Peel Essential Oil

8,6	2,3
0,3	0,8
<i>Lemon Citrus</i>	39
87 Meski	9
1 Risso	0,3
0,7	Citrus reticulata
39	86
9	1,9

Fuete: (Urrunaga, Carpio, Gutierrez, & Tomaylla, 2022)

Citrus rinds harbor significant amounts of essential oils (0.5 to 3.0 kg per ton of fruit). Citrus fruits with thicker skins, such as sour orange (*Citrus aurantium*), grapefruit (*Citrus paradisi*) and bergamot (*Citrus bergamia*), show a high concentration of essential oils in contrast to thin-skinned citrus varieties. (Urrunaga, Carpio, Gutierrez, & Tomaylla, 2022)

2. Essential Oil Extraction

Extraction methods are known as emerging or unconventional technologies, those that have the potential to improve and optimize the process of obtaining essential oils from plants and natural products, in addition, they seek to be more efficient, environmentally friendly and, in some cases, more economical than traditional extraction methods. (Biamonte, 2017)

According to previous research, these approaches have the ability to improve yields in obtaining bioactive compounds. They are more accessible and easier to operate, generating high-quality compounds in significant quantities without requiring large amounts of solvents or high energy consumption. This is due to the short times required for extraction, which typically range from 5 to 40 minutes. (Caputo, Quintieri, Cavalluzi, & Lentini, 2018)

*Conventional extraction, **Unconventional extraction.3MAE 2: 8 min at 100 °C

0,19	0,8	Citrus paradisi	32
90	8,1	1.1 Extraction time: 8 h Temperature: 17°C to 20°C	0,1 Ethanol: 23.65% Water: 6.53%
	Ultrasound assisted extraction**	Solvents: ethanol:water (1:1), ethanol and water Extraction time: ultrasound power at 50%, 5 min and ultrasound power at 100% for 30 min. Temperature: 25°C to 30°C	Ethanol/water (50%): 36.92%; Ethanol/water (100%): 41.38% Ethanol (50%): 32.74%; Ethanol (100%): 40.90% Water (50%): 43.20%; Water (100%): 44.94%
	Microwave assisted extraction**	Solvents: ethanol:water (1:1), ethanol and water Extraction time: 3s to 20s Temperature: 80°C to 85°C	Ethanol/water: 9.52% Ethanol: 30.21% Water: 22.73%
<i>C. aurantifolia</i> <i>Swingle</i>	Maceration*	Solvents: 120 mL of 1% citric acid solution. Extraction time: 1 hour. Temperature: 90°C	Total phenols = 34.00 ± 0.04 – 37.67 ± 0.05 mg/g extract
	Yield (g oil/g sample)	Citrus jambhiri Extraction time: 5 min, 800 W. Temperature: 120°C	0,72%*
	1,06%	Citrus sinensis var.	2.31%

<i>Citrus grandis</i>		meski Temperature: 40°C	80% ethanol= 18.46% 50% ethanol= 15.64% 100% ethyl acetate= 5.12%
	1,75%	Citrus aurantium Linn. Extraction time: 60 min Temperature: 45 °C Frequency: 35 kHz	1.29% 80% ethanol= 19.24% 100% methanol= 13.84%
<i>Citrus paradisi</i>	0,79%	Citrus limon (L.) Burm. Extraction time: 24 hours. Temperature: 50°C	0,21%
	Microwave assisted extraction**	Solvent: water Power: 10 W MAE 1: 20 min at 80 °C MAE 2: 8 min at 100 °C MAE 3: 20 min at 100 °C	Interval between treatments between 18.0% to 21.5%

*extracción convencional, **extracción no convencional.

Fountain: (Wong, Aguilar, Veana, & Muñiz, 2020)

4 Bacteria

Bacteria require a minimal amount of nutrients for their growth, including water, a carbon source, a nitrogen source, and some mineral salts. These nutrients are used to carry out the biosynthesis of new cellular components, which requires energy from the environment. (Caycedo, Corrales, & Trujillo, 2021)

At present, a wide variety of microorganisms have been identified, comprising thousands of species. Some of these microorganisms are able to survive in extreme environments. In nature, they perform vital functions, such as decomposing organic matter and participating in biogeochemical cycles. (Rios & Bardales, 2022) (Molina, Paños, Reyes, & Ruiz, 2021)

These microorganisms, which are less than 1.0 mm in size according to our anthropocentric perception based on the resolving capacity of the human eye, are single-celled organisms that cannot be seen with the naked eye, but can be detected by using conventional light microscopes or, in greater detail, by scanning and transmission electron microscopes. (Sanchez, 2018)

Microbiological causes generate negative changes in food, however, there is a perception about the role of microorganisms, often relating them to unhygienic environments and diseases. (Ballesteros, Paños, & Ruiz, 2018)

Microorganisms obtain food from the substrate where they grow and secrete enzymes to break down complex compounds into simpler soluble foods. Enzyme activity is affected by temperature, pH, the presence of salts, and time. Microorganisms can survive in a wide temperature range and some require oxygen to grow, while others can thrive in the absence of oxygen. pH also influences growth, with some bacteria and fungi preferring low pH, but the optimal range for bacteria is 6.0 to 8.5. (Cervantes, Orihuela, & Rutiaga, 2017)

c. Different letters in the same column indicate significant differences between the strains tested (P<0.05)4 Diameters of inhibition halos (mm) of AE on strains tested

	Citric	Type of extraction	Conditions
<i>Yield</i> ES147	C. aurantium	Maceration*	Solvents: ethanol: water (1:1), ethanol and water
<i>Ethanol/Water:</i> 29.28% Ethanol: 23.65%	7,17 ± 2,56 b	Ultrasound-assisted extraction**	Solvents: ethanol: water (1:1), ethanol and water
<i>Ethanol/water</i> (50%): 36.92%; Ethanol/water (100%): 41.38%	14,50 ± 3,15 c	Microwave- assisted extraction**	Solvents: ethanol: water (1:1), ethanol, and water
<i>Ethanol/Water:</i> 9.52% Ethanol: 30.21%	C. aurantifolia Swingle	Maceration*	Solvents: 120 mL of 1% citric acid solution.

Total phenols = 34.00 ± 0.04 \u2012 37.67 ± 0.05 mg/g extract

b. ni: no inhibi3o

Microwave-assisted extraction**

(Vignola, Serra, & Andreatta, 2020)

3. Determination of the minimum inhibitory concentration (MIC)

Assessment of the susceptibility of bacteria to antimicrobials can be carried out using phenotypic (such as dilution and diffusion), biochemical, or genetic methods. Among them, phenotypic methods are the most commonly used initially due to their simplicity when it is desired to determine the sensitivity of microorganisms to a new antimicrobial compound. While diffusion techniques provide us with a preliminary assessment of the antimicrobial capacity of the compounds we are studying, dilution techniques allow us to determine the Minimum Inhibitory Concentration (MIC). (Pinheiro, y otros, 2022) (Muthular, y otros, 2019)

The Minimum Inhibitory Concentration (MIC) is the minimum amount of an antimicrobial (measured in µg/mL) needed to stop the visible growth of a microorganism after 24 hours of incubation at 37°C. MIC is considered the most reliable method for assessing antimicrobial susceptibility, outperforming other methods. In addition to detecting rare resistances, it provides definitive answers when other methods are inconclusive. (Horna, Silva, Vicente, & Tamariz, 2005)

CMI, was determined in applications of essential oils that act as antimicrobial agents that are frequently studied for their ability to protect food against pathogenic microorganisms (Gonzalez, Salazar, & Fuertes, 2022)

The antimicrobial activity of different ECs (lemon, orange, tangerine, bay leaf, and eucalyptus) in beneficial bacteria such as *Lactobacillus plantarum* ES147 and ATCC 8014, pathogens such as *Escherichia coli* ATCC 25922 and a food-spoilage bacterium, *Leuconostoc mesenteroides* MS1. Oils with significant inhibition halos were tested for minimal inhibitory and bactericidal concentrations. Lemon and eucalyptus

showed bactericidal effect on all strains. Orange and tangerine had little antimicrobial effect. Laurel, tangerine, and orange had little impact on beneficial bacteria and generated large zones of inhibition in pathogenic or food spoilers. (Vignola, Serra, & Andreatta, 2020)

4. Relationship of antibacterial activity

The antimicrobial activity of citrus peel essential oils, with antimicrobial properties, may be an alternative to preservatives in food, taking advantage of a by-product of the juice industry. Its effectiveness varies according to the chemical composition, influenced by cultivation and extraction. (Perez, y otros, 2017)

Fruits have been found to contain a wide variety of nutritional compounds, including antioxidants such as phenolic acids, anthocyanins, carotenoids, flavonoids, vitamin C, and vitamin E. Therefore, incorporating more of these fruits into the diet helps prevent chronic diseases

(Mora, Cabrera, Alarcón, & Garcia, 2022)

5 Assessment of the antibacterial activity of citrus essential oils

(Torrenegra, Pajaro, & Leon, 2017) According to them, monoterpenes are the most abundant volatile metabolites in EAs. The main compound found is limonene for both HD and MWHD methods in all the samples evaluated, the identification of the components, retention times and abundance percentages are reported in the [Table 5](#).

Relative abundance (%) of essential oils Major components detected in the AEs obtained by the method of steam entrainment hydrodistillation (HD) and microwave-assisted hydrodistillation (MWHD).

Solvent: 60 mL of 1% citric acid solution.	Total phenols = 38.00 ± 0.02 \u2012 41.43 mg/g extract							
	Maceration*		Solvents: ethanol, methanol, acetone, ethyl acetate (50%, 80%, 100%) Material/solvent ratio: 1:15 w/v		The highest yield was using ethanol.		C. paradisi	
	Solvents: ethanol, methanol (50%, 80%, 100%) Material:solvent ratio: 1:20 w/v	The highest yield was using ethanol.	C. medica L. C. sinensis L. C. lemon L	Hot Water Extraction*	Solvent: preheated water	Range from 18.0% to 21.5%	HD	Microwave-assisted extraction**
Solvent: water Power: 10 W	Interval between treatments between 18.0% to 21.5%	*Conventional extraction, **Unconventional extraction.	1.68 (9.09)	1.77 (9.00)	0.70 (9.91)	0.90 (9.91)	0.45 (9.98)	0.65 (9.88)
β-mireno	1.17 (12.01)	1.39 (12.45)	Lemon	Orange	Tangerine	Lactobacillus plantarum	10.00 ± 3.40 b	7.17 ± 0.98 A

7.67 ± 0.52 A	Lactobacillus plantarum	7.17 ± 2.56 b	nor	6.67 ± 0.82 A	Escherichia coli	14.50 ± 3.15 C	5.50 ± 0.63 A	6.50 ± 0.55A
Leuconostoc mesenteroides	7.00 ± 2.28 to	nor	nor	to. Diameter of inhibition halo including 5 mm disk, expressed as the average of 3 replicates	B. Ni: Did not inhibit	c. Different letters in the same column indicate significant differences between the strains tested (P<0.05)	0.54 (15.90)	0.66 (16.09)

Tiempo de retención (t_R)

Abundancia relativa (%) de los aceites esenciales

On the other hand, the results of the evaluation of the antibacterial susceptibility of the essential oils obtained by the HD and MWHD methods (Table 1) allowed us to identify the extracts capable of inhibiting the growth of the bacterial strains, taking as a selection criterion, those that were able to inhibit the three strains by more than 90% (Table 6). Therefore, the MIC was determined (Table 7).

98.00±0.336 Antibacterial sensitivity of essential oils.

Samples	Bacterial growth inhibition percentage				
	<i>S. aureus</i>	<i>S. epidermidis</i>	<i>K. pneumoniae</i>	<i>P. aeruginosa</i>	<i>E. coli</i>
<i>C. sinensis</i> HD	Compound	Percentage relative abundance (tR, min)*	92,50±0,33	<i>C. sinensis</i>	<i>C. aurantifolia</i>
<i>C. reticulata</i> MWHD	<i>C. paradisi</i>	91,03±0,30	HD	MWHD	HD
MWHDHD	HD	MWHD	HD	MWHD	α-pinene
0.38 (9.89)**MWHD	0.59 (9.90)	1.68 (9.09)	1.77 (9.00)	0.70 (9.91)	0.90 (9.91)
0.45 (9.98)HD	0.65 (9.88)	β-Mireno.	1.17 (12.01)	1.39 (12.45)	1.00 (12.01)
1.09 (12.45)MWHD	0.90 (12.10)	1.00 (12.05)	6.11 (11.89)	6.25 (11.99)	Limonene
88.80 (13.05)HD	0.20 (13.01)	69.28 (13.36)	78.80 (13.61)	92.22 (13.34)	93.55 (13.55)
72.11 (12.90)MWHD	75.22 (12.91)	Linalool	0.60 (15.09)	0.89 (15.56)	14.60 (18.14)
17.89 (18.09)	3.03 (14.79)	3.89 (14.88)	0.54 (15.90)	0.66 (16.09)	Retention Time (tR)

(Ambrosio, y otros, 2019) According to the IMC, it is determined with the use of inoculated and standardized broth, to which solutions of the essential oil at different concentrations are added, causing dilution. The values are presented in the [board 6007](#) Minimum inhibitory concentration (MIC)

Relative abundance (%) of essential oils	Essential oil	Extraction method	CMI ($\mu\text{g/mL}$)
Percentage of Bacterial Growth Inhibition	<i>Samples</i>	S. aureus	S. epidermidis
		E. coli	C. sinensis
	90.00 \pm 1.00	92.50 \pm 0.33	92.98 \pm 0.25
		90.90 \pm 0.50	91.03 \pm 0.30
	93.55 \pm 0.66	92.80 \pm 0.11	C. paradisi
		92.50 \pm 0.33	91.98 \pm 0.25
	<i>C. paradisi</i>	91.00 \pm 0.33	90.33 \pm 0.11
		92.78 \pm 0.11	C. aurantifolia
90.10 \pm 0.89	90.23 \pm 0.05	91.09 \pm 0.78	91.07 \pm 0.78
		90.99 \pm 0.25	90.47 \pm .33
	92.98 \pm 0.33	92.17 \pm 0.67	C. reticulata
		92.08 \pm 0.15	92.00 \pm 0.22
	<i>C. reticulata</i>	91.00 \pm 0.33	91.00 \pm 0.25
		92.22 \pm 0.35	Gentamicin (control)
	98.00 \pm 0.50	Phone: 97.00 \pm 0.55	98.00 \pm 0.25
		MWHD	900
Bacterial strains	<i>Essential oil</i>	Extraction Method	MIC ($\mu\text{g/mL}$)
		HD	900
	<i>C. paradisi</i>	MWHD	900
		HD	1000
	<i>C. aurantifolia</i>	MWHD	1000
		HD	1000
	<i>C. reticulata</i>	MWHD	1000
		HD	900
<i>P. aeruginosa</i>	<i>C. sinensis</i>	MWHD	900
		HD	1000
	<i>C. paradisi</i>	MWHD	900
		HD	1000
	<i>C. aurantifolia</i>	MWHD	1000
		HD	1000
	<i>C. reticulata</i>	MWHD	1000
		HD	900
<i>E. coli</i>	<i>C. sinensis</i>	MWHD	900
		HD	700

<i>C. paradisi</i>	MWHD	600
	HD	800
<i>C. aurantifolia</i>	MWHD	700
	HD	900
<i>C. reticulata</i>	MWHD	800
	HD	800

6 Conclusions

In conclusion, citrus fruits of the genus "Citrus" have a diverse chemical composition that includes essential components such as limonene, alcohols, aldehydes and terpenoids, which the essential oils extracted from these fruits demonstrate antimicrobial and preservative properties, presenting a potential as a promising alternative to conventional preservatives in the food industry.

The antimicrobial activity of these oils is linked to their chemical composition and varies according to the bacterial strains studied, the extraction methods, Although some citrus essential oils have a selective effect on pathogenic bacteria without significantly affecting beneficial bacteria, it is essential to balance their use to maintain the health of the microbiota, however research on the antibacterial activity of citrus essential oils highlights their potential as natural antimicrobial agents in the food industry, requiring a deeper understanding of their action on different strains and their Interaction with other preservation methods for optimal application.

7 Recommendation

It is advisable to conduct research that focuses on optimizing these methods. This could include evaluating different extraction techniques to maximize the yield of bioactive compounds and ensure efficient and sustainable production of citrus essential oils.

8 Bibliography

- Alvarez, A. B., Rodriguez, D. R., & Polo, J. A. (2022). Antibacterial effect of the peel essential oil of citrus latifolia tanaka "limon" on staphylococcus aureus compared with oxacillin. *Peruvian Journal of Integrative Medicine*.
- Ambrosio, C., Ikeda, N., Miano, A., Saldaña, E., Moreno, A., Stashenko, E., . . . Gloria, E. (2019). Deciphering the selective antibacterial activity and chemical composition of citrus essential oils. *Scientific Reports* .
- Badr, R., Alokalah, B., & Al-Amir, L. (2021). Chemical composition and antibacterial activity of the peel essential oils extracted from citrus fruits. *Agriculture and Applied Biology*, 114-123.
- Ballesteros, M., Paños, E., & Ruiz, J. (2018). Microorganisms in Primary Education. Conceptions in children. *Journal of Research and Didactic Experiences*.
- Biamonte, J. W. (2017). Quantum machine learning. *Nature*, 549(7671), 195-202. Quantum Artificial Intelligence (Quantum AI). *Blockchain Technology*, pp. Nature, 549(7671), 195-202.

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- Bozkurt, T., Aka, Y., & Gulnaz, O. (2017). Chemical composition of the essential oils from some citrus species and evaluation of the antimicrobial activity. *Environmental Science, Toxicology and Food Technology*, 29-33.
- Caputo, L., Quintieri, L., Cavalluzi, M., & Lentini, G. (2018). Antimicrobial and Antibiofilm Activities of Citrus Water-Extracts Obtained by Microwave-Assisted and Conventional Methods. *Biomedicines*.
- Caycedo, L., Corrales, L., & Trujillo, D. (2021). Bacteria, Their Nutrition and Growth: A Look from Chemistry. *Scielo*.
- Hunt, I. (2016). *Physico-chemical analysis to determine the quality of fruits*.
- Cervantes, J., Orihuela, R., & Rutiaga, J. (2017). About the Development and Control of Microorganisms in Papermaking. *Technological awareness*.
- Denkova, R., Teneva, D., Tomova, T., Goranov, B., Denkova, Z., Shopska, V., . . . Hristova, Y. (2020). Chemical composition, antioxidant and antimicrobial activity of essential oils from tangerine (*Citrus reticulata* L.), grapefruit (*Citrus paradisi* L.), lemon (*Citrus lemon* L.) and cinnamon (*Cinnamomum zeylanicum* Blume). *DE GRUYTER*, 175-185.
- Gonzalez, K., Salazar, M., & Fuertes, C. (2022). Antibacterial activity of *Minthostachys mollis* Griseb essential oils. "Muña" and *Piper carpubunya* Ruiz & Pav. "Pinku." *Science and Research*.
- Granados-Chinchilla, F., & Andrea Molina. (2019). Chemical composition and antimicrobial activity of *Psidium guajava* and *Cymbopogon citratus* essential oil. *Mesoamerican Agronomy*, vol. 30, no. 1, pp. 147-163.
- Horna, G., Silva, M., Vicente, W., & Tamariz, J. (2005). Minimum inhibitory concentration and minimum bactericidal concentration of ciprofloxacin in uropathogenic bacteria isolated at the National Institute of Neoplastic Diseases. *Revista Médica Herediana*.
- Hsouna, A., Halima, N., Delgado, S., & Hamdi, N. (2017). Citrus lemon essential oil: chemical composition, antioxidant and antimicrobial activities with its preservative effect against *Listeria monocytogenes* inoculated in minced beef meat. *Lipids in Health and Disease*.
- Jimenez, W., Zamora, J., Campoverde, J., Mariscal, & Walter. (2022). Antioxidant and antimicrobial activity of citrus *sinensis*, *citrus paradisi*, and *citrus reticulata* essential oil. *Reciamuc*, 399-407.
- Klimek, M., Szopa, A., & Ekiert, H. (2020). Citrus limon (Lemon) Phenomenon—A Review of the Chemistry, Pharmacological Properties, Applications in the Modern Pharmaceutical, Food, and Cosmetics Industries, and Biotechnological Studies. *Plants*.
- Lopez, T. (2014). Essential oils. *Elsevier*, 88-91.
- Miladys Esther Torrenegra Alarcón, & Nerlis Paola Pájaro. (2017). In vitro antibacterial activity of essential oils of different species of the genus *Citrus*. *Scientific research article / <http://dx.doi.org/10.15446/rcciquifa.v46n2.67934>*.
- Mitidieri, M., Barbieri, M., Brambilla, M., & Piris, E. (2021). In vitro effect of lemon essential oil and garlic extract on *Monilinia fructicola* growth. *Scielo*.

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- Molina, J., Paños, E., Reyes, J., & Ruiz, J. (2021). Microorganisms and hygiene habits. Longitudinal study in the initial years of Primary Education. *Eureka Journal on Science Teaching and Dissemination*.
- Mora, J., Cabrera, D., Alarcón, A., & Garcia, F. (2022). Comparative study of polyphenols or total phenols and antioxidant activity of the citrus sinensis shell. *RECIAMUC*, 459-469.
- Muthular, M., Passero, P., Fernanda, B., Jewtuchowicz, V. M., Brusca, M., & Pérez, C. (2019). Inhibitory effect of diethylstilbestrol on clinical strains of *Candida albicans* susceptible and resistant to azoles. *ScienceDirect*.
- Pérez, A., Vitola, D., Villareal, J., Noya, B., Pérez, P., Ramírez, S., & Rangel, P. (2017). Antimicrobial Activity of Sweet Orange (*Citrus Sinensis*) and Creole Lemon (*Citrus Aurantifolia*) Essential Oils. *LIMENTECH*.
- Pinheiro, C., Perez, A., Pereira, J., Abrantes, J., Souza, L., Araújo, M., . . . Oliveira, E. (2022). Investigation on mechanism of antifungal activity of citral against *Cladosporium sphaerospermum* Penz. *Annals of Biology*, 43-53.
- Raspo, M., Vignola, M., Andreatta, A., & Juliani, R. (2020). Antioxidant and antimicrobial activity of citrus essential oils . *Food Bioscience*, 2-27.
- Ríos, O., & Bardales, L. (2022). Effect of effective microorganisms (me), in the rearing of Nile tilapia. *Journal of Amazonian Veterinary and Zootechnics*.
- Rivas Yauce, M. A. (2020). *Evaluation of lemon essential oil (Citrus aurantifolia swingle) as a natural preservative in pork (Sus scrofa domesticus)*. But.
- Sanchez, J. (2018). Microorganisms in human life and chronic degenerative diseases. *Journal of the Andean Jungle Research Society*.
- Sharmeen, J., Mahomoodally, F., Zengin, G., & Maggi, F. (2021). Essential Oils as Natural Sources of Fragrance Compounds for Cosmetics and Cosmeceuticals. *Molecules*.
- Song, X., Liu, T., Wang, L., Liu, L., Li, X., & Wu, X. (2020). Antibacterial Effects and Mechanism of Mandarin (*Citrus reticulata* L.) Essential Oil against *Staphylococcus aureus*. *College of Food Engineering and Nutrition Science*, 1-10.
- Suárez Rodríguez, E. B. (2012). Evaluation of the Antibacterial Activity of Lemon Grass Essential Oils (*Cymbopogon citratus* DC. Stapf), eucalyptus (*Eucalyptus* ssp.) and cloves (*Syzygium aromaticum* L), alone and in combination, against *Pseudomonas aeruginosa* ATCC 27853, . *Tese em Espanhol / LILACS, MOSAIC - Integrative Health / ID: biblio-877509*.
- Suárez, Z., Tobar, M., Pérez, J., Hurtano, A., & Ospina, J. (2017). Evaluation of the inhibitory capacity of essential oils in *Staphylococcus aureus* and *Escherichia coli*. *Biotechnology in the Agricultural and Agroindustrial Sector*.
- Torrenegra, M., Pájaro, N., & León, G. (2017). In vitro antibacterial activity of essential oils of different species of the genus *Citrus*. *Colombian Chemical Sciences - Pharmaceuticals*.
- Torres, L., & Franco, Y. (2015). *EVALUATION OF THE PROCESS OF EXTRACTING ESSENTIAL OILS FROM LEAVES OF Citrus aurantifolia (SUBTLE LEMON)*,

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Citrus sinensis (ORANGE) AND *Citrus nobilis* (MANDARIN) BY HYDRODISTILLATION. Quevedo.

- Urrunaga, M., Carpio, C., Gutierrez, G., & Tomaylla, C. (2022). Physicochemical properties, chemical composition and antioxidant activity of Citrus jambhiri (Rough Lemon) essential oil. *Journal of the Chemical Society of Peru*.
- Vignola, M., Serra, M., & Andreatta, A. (2020). Antimicrobial Activity of Various Essential Oils on Beneficial, Pathogenic and Food Spoilers Bacteria. DOI: <https://doi.org/10.33414/rtyc.37.92-100.2020>.
- Wong, J., Aguilar, P., Veana, F., & Muñoz, D. (2020). Impact of green extraction technologies for obtaining bioactive compounds from citrus fruit residues. *Journal of Chemical-Biological Sciences*, 1-11.
- Wong, J., Zárate, P., Veana, F., & Muñoz, D. (2021). Impact of green extraction technologies for obtaining bioactive compounds from citrus fruit residues. *Scielo*.