



Evaluation of antibiotic sensitivity pattern in clinical isolates of *Pseudomonas aeruginosa* in Dhiraj Hospital, Vadodara, Gujarat.

¹Dr. Harsh vinubhai singel, ²Dr. Tanuja javadekar

¹Assistant Professor, Microbiology Department, Smt. B. K. Shah Medical Institute & Research Centre, Sumandeep Vidyapeeth Deemed to be university, Piparia, Vadodara, Gujarat

²Professor & Head, Microbiology Department, Smt. B. K. Shah Medical Institute & Research Centre, Sumandeep Vidyapeeth Deemed to be university, Piparia, Vadodara, Gujarat.

Corresponding Author: **Dr harsh vinubhai singel**

Article History

Volume 6 Issue 12, 2024

Received: 25 May 2024

Accepted: 30 June 2024

doi:

10.48047/AFJBS.6.12.2024.1735-1741

ABSTRACT:

INTRODUCTION- *Pseudomonas aeruginosa* is a straight or slightly curved, motile gram-negative bacilli, strict aerobe that belongs to the family Pseudomonadaceae. It represents a phenomenon of antibiotic resistance and exhibits all known mutational and enzymatic mechanisms of bacterial resistance thus imperilling the selection of appropriate treatment.

METHOD- Fifty consecutive isolates of *Pseudomonas aeruginosa* obtained from various clinical specimens were processed at Central Microbiology Laboratory of Dhiraj Hospital between the periods of December 2023 to May 2024. Specimens were cultured on Blood agar, MacConkey agar and Nutrient agar and incubated at 37°C for 24 hours. Colonies were identified by its characteristics and biochemical reactions. **RESULTS-** A total of 50 isolates of *Pseudomonas aeruginosa* was found from clinical samples with 56% samples from pus, 24% from sputum, 5% from endotracheal tube secretion, 2% from CSF, and 8% from others. Most of samples were of male 72%. **CONCLUSION:** Healthcare settings largely contribute as reservoirs of pathogenic strains of *Pseudomonas aeruginosa*. They are known to utilize their highest levels of intrinsic and acquired resistance mechanisms against most antibiotics.

Key word: *Pseudomonas aeruginosa* (*P. aeruginosa*), chronic obstructive pulmonary disease (COPD), ventilator-associated pneumonia (VAP), multi-drug resistance (MDR).

INTRODUCTION:

Pseudomonas aeruginosa is a straight or slightly curved, motile gram-negative bacilli, strict aerobe that belongs to the family Pseudomonadaceae.

It is the pseudomonad most frequently recovered from clinical specimens. It is increasingly recognized as emerging opportunistic pathogen of clinical relevance which causes infection in patients with burn wounds, cystic fibrosis, organ transplants, urinary tract and lower respiratory tract infections which can be severe and even life-threatening in immunocompromised hosts.

It represents a phenomenon of antibiotic resistance and exhibits all known mutational and enzymatic mechanisms of bacterial resistance thus imperiling the selection of appropriate treatment¹.

Pseudomonas aeruginosa is a multi-drug resistance (MDR) opportunistic pathogen, causing acute or chronic infection in immunocompromised individuals with chronic obstructive pulmonary disease (COPD), cystic fibrosis, cancer, traumas, burns, sepsis, and ventilator-associated pneumonia (VAP) including those caused by COVID-19⁴⁻⁶. *P. aeruginosa* in biofilm states may survive in a hypoxic atmosphere or other extremely harsh environments⁷⁻⁸. This paper broadly reviews the recent progress in *P. aeruginosa* research towards the regulatory and functional mechanisms of virulence factors, gene expression regulators, secretion systems, quorum sensing, and antibiotic resistance, as well as host-pathogen interaction, new technologic advances, and therapeutic development (Fig.1)

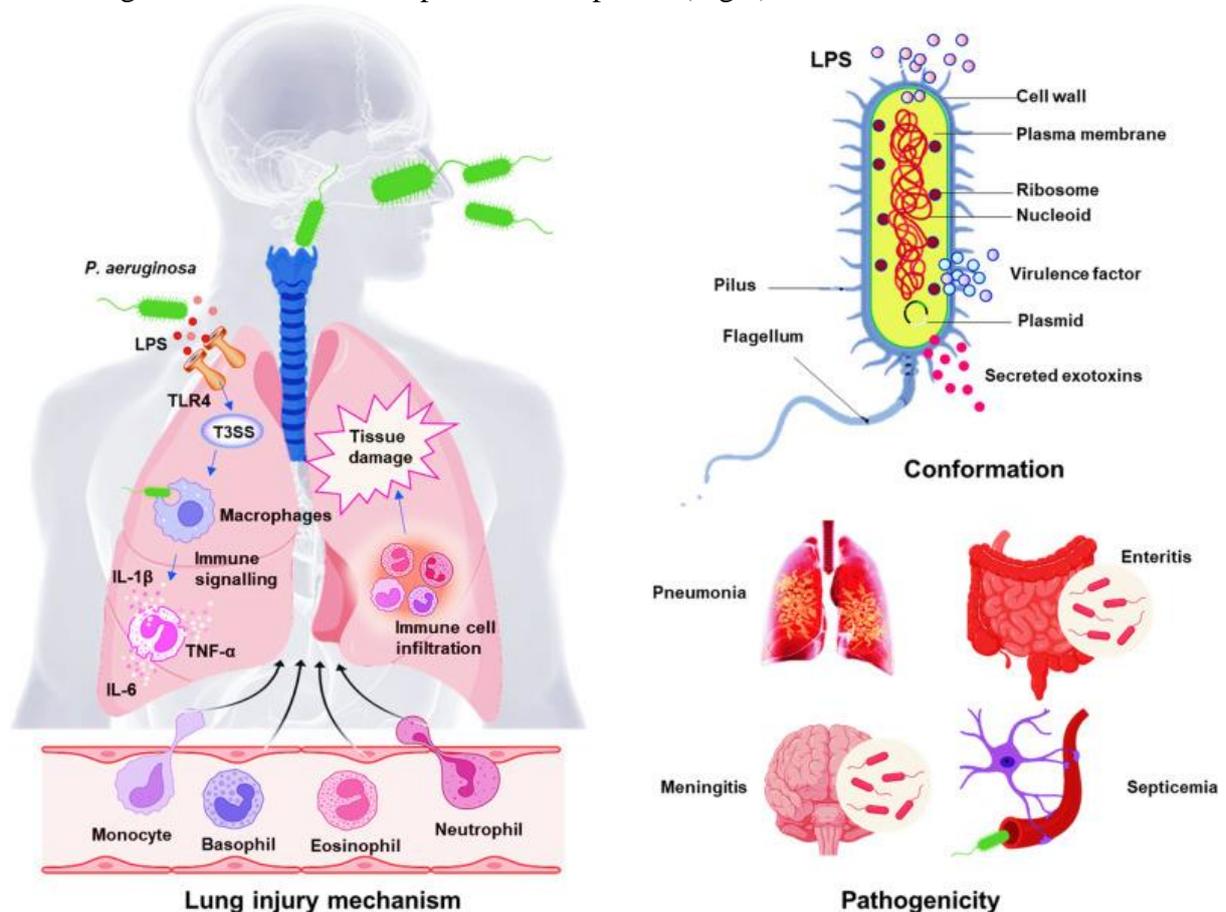


Fig-1: broadly reviews

ANTIMICROBIAL RESISTANCE MECHANISMS-

antimicrobial resistance in *P. aeruginosa* include outer membrane porins and permeability alterations, efflux pumps, antibiotic-inactivating enzymes, and target binding site mutations. Many resistance mechanisms are often present and expressed simultaneously in a given patient with a *P. aeruginosa* infection. The terms MDR, extensively drug resistant (XDR), and pandrug-resistant (PDR) are often used to characterize the different patterns of multidrug resistance exhibited by *P. aeruginosa*. An MDR isolate is nonsusceptible to at least one agent in three or more antibiotic classes with intrinsic activity. An XDR isolate is nonsusceptible to

at least one agent in all but two or fewer antibiotic classes with intrinsic activity, and a PDR isolate is nonsusceptible to all agents with intrinsic activity.

Resistance mechanisms present in *P. aeruginosa* can be classified as intrinsic, acquired, or adaptive. Intrinsic resistance mechanisms stem from genes that encode the inherent properties of cell structures and composition that provide protection against toxic molecules and antimicrobials. Acquired resistance mechanisms result through mutation of intrinsic genes or horizontal acquisition from other bacteria through transferring plasmids carrying genetic materials encoding for antibiotic resistance. Acquired resistance typically occurs in response to selective antibiotic pressures. These mechanisms are stable and can be transferred vertically (e.g., upon bacterial replication) or horizontally (e.g., resistance genes by plasmids). Adaptive resistance is induced in the presence of specific antibiotics and other environmental stresses and is transient, given that susceptibility is restored upon removal of the stimuli. This type of resistance mainly relies on induced alterations in gene expression, resulting in increased protein production or alterations in antibiotic targets.

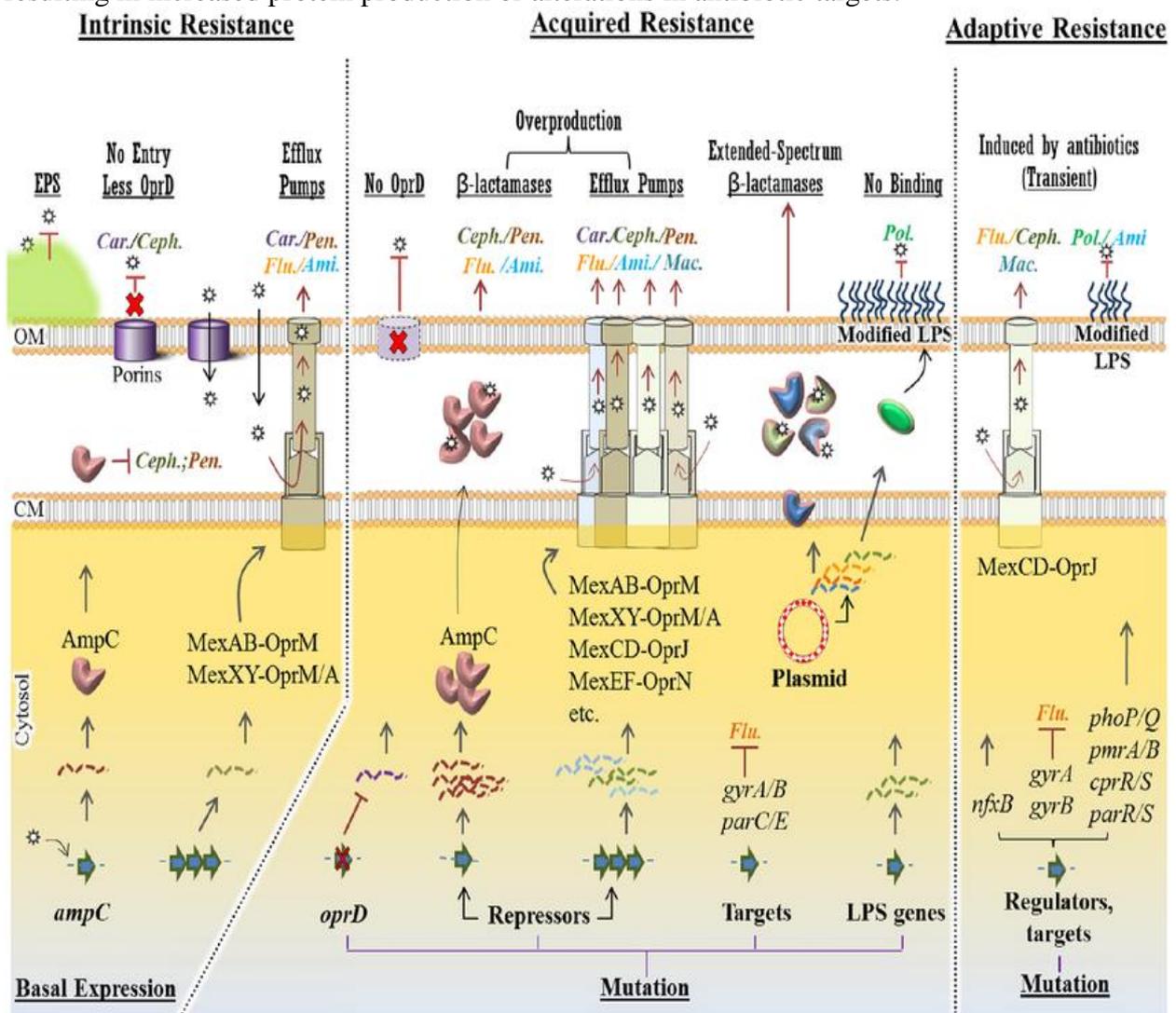


Fig-2: Intrinsic, acquired, and adaptive mechanisms confer antibiotic resistance in *P. aeruginosa*.

Efflux Pump Systems-

P. aeruginosa has a robust efflux pump system. The primary purpose of these pumps is to expel toxic environmental compounds or metabolites from the cytoplasm that might

otherwise disorganize the cytoplasmic membrane. Substrates of these pump systems include many clinically relevant antibiotics such as β -lactams, fluoroquinolones, amino glycosides, macrolides, tetracyclines, sulfonamides, and chloramphenicol, among other compounds. Multidrug-resistant isolates are very likely to have efflux pump system up-regulation. *P. aeruginosa* has several multidrug efflux pump systems. Of the five protein efflux system families described to date, most of those expressed in *P. aeruginosa* are members of the same (i.e., resistance-nodulation-cell division) superfamily. These efflux systems usually have three components: a cytoplasmic membrane pump, a cytoplasmic membrane “exit” porin, and a linker protein. The best-described pump system in *P. aeruginosa* is MexAB-OprM, which is expressed in all isolates to varying degrees. Wild-type strains tend to have relatively low expression, but mutations in the *mexR* repressor gene can result in pump overexpression. Overexpression of MexAB-OprM results in high-level resistance (e.g., increases in MIC by 8-fold) to a range of antibiotics. Genetic deletion of this pump restores susceptibility to many agents that are not considered clinically active against *P. aeruginosa* such as amoxicillin, cefuroxime, and tetracycline. The antipseudomonal agents perhaps most affected by efflux pumps are β -lactams and aminoglycosides, with fluoroquinolones possibly less affected.

MATERIAL AND METHODS:

Fifty consecutive isolates of *Pseudomonas aeruginosa* obtained from various clinical specimens were processed at Central Microbiology Laboratory of Dhiraj Hospital between the periods of March 2024 to May 2024.

Specimens were cultured on Blood agar, MacConkey agar and Nutrient agar and incubated at 37°C for 24 hours. Colonies were identified by its characteristics and biochemical reactions. The samples were further processed for antibiotic sensitivity testing using VITEK 2 automated system and interpreted using CLSI guidelines².

RESULTS:

A total of 50 isolates of *Pseudomonas aeruginosa* was found from clinical samples with 56% samples from pus, 24% from sputum, 5% from endotracheal tube secretion, 2% from CSF, and 8% from others. Most of samples were of male 72%.

The result showed highest sensitivity to drug amikacin (54%), followed by Piperacillin/Tazobactam (50%), Ciprofloxacin (46%), Cefepime (42%), Cefosalbactam (40%), Levofloxacin, Doripenem, Meropenem, Gentamicin (38%), Ceftazidime (36%), Imipenem (34%), Ticarcillin/Clavulanic Acid (32%). The least sensitive drug was Tigecycline (8%).

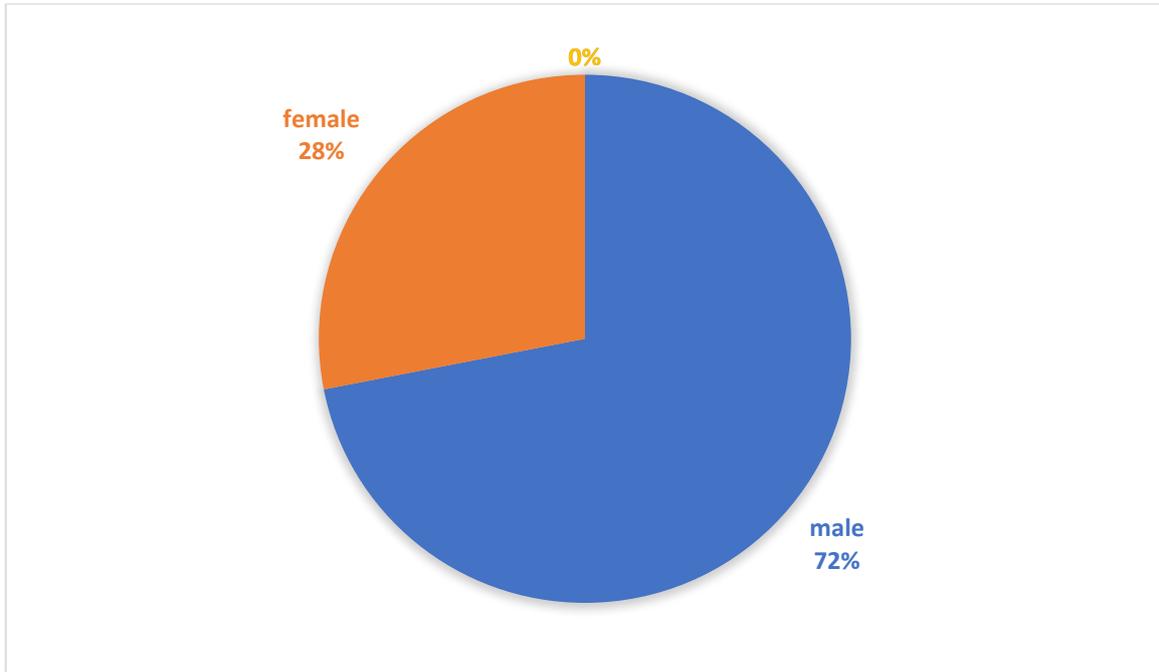


Fig-3: male – female ratio

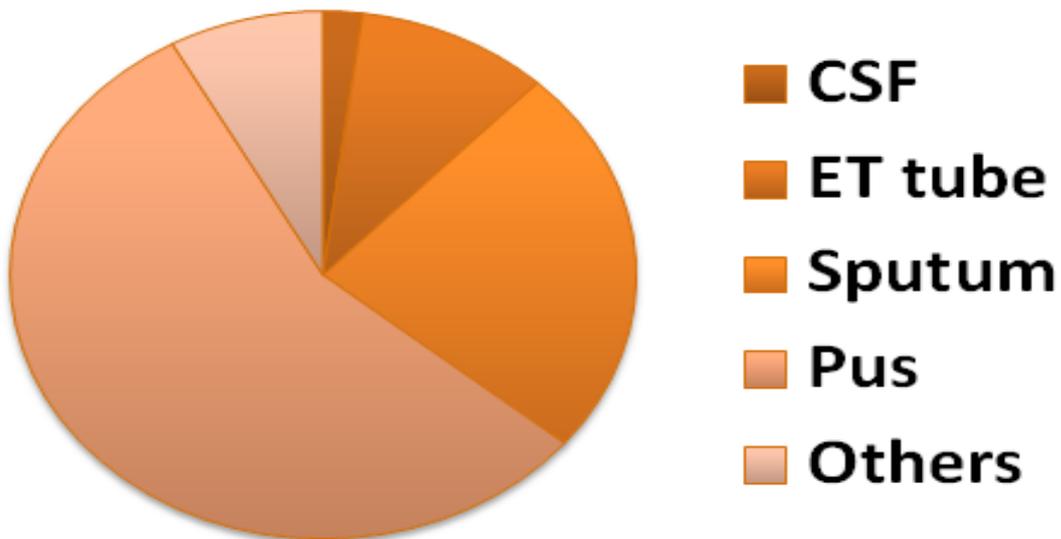


Fig-4: various samples

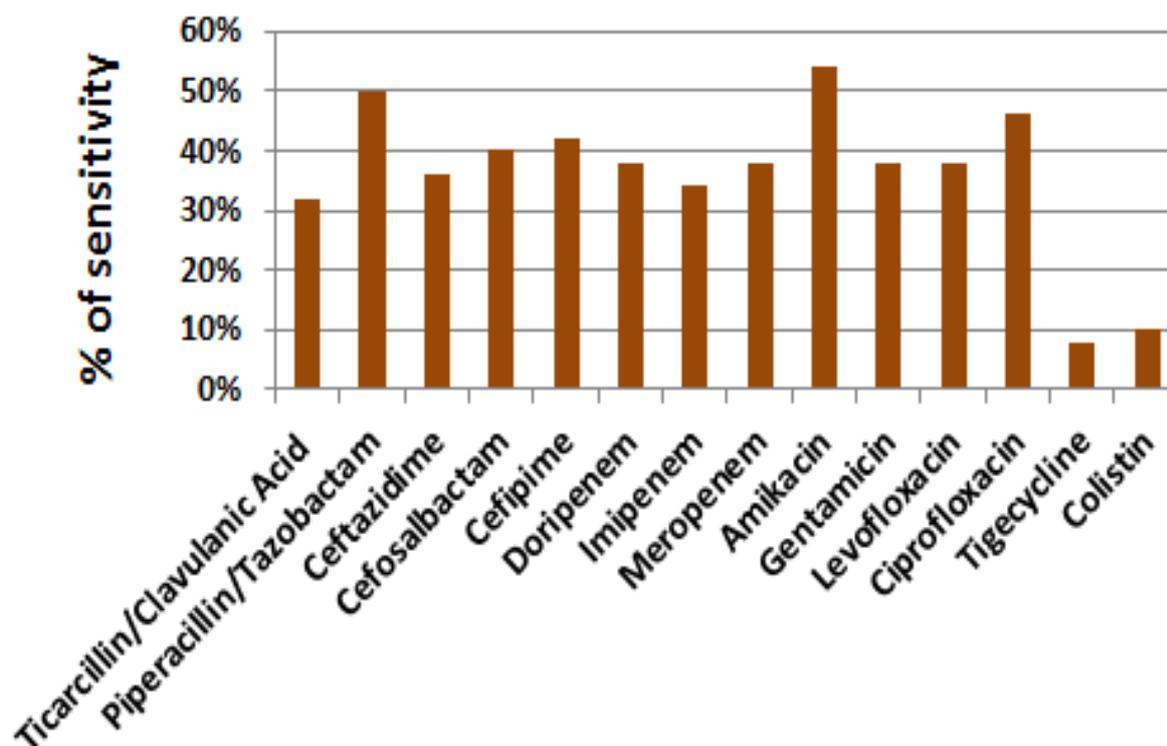


Fig -5: showing antibiotic sensitivity.

DISCUSSION:

P. aeruginosa is one of the most isolated health-care associated pathogen worldwide. A total of 50 isolates of *Pseudomonas aeruginosa* was found from clinical samples with 56% samples from pus, 24% from sputum, 5% from endotracheal tube secretion, 2% from CSF, and 8% from others. Most of samples were of male 72%.

Most of samples were from male patients (63%), which is same as reported by Sharma et.al, Viren et. al, Ali Hussein et al, Shampa et al, and Rakesh et al. Previous study show higher isolation of *P.aeruginosa* from intensive care unit⁹.

The occurrence of *P.aeruginosa* is found to be higher in males, inpatients in age group >60, 41 years and in surgery department, which is same as reported by Viren et. al¹⁰, Ali Hussein et al¹², Shampa et al¹³ and Rakesh et al.¹⁰.

P. aeruginosa rapidly acquires high-level resistance to these drugs which may have impacts on both clinical and economic outcomes. Sensitivity to Meropenem, Cefepime, and Piperacillin-Tazobactam was better (67%, 62%, and 59%). Amikacin, Meropenem, Cefepime, and Piperacillin-Tazobactam could be used to treat *Pseudomonas* infections.

CONCLUSION:

Healthcare settings largely contribute as reservoirs of pathogenic strains of *Pseudomonas aeruginosa*. They are known to utilize their highest levels of intrinsic and acquired resistance mechanisms against most antibiotics. Multidrug resistance is also increasing³.

REFERENCES:

1. Rivani, Erizka & Widodo, Agung. (2019). Susceptibility Pattern of *Pseudomonasaeruginosa* Isolated From Blood, Urine, Pus, and Sputum Specimens in RSUD Dr. Soetomo. 10.13140/RG.2.2.24184.60164.
2. CLSI (Clinical and Laboratory Standard Institute) 2022, Performance Standard for Antimicrobial Susceptibility Testing, 32nd edition, M100.

3. Pachori P, Gothalwal R, Gandhi P. Emergence of antibiotic resistance *Pseudomonas aeruginosa* in intensive care unit; a critical review. *Genes Dis.* 2019;6(2):109-119. Published 2019 Apr 17. doi:10.1016/j.gendis.2019.04.001
4. Rossi E, et al. *Pseudomonas aeruginosa* adaptation and evolution in patients with cystic fibrosis. *Nat. Rev. Microbiol.* 2021;19:331–342. doi: 10.1038/s41579-020-00477-5. [PubMed] [CrossRef] [Google Scholar]
5. Jurado-Martin, I., Sainz-Mejias, M. & McClean, S. *Pseudomonas aeruginosa*: an audacious pathogen with an adaptable arsenal of virulence factors. *Int. J. Mol. Sci.* **22**, 1–35 (2021). [PMC free article] [PubMed]
6. Cendra MDM, Torrents E. *Pseudomonas aeruginosa* biofilms and their partners in crime. *Biotechnol. Adv.* 2021;49:107734. doi: 10.1016/j.biotechadv.2021.107734. [PubMed] [CrossRef] [Google Scholar]
7. Sinha, M. et al. *Pseudomonas aeruginosa* theft biofilm require host lipids of cutaneous wound. *Ann. Surg.* **5252**, 1–23 (2021). [PMC free article] [PubMed]
8. Tang P, et al. BNT162b2 and mRNA-1273 COVID-19 vaccine effectiveness against the SARS-CoV-2 Delta variant in Qatar. *Nat. Med.* 2021;27:2136–2143. doi: 10.1038/s41591-021-01583-
9. Jaswinder Sharma¹, Prevalence and Antimicrobial Susceptibility Pattern of *Pseudomonas Aeruginosa* Isolated from Pus Samples in a Tertiary Care Hospital, Bathinda, ISSN (Online): 2393-915X; (Print): 2454-7379 | ICV (2015): 77.83.
10. Rakesh R.M, Ninama G. L., Mistry K., Parmar R, Patel K, Vegad M M. Antibiotic resistance pattern in *Pseudomonas aeruginosa* species isolated at a tertiary care Hospital, Ahmadabad. *National Journal Of Medical Research.* 2012;2:156-58.