

<https://doi.org/10.33472/AFJBS.6.6.2024.880-890>



African Journal of Biological Sciences

Journal homepage: <http://www.afjbs.com>



Research Paper

Open Access

The GC-MS and antioxidant activity analysis using organic solvent extracts of *Leucas aspera*

KAMALAMBIGESWARI R^{1*}, KAMALESH A², SHIBIN B³, PUTTUR ANUSHA⁴, JOSEPH M⁵

¹Department of Industrial Biotechnology, BIHER

Article Info

Volume 6, Issue 6, May 2024

Received: 28 March 2024

Accepted: 30 April 2024

doi: 10.33472/AFJBS.6.6.2024.880-890

ABSTRACT

Drugs can be generated from many plants. *Leucas aspera* is known as “Thumbai” were taken and 5g of Leaf was treated with 5ml of ethanol and Acetone respectively, and the flowers of *Leucas aspera* were also taken. 1g of flower is treated with the 10ml of Ethanol and Acetone each to get the extract. The Antibacterial activity of the extract was found to be in 100 µg/ml of the flower extract of acetone and the maximum antioxidant activity was also found at 120 µg/ml of flower extract of acetone. The GC-MS analysis were done for the identification of bioactive compounds.

KeyWords : Flower extract, Anti-bacterial activity, Anti-oxidant activity, GC-MS analysis

INTRODUCTION

Nature is a richest source of medicinal plants diversity. Many novel drugs have been isolated from it and most of these novel drugs are based on their use in traditional medicine. In ancient days, the treatment of diseases is done by using medicinal plants, herbs due to their potential to cure diseases^{1,2}. In developing countries the interest of medicinal plants is increasing as the herbal medicine have been reported as safe and also the adverse effect is low particularly when compared with synthetic drugs^{3,4}. Based on folklore medicine, the plants from the genus *Leucas* have different kinds of therapeutic activities. *Leucas aspera* belongs to the family Lamiaceae commonly known as ‘Thumbai’ (in Tamil) ‘Thumbe’, ‘White dead nettle’ (English), ‘Dronapushpi’, ‘Chitrapatrika’ (Sanskrit). The whole plant is traditionally important because it has many therapeutic values^{5,6}.

MATERIALS AND METHODS

Preparation of the Extract

5g of leaf was taken from the plant and 5ml of ethanol was added to it. Another 5g of leaf treated with 5ml of acetone⁷. Both was then grinded well then by using pestle and mortar and the filtrate was extracted as previous procedure. 1g of flower taken from the plant and treated with 10ml of Ethanol and Acetone respectively and the filtrate was obtained⁸.

ANTIBACTERIAL ACTIVITY

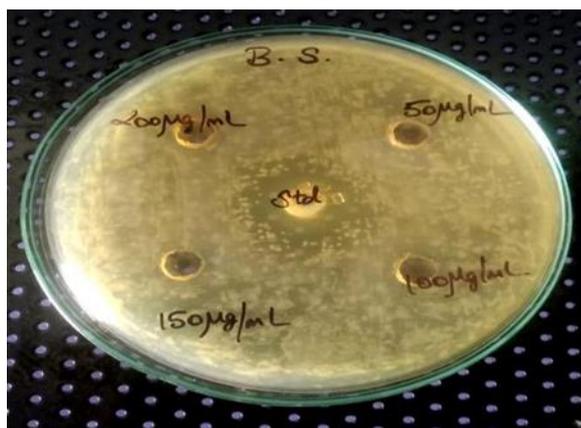
Nutrient agar was prepared and poured in the sterile Petri dishes and allowed to solidify. The overnight-grown bacterial colony was used for the preparation of the bacterial suspension. A single colony was taken and dissolved in normal saline solution *and* was swabbed on it. Wells were prepared by using a gel puncture of 10 mm diameter on all of the prepared agar plates. Then, the test sample was added over the agar wells. Streptomycin (25µL) was used as positive control (Standard). The plates were then incubated at 37°C for 24hours⁹. After incubation the inhibition diameter was measured and units are mm.

ANTIOXIDANT ASSAY**DPPH ASSAY**

The Radical Scavenging Activity of test sample was determined by using DPPH assay according to Chang et al., (2008)¹⁰ with small modification. The decrease of the absorption at 517nm of the DPPH solution after the addition of the antioxidant was measured in a cuvette containing 2960µl of 0.1mM ethanolic DPPH solution mixed with 20 to 120 µg/ml of test sample and vortexed thoroughly¹¹. The setup was left at dark in room temperature and the absorption was monitored after 30minutes. The ability of the plant extract to scavenge DPPH radical was calculated¹² by the following equation:

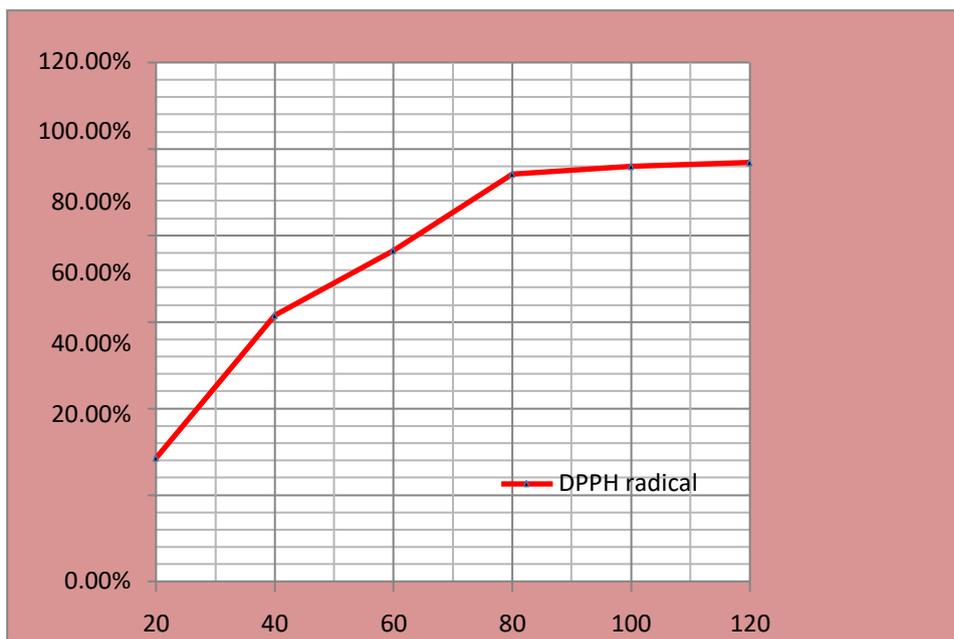
$$\% \text{ of DPPH Radical Scavenging Activity} = \frac{\text{Abs. control} - \text{Abs. sample} * 100}{\text{Abs. control}}$$

Abs. control is the absorbance of DPPH radical + methanol; Abs. sample is the absorbance of DPPH radical + test sample

RESULTS AND DISCUSSION**ANTIBACTERIAL ACTIVITY**

The agar well diffusion method is the most commonly used method to examine the antimicrobial activity. The anti bacterial activity in plates was observed which was swabbed with *Bacillus sp.*, showed 0.3 mm zone of inhibition to 100µg/ml when used streptomycin as the standard¹³. The zone of inhibition was measured for the standard which is about 0.5mm.

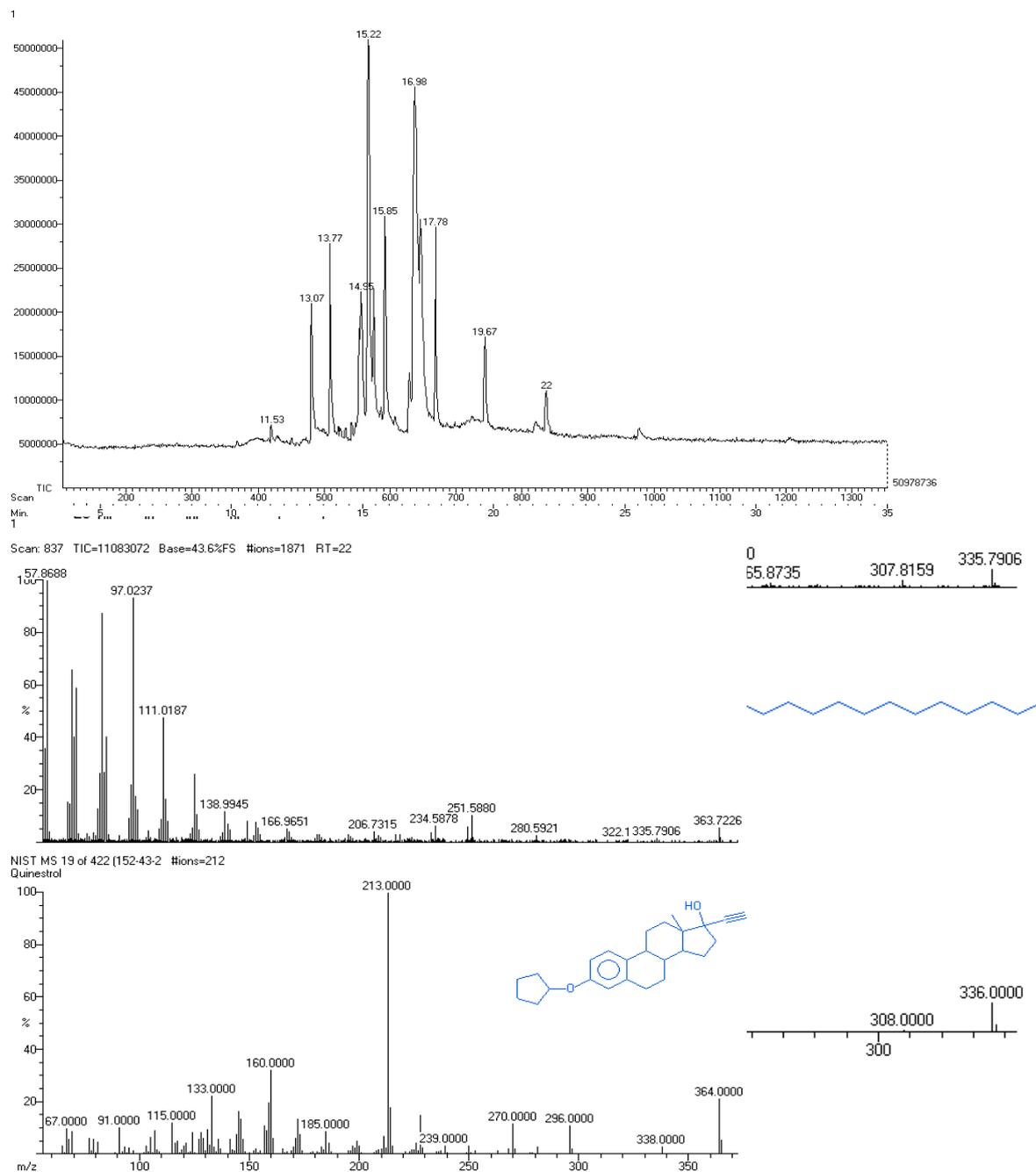
ANTIOXIDANT ACTIVITY



The 2,2-Diphenyl-1-picrylhydrazyl (DPPH) is a popular, quick, easy, and affordable approach for the measurement of antioxidant properties that includes the use of the free radicals used for assessing the potential of substances to serve as hydrogen providers or free-radical scavengers (FRS). The decrease of the absorption at 517nm of the DPPH solution after the addition of the antioxidant was measured in a cuvette containing 2960µl of 0.1mM ethanolic DPPH solution mixed with 20 to 120 µg/ml of test¹⁴. Anti-scavenging activity was found to be increasing and was found to be 120 µg/ml. Damage caused by ROS may be reduced by using antioxidants, which are chemicals that can stop other molecules from oxidizing. Due to their potential to contribute electrons that may neutralize radical production, antioxidants are useful in lowering and preventing further damage through free-radical responses

GC MS REPORT

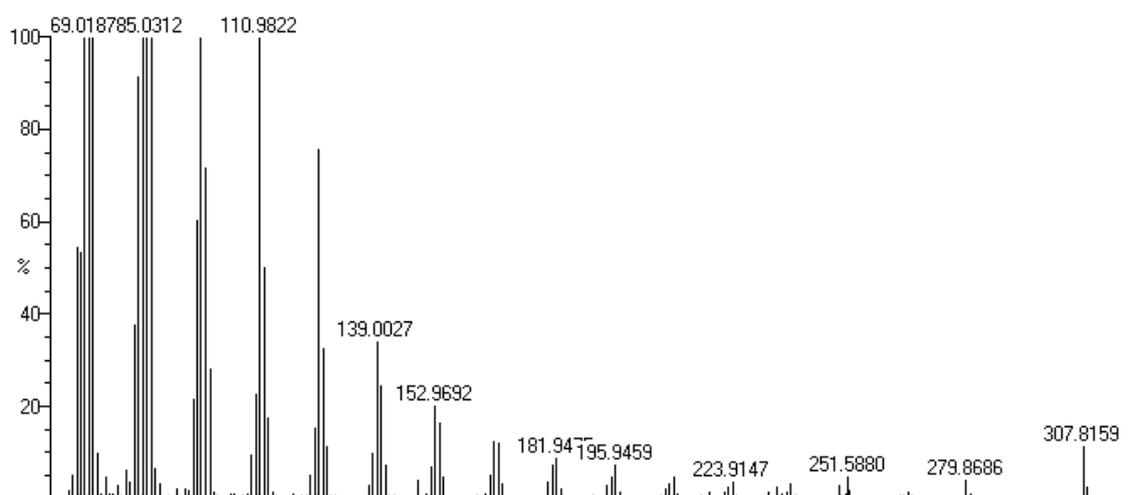
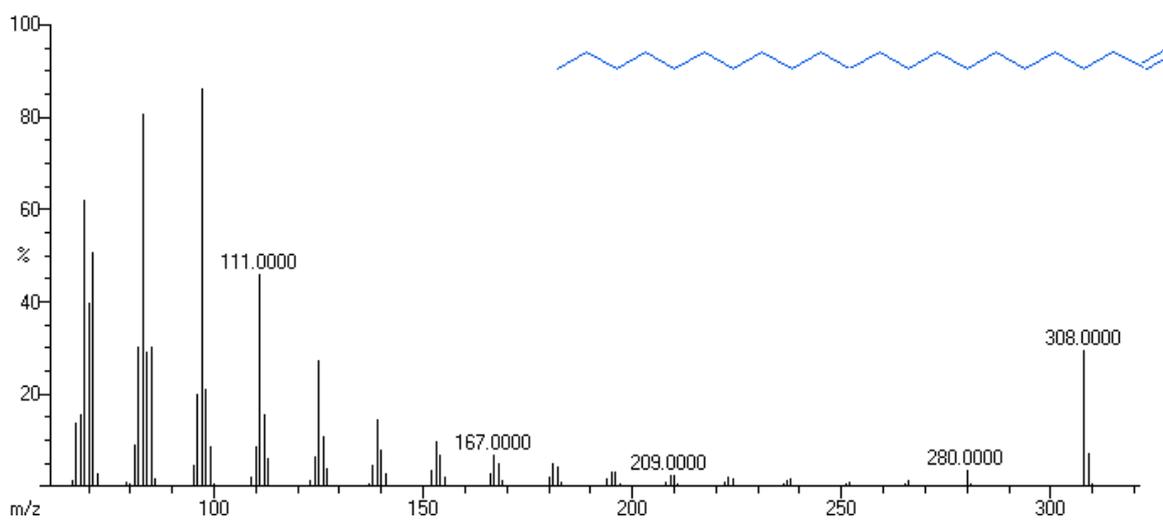
A good chromatogram of each standard sample. A mass spectrum of each sample is obtained. Chromatographs and mass spectra of all unknown mixtures has been depicted. The retention time and compound category has been identified ^{15,16}.



KAMALAMBIGESWARI R /Afr.J.Bio.Sc. 6(6) (2024)

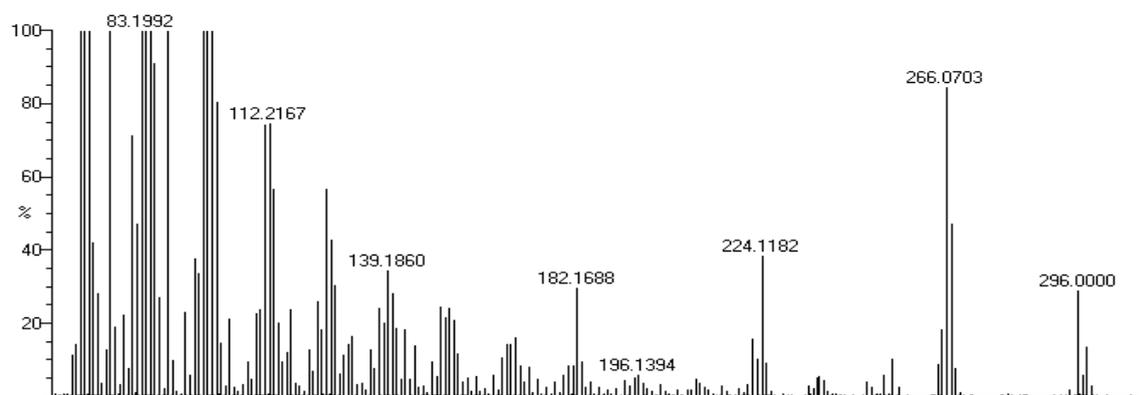
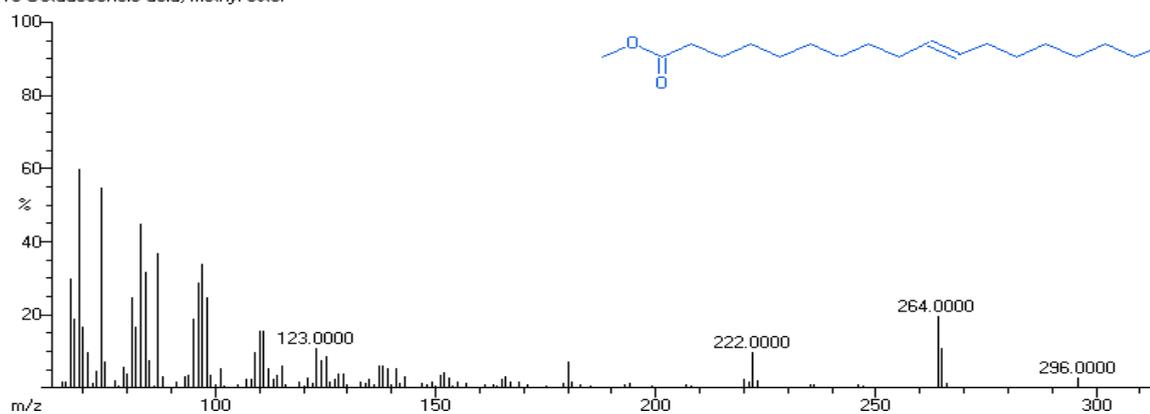
1

Scan: 669 TIC=29631120 Base=100%FS #ions=1940 RT=17.78

NIST MS 1 of 40 (1599-67-3) #ions=136
1-Docosene

1

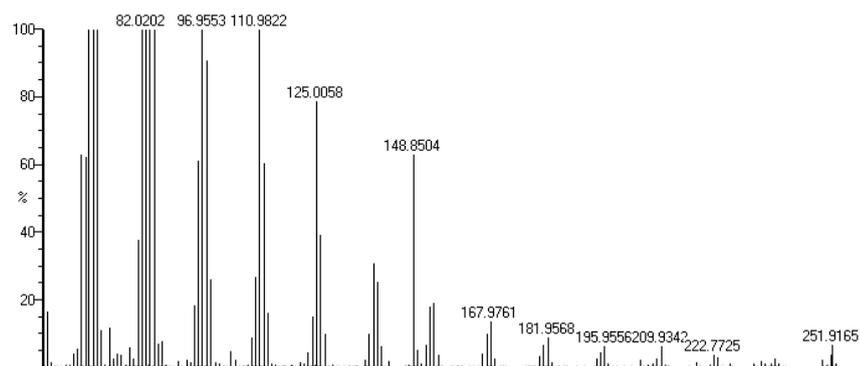
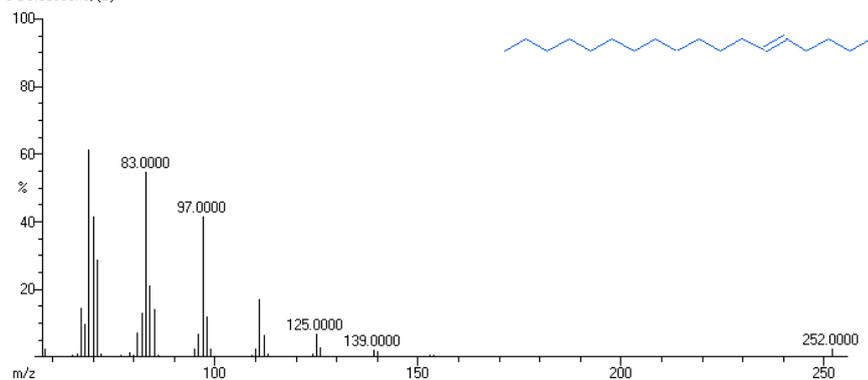
Scan: 637 TIC=45612416 Base=100%FS #ions=1985 RT=16.98

NIST MS 1 of 40 (13481-95-3 #ions=249
10-Octadecenoic acid, methyl ester

KAMALAMBIGESWARI R /Afr.J.Bio.Sc. 6(6) (2024)

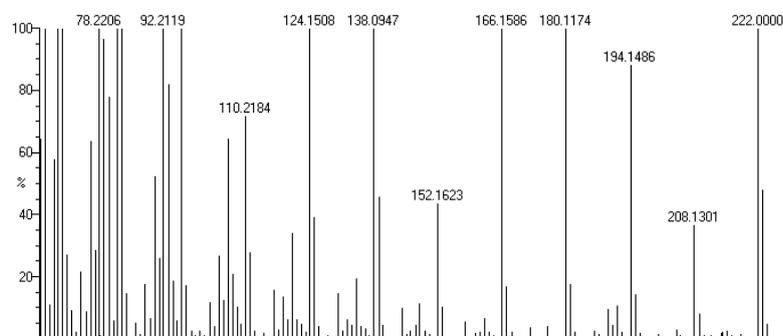
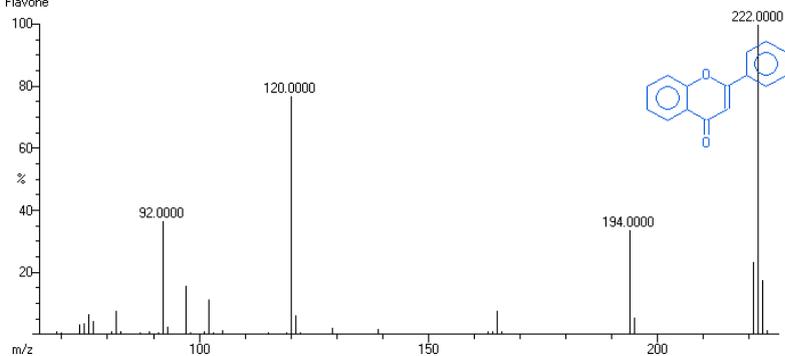
1

Scan: 592 TIC=30931280 Base=100%FS #Ions=1884 RT=15.85

NIST MS 561 of 702 (7206-21 #Ions=104
5-Octadecene, [E]-)

1

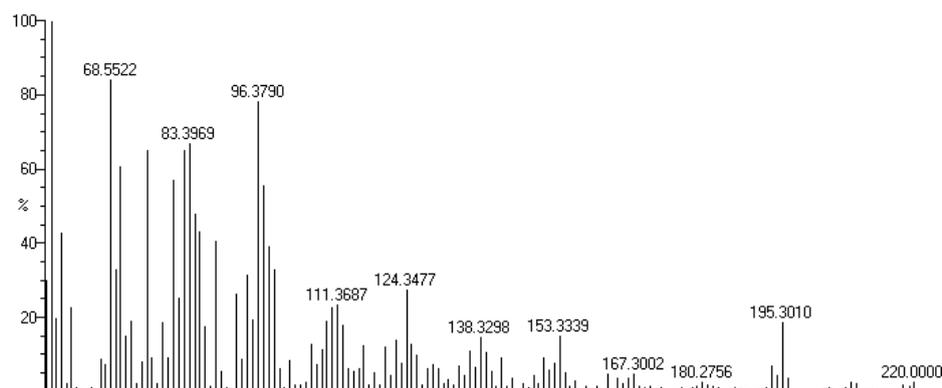
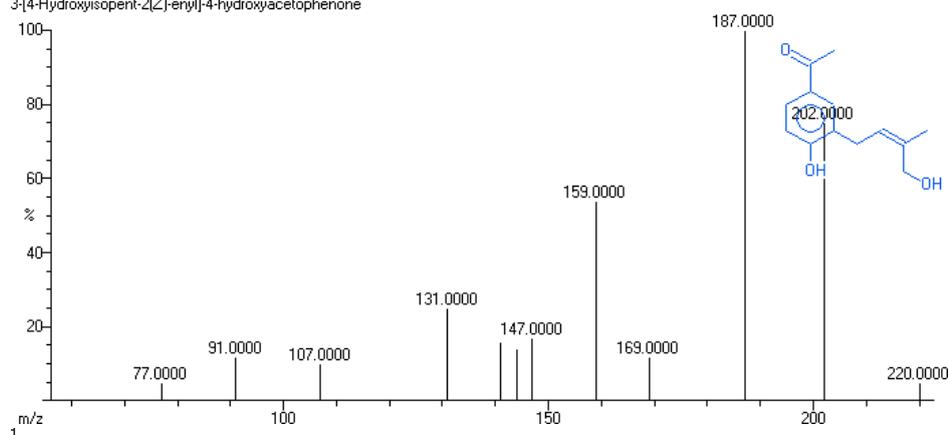
Scan: 567 TIC=50978736 Base=100%FS #Ions=1951 RT=15.22

NIST CAS# 525-82-6 #Ions=78
Flavone

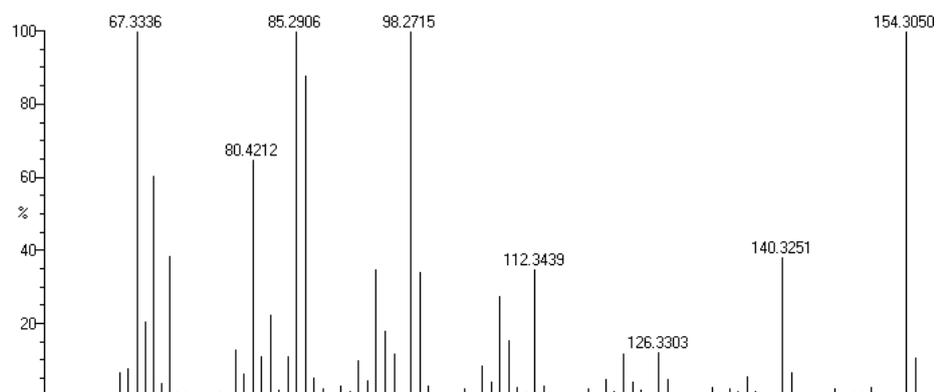
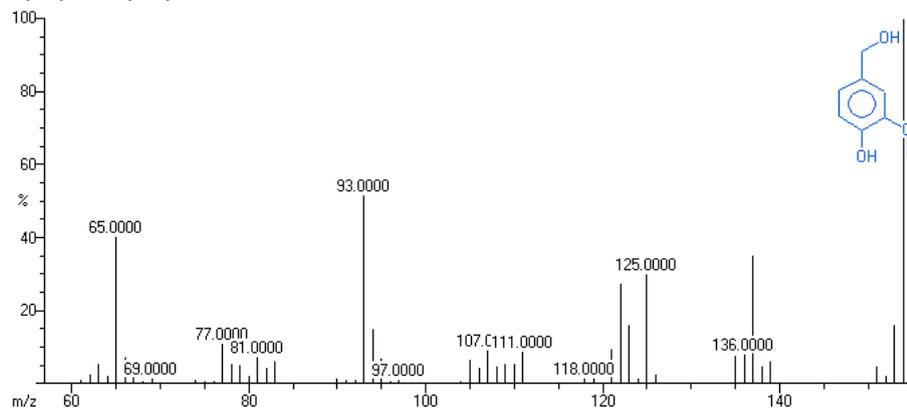
KAMALAMBIGESWARI R /Afr.J.Bio.Sc. 6(6) (2024)

1

Scan: 556 TIC=22345936 Base=80.2%FS #ions=1993 RT=14.95

NIST MS 2 of 811 (24672-83- #ions=13
3-[4-Hydroxyisopent-2(Z)-enyl]-4-hydroxyacetophenone

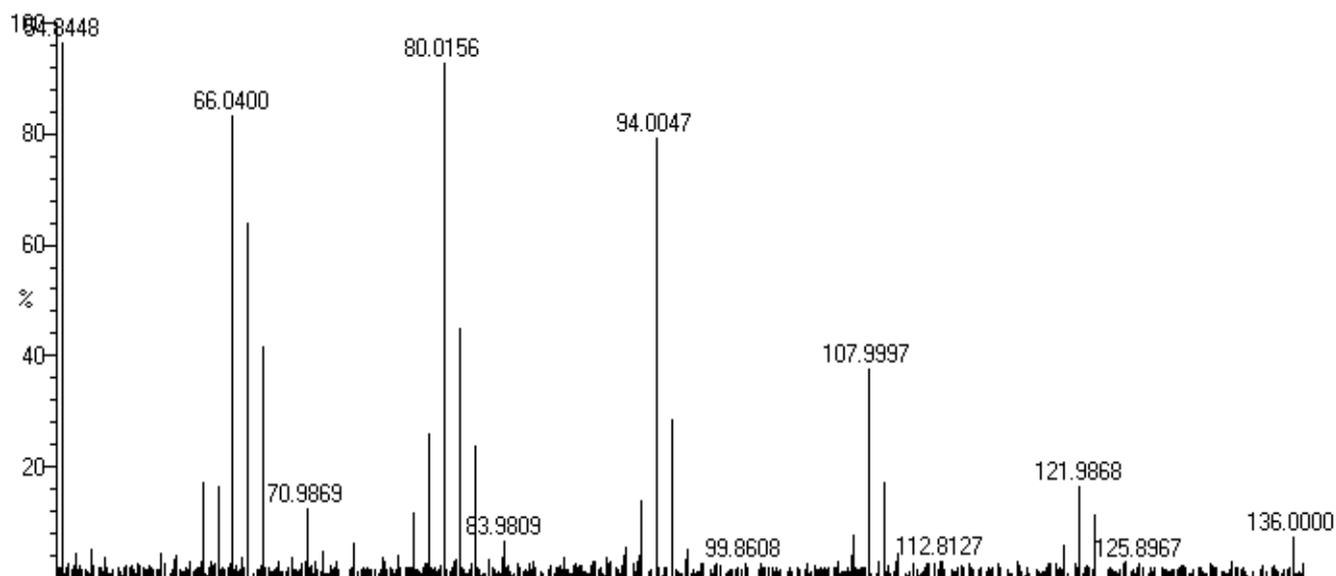
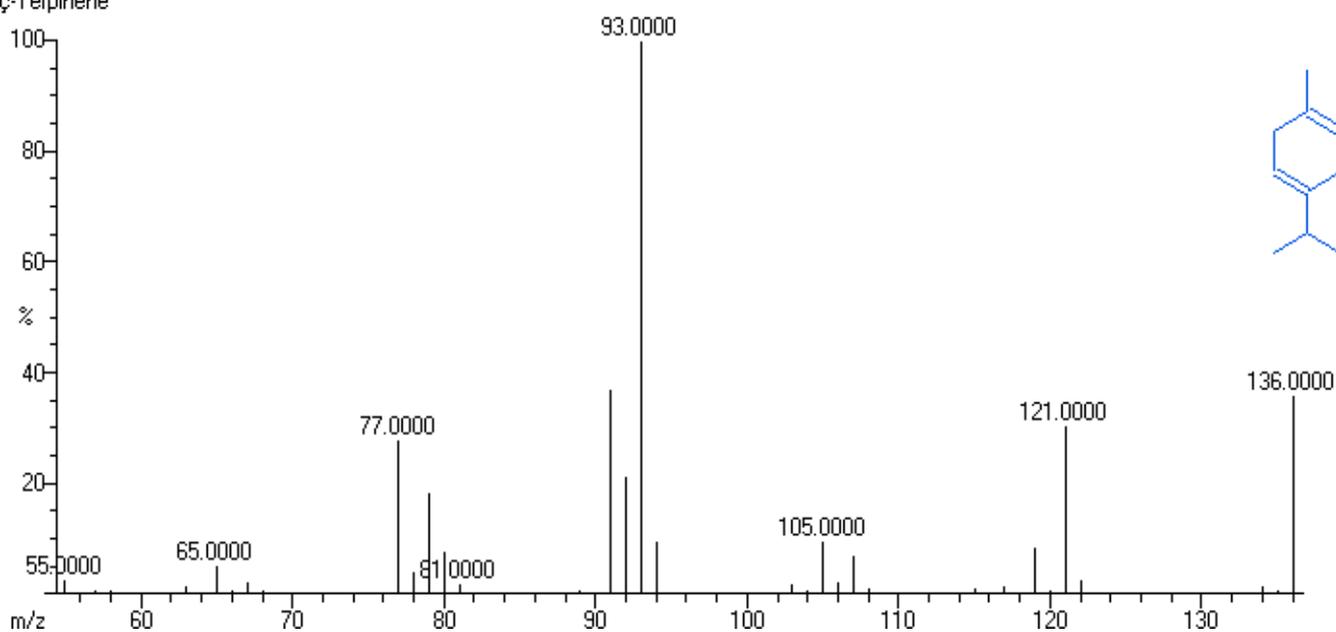
Scan: 481 TIC=21002352 Base=100%FS #ions=1801 RT=13.07

NIST MS 15 of 832 (498-00-0 #ions=92
4-Hydroxy-3-methoxybenzyl alcohol

KAMALAMBIGESWARI R /Afr.J.Bio.Sc. 6(6) (2024)

1

Scan: 420 TIC=7148432 Base=17.2%FS #ions=1683 RT=11.53

NIST MS 15 of 549 (99-85-4) #ions=71
 ζ -Terpinene

CONCLUSION

Leucas aspera is a medicinal plant which is traditionally important. *Leucas* genus has many species. The bioactive compounds such as diterpenes, lignans, flavonoids, squalene of this plant has many therapeutic values. *Leucas aspera* extracts have antimicrobial, antioxidant, anticancer, antidiabetic activity. The phytochemical compounds present in *Leucas aspera* can be harnessed for human therapeutic purpose due to its pharmacological activities.

REFERENCE

1. Kelly K. *History of medicine*. New York: Facts on file; 2009. pp. 29–50.
2. Tucakov J. *Healing with plants – phytotherapy*. Beograd: Culture; 1971. pp. 180–90.
3. Sasaki RT, Flório FM, Basting RT. Effect of 10% sodium ascorbate and 10% α -tocopherol in different formulations on the shear bond strength of enamel and dentin submitted to a home-use bleaching treatment. *Oper Dent* 2009; 34:746-752.
4. Mukherjee P. W. *Quality Control of Herbal Drugs: An Approach to Evaluation of Botanicals*. 2002, New Delhi, India: Business Horizons Publishers
5. Baldwin C. A., Anderson L. A., Phillipson J. D. What pharmacists should know about *Ginseng*. *Pharm. J.* 1986 237 583–586
6. Bandaranayake W. M. “Quality control, screening, toxicity, and regulation of herbal drugs,” in *Modern Phytomedicine. Turning Medicinal Plants into Drugs* eds Ahmad I., Aqil F., Owais M. (Weinheim:Wiley-VCH GmbH & Co. KGaA;) 2006, 25–57
7. Jeyanthi Rebecca L, Kamalambigeswari.R. Extraction of Antioxidant Lutein from Various Flowers. *International journal of Pharmaceutical Sciences review and research*. 2016, 39. 122-124.
8. Sandoval M, Charbonnet RM, Okuhama NN, Roberts J, Krenova Z, Trentacosti AM, et al.. Cat's claw inhibits TNF alpha production and scavenges free radicals: role in cytoprotection. *Free Radic Biol Med* 2000;29:71-78.
9. Kum KY, Lim KR, Lee CY, Park KH, Safavi KE, Fouad AF, et al.. Effects of removing residual peroxide and other oxygen radicals on the shear bond strength and failure modes at resin-tooth interface after tooth bleaching. *Am J Dent* 2004;17:267-270.
10. Kamalambigeswari R, Sharmila S, Kowsalya E, Janani S, Deva V. Extraction of Omega-3 Fatty Acid-methyl stearate from Soil Fungi (*Fusarium* sp.). *Research Journal of Pharmacy and Technology* 2019; 12 (9): 4295-4298
11. Chang HC, Dimlich DN, Yokokura T, Mukherjee A.). Modeling spinal muscular atrophy in *Drosophila*. *PLoS ONE* 2008; 3(9): e3209.
12. Kamalambigeswari R, Alagar S, Sivvaswamy N .Isolation, identification, screening and optimisation of pectinase producing soil fungi (*Aspergillus niger*). *IJRPS* 2018; 9 (3): 762-768
13. Tay LY, Kose C, Loguercio AD, Reis A. Assessing the effect of a desensitizing agent used before in-office tooth bleaching. *J Am Dent Assoc* 2009;140:1245-1251.
14. Dreifuss AA, Bastos-Pereira AL, Avila TV, Soley B da S, Rivero AJ, Aguilar JL, et al.. Antitumoral and antioxidant effects of a hydroalcoholic extract of cat's claw (*Uncaria tomentosa*) in an in vivo carcinosarcoma model. *J Ethnopharmacol* 2010;130:127-133.

15. McLafferty, F. *Interpretation of Mass Spectra*; 3rd ed.; University Science Books: Mill Valley, CA, 1980.
16. Silverstein, R. M.; Bassler, G. C.; Morrill, T. C. *Spectrometric Interpretation of Organic Compounds*; 4th ed.; Wiley: New York, NY, 1981.