https://doi.org/ 10.33472/AFJBS.6.10.2024.738-744



RESEARCH ARTICLE

Optimization and Evaluation of 5-Fluorouracil-Entrapped edge-activating Transfersome as a New Prospective Thermo Sensitizer at Different Fascinating Importance Illustration Agent Study for: Ex-vivo study Nidhi Jain^{1*}, Anup Kumar Chakraborty²

¹Research Scholars, Department of Pharmacy, Oriental University, Indore, India.
 ²Professor, Department of Pharmacy, Oriental University, Indore, India.
 *Corresponding author: Nidhi Jain, Research Scholars, Oriental University, Indore, India.

Article History Volume 6,Issue 10, 2024 Received:17 Apr 2024 Accepted : 05 May 2024 doi: 10.33472/AFJBS.6.10.2024.738-744

ABSTRACT

The Topical-delivery possessions of the new prospective nano-carriers drug liberation systems, which were accustomed increase the time retention on the anticancer agent in the topically body. In this drug distribution systems occurred extensively exploited to distribute anticancer drugs. The 5-fluorouracil entrapped soya-lacithin30% transferosomal preparation in this broad-fields have the great beneficial effect. Now, we obtained that while the rational quantity of 5-fluorouracil and soya-lacithin 30% was 1:1. Results showed that the average particle size, polydispersity index, and zeta potential were 35.58 ± 0.56 , 0.285 ± 0.125 , and 14.50 ± 0.8 respectively, for 5FU loaded transfersomes. And image shown that TEM, FTIR Study, X-ray diffraction study mention below. The highest encapsulation efficiency achieved was 85.05 ± 0.58 for P-16 formulation.

Keywords: Anticancer, 5-FU, transferosome, QbD, Box behenkan design, soya Lecithin (30%).International Journal of Drug Delivery Technology (2024).

INTRODUCTION

Topical drug distribution system (transdermal drug distribution system) are numerous implied benefit in conventional drug delivery system like restraint in different type of pre systemic metabolism^{1,2} Now, topical treatment of 5-FU has established carefulness since of its antiinflammatory, antioxidant, and immune modulatory possessions which may present a therapeutic threshold for treatment of UV-ray generated to skin cancer. It effectively inhibits both photo carcinogenesis cell and skin tumor generating cells. It also acts high efficacy in inhibition of skin cancer by thymidylate synthase (TS) inhibitor³. Bracke of the enzyme cell membrane and blocked of the action of the action DNA replication. In the face of the effectively use of 5-FU as an efficient and protected compound for chemotherapy. Distribution of medications to the skin is an efficient and targeted drug delivery system for local topical disorders. Topical hydrogel preparations deliver a convenient distribution system for drugs, since they are non -greasy, hydrophilic nature and can be readily separated from the skin. Dermal delivery system is alluring route of drug administration for topical treatment for both local and systemic effect show^{4, 5}.

Transfersomes vesicles are shown as extra elasticity came pair to the typical liposomes and therefore well suitable permeation for penetration of the topical skin. The transfersomeshave permeated the epidermis penetration power and diffusion are problematic by holding themselves alongside an intra-cellular stopping phospholipid of the st. corneum^{6, 7}. It are very specific through transfersomes, since the size of vesicles deformability which permeation the entrance as a result of the mechanical pressure of encircling, in a self- regulating method. Provide elasticity and good resistance of surrounding membrane is controlled by appropriate quantity added edge activator agent (surfactants) result in mechanisms in the appropriate quantity or rational quantity using lipids molecules^{8, 9}. Cancer' is thus a generic hereditary termed purpose used to illustrate a collection group of around a many diseases that develop when malignant forms of abdominal and abnormal cell growth multiplication in one or more body organs. Cancer arrived from later a series of hereditary mutations eliminate the normal confidential cell growth. These cancer cells survive and growth to divide and spread to produce tumours^{10, 11}.

Nidhi Jain / Afr.J.Bio.Sc. 6(10) (2024)

The continual exposure UV may origin an irregular skin discoloration, wrinkles, decrease of skin flexibility and elasticity, skin aged and a discoloration of skin obstruction occupations¹. These fluctuations in the skin are on the way to as photo aging. The photo aging and critical exposure to UV radiation causes in the growth of, melanoma, SCC and BCC. Continual exposure UV radiations origins stimulation of dangerous inflammation, oxidative anxiety, DNA nucleotide damage (development and formation of cyclobutane pyrimidine dimmers), gene mutilation and immune suppression. In additionally to this, UV radiation cause alteration and mutations of p53 malignant tumor suppressor genetic material (gene)^{12, 13}.Also, UV radiation can create up regulation of gene appearance over and done with intracellular wave introduction route there by introducing to growth and development of skin cancer at tumor promotion stage ^{14,}. They also produce the creation of reactive oxygen species (ROS), which produces oxidative stress in skin cells¹⁵. ROS (reactive oxygen species) have been exposed to stimulate transcription activity formulated for instance activate Protein -1 (AP-1) and nuclear factor kB (NF-kB), they participate to cell proliferation and then cell death^{16, 17}.

MATERIAL AND METHODS

Materials

Determined drug 5-FU were a bought from S.K. Traders (Indore). Tween 80, Rhodamine and Phospholipid were bought from, New Modern Chemical Corporation Mumbai (HPLC grade), soya lecithin 30% was purchased at HiMedia Laboratory, carbopol 940 and Sopn 80 were purchased from LOBA Chemie Pvt. Ltd, Mumbai, India. 5-FU marketed product name in flonida cream 1% was procured from Rohan Chemist Indore. All additional chemicals used in the study were of analytical grade. Purified water from ultra-pure water system (Synergy UV water purifier system, India) was used throughout the study.

Preparation of Transferosome

Transfersome was doing processed through the performance described in Patel et al., (2009) with slight modification. The transfersome had formulated through rotating evaporating sonication (phospholipid film hydration) method to be used. The amalgamation of assembling are anticancer active consequents drug 5-FU, soya lecithin 30% (phospholipid) and non-ionic surfactant as a used in edge activator agent just like tween-80/span-80 were taken round bottom flask (RBF) and expended in uniformly ethanol^{19, 20}.

When the heated approximate temperature 55°C so organic solvent removed by evaporation method, while a skinny lipid film was obtained inner portion the round bottom (RBF) flask. The skinny film layer had deposited and spread in a short time period (12hrs) to be total evaporation and from a skinny film layer. Then by phosphate buffer (pH 7.4) used the hydrated skinny film layer upon still the sonicated approximate 30 minute (room temperature) in a probe sonication to reduction the vesicle size to the from an uniform size vesicles. Then, their found in suspension like transfersome again hydrated in phosphate buffer solution in low temperature 2-8oC (at 1hr). Preserve in transferosome in a well-closed and air tight container. Same as, empty transfersomes vesicles (without 5-FU) were formulated.

Optimization of Drug Loaded Transfersomes

Box-behnken design–response surface methodology (BBD– RSM) be situated operated to computerized software arrangement different examine that were factor affected stimulus of the triple precarious preparation variables just like vesicle size distribution, percentage drug loading (w/w) and EE % (w/w) that were determined arranged transfersomes. Considered for all element, the varieties of investigates were a selection of the origin of the consequences of primary investigates & the possibility of prepared transfersomes on the great standards value determined ^{10, 11}.

The soya Lecithin (30%) phospholipid and edge activator ratio, time will be need to sonication, drug ratio as 5-FU and then rotational velocity that were choosing self-regulating variables find out. On show vesicle sizes distribution, percentage drug loading and EE % be situated designated as the reliant variables show. 5-FU ratio show that 10mg, dependents to formulation. Standards value are shown on total variables and batch number that were be described on Table 2.

In this current research spending main three features, as well as three point multiple strategies through three distinctive retorts be situated analysis. Since have outcomes different three reliant standards value be situated achieved reaching starting entrapment efficacy 84.7% to 87.24 % designed for, 0. 013 % to 8.042 % for DL and 35.09 nm to 56.25 nm for PS. The situation that was detected in quadratic equation just like Y = Ax2+Bx+c had greatest built-in designed different retorts it just like, mean particle size distribution, % Entrapment Efficacy and % Drug Loading. In this following underneath declared quadratic model have to generate below:

Y1 (PS) = 50.48 + 0.070X1 - 3.81X2 - 2.24X3 -0.98 X1 X2 + 0.070 X1 X3 - 2.89 X2 X3 - 4.20 X1 2 - 5.15 X2 2 -3.15 X3 2 (1)

Y2 (EE%) = 98.84950 + 2.06375X1 - 0.46242 X2 - 1.28528X3 +0.085250 X1 X2 - 0.070750 X1 X3 + 0.012725 X2 X3 - 1.02425 X1 2 + 4.20750E-003 X2 2 + 0.013708 X3 2 (2)

Y3 (DL%) = 15.94245 - 1.39412 X1 + 0.049570 X2 - 0.57567 X3 + 0.041625 X1 X2 + 0.050550 X1 X3 + 0.017504 X2 X3 - 0.80580 X1 2 - 0.018016 X2 2 + 2.81425E-003 X3 2

Where, X1, X2 and X3 defines as disguised integrities that will rational assessment of soya lecithin (30%) phospholipid and edge activator ratio, time will be need to sonication, rotation speed and drug ratio as 5-FU correspondingly. The optimistic (+) symbol of an element that were affected exceeding equation designates synergistic outcome or an improvement to particular reply and dissipation visa.

IR absorption spectroscopy for drugs

Drug illustrations were trituration particularly through dried

Nidhi Jain / Afr.J.Bio.Sc. 6(10) (2024)

separately. One by one drugs (5-FU and topotecan pellets) were pass out IR-ray in ranges between 400-4000cm-1^{22, 23}.

X-ray diffraction

X-ray diffraction Study of pure drug 5-FU sharp diffraction peak showed at 2θ value of 13.2, 15.5, 16.6, 18 and 23, soya lecithin (30%) phospholipid presented specific peak at 2θ value of 11.3 and 23.5. Their crystal-like structure of mixture for 5-FU was clearly peak shown in figure 3.

In vitro Drug Release

The results of *in-vitro* release curve study of 5-FU entrapped transfersomes, pure drug suspended formulation and marketed available product using in a phosphate buffer solution of pH 5.5 ($37 \pm 0.5^{\circ}$ C) are effects be present in Figure 4.

Release kinetics for 5-FU entrapping transfersomes

Release kinetics study performed the optimized formulation of 5-FU entrapping transfersomes was associated to various kinetic models. An obtained outcomes represent that the best fitted model was data within the Higuchian equation (R2 = 0.987). In this model describes the drug release profile from an unsolvable medium in time dependent parameter that were based on Fickian diffusion^{24, 25}.

Drug Stability studies

The drug stability studies shown that here was an insignificant enhance in the particle size range from 35.45 ± 0.58 to 36.85 ± 3.43 nm through the storage temperature (4oC and 25oC). The early % EE of the elevated transfersomes.

Ex-vivo skin permeation studies

Skin penetration study of different formulations by Confocal Laser Microscopy [CLSM]

To the study in skin penetration effect determined in naval transferosomal hydrogel preparation (PDH, MP, OTS and TH) by confocal laser microscopy evaluation performing invitro applied on goat skin. Goat skin was working for the intention to evaluated skin penetration capability of prepared hydrogel since that goat's skin is physically and physiologically like that of human skin. The skin saturations of all the transferosomal formulations (PDH, MP, OTS and TH) were pretreated evaluated to Rhodamine 123.

RESULTS AND DISCUSSION

Optimization of transferosome and FT-IR- VisSpectral Analysis

Physical appraisal of transferome preparation shown that image on Figure 1. FTIR spectrum of puree from drug of 5-FU shown as distinctive peaks at 3736 cm-1 for NH stretching. The C=O stretching indicate peaks 1733 cm-1 and -C=C- stretching shown at 1660 cm-1. The characteristic of CH in plane deformation start on 1568.6cm-1 and end of plane deformation wereexposed at 399.193cm-1 respectively and data interval 0.964233 cm-1.





Figure 1: Optical approval for Transferosomal preparation and FT-IR imagination

The Entrapment Efficiency (% EE) of deformable Nano size vesicles preparations were show selected in the value range of 68.25 ± 0.24 to 85.05 ± 0.58 (Table 9T-10). The greater % EE considerably (P<0.05) with enhancing edge activator surfactant concentration since 5-10% (w/w) for transfersomes formulated by Tween 80, again enhanced edge activator surfactant concentration to 10% (w/w) after that 15% (w/w) show that a considerable changes of EE% (P<0.05) decreased, Table 9T-10. The rational value concentration 90:10% (w/w) showed that ideal EE%, Table 2. Upon integration of edge activator surfactant in small concentration, so appeared growth in vesicle size however; further enhanced in the edge activator surfactant concentration so may have directed to small pore creation in the bilayers formulated transferosame. While edge activator surfactant more than 15% concentration, integrated micelles cohabited through the transfersomes as with the significance of lesser drug 5-florouracil entrapment owing to the stiffness and lesser size of varied micelles [3, 9]. Patel et al. described that, the consequence of soya lecithin (30%) phospholipids and edge activator surfactant rational value in the lipid constituents of transfersomal vesicles depend on the EE% of lipophilic drug, 5-florouracil, and the decreased with increasing edge activator surfactant efficacy concentration. Drug loading too interact upon of soya lecithin (30%) phospholipid & edge activator surfactant rational value.

The current experiment shown that drug loading % values was increased as well as soya lecithin (30%) phospholipids and edge activator surfactant concentration for only in few batches, and overall reliant variables is limited values shown but edge activator surfactant influences significantly in a positive behavior. This Model shown that F-value of 15.85 indicates the model is prominently. There is just only a 0.08% coincidental changes that an F-value. This big could take place due to noise. This values of "Prob > F" below 0.0500 specify model rapports are greatly.

The zeta potential was shown that good strength and stability range from -9.22 to 14.26mv. Detected values of zeta potential signify that the surface potential charges of the particles in different prepared formulation was shown negative values The vesicles size investigation values for transfersomes determination with the help Malvern Mastersizer exposed size ranging from 35.19 nm to 57.25 nm..





Figure 2 Fig. 9F-11. 3d Surface plots showing the effect of variables on (A) Particle Size (B) % Entrapment Efficiency and (C) % Drug loading.

Table 1: Different Adaptable factorsalso their determinestageswith the help of Box-Behnken design modelaimed at optimized of 5-FU entrapped transfersomes

A dontable factors	Stages			
Adaptable factors	-1	0	1	
Liberated Adaptable Factors				
X1= Soya Lecithin (30%) phospholipid: edge activator ratio,	95:05:00	90:10:00	85:1 5:00	
X2= time will be need to sonication	10min	20min	30m in	
X3= RPM (round per minute)	20	40	60	
Deliberated Adaptable Factors				
Y1= particle size distribution (PSD)	minimize			
Y2= %EE	maximize			
Y3= %DL	constants			

The outcome shown that very short% bias, its indicating that have the optimizing different formulation be present constant & rational variables. First variables independent variables shown as X1= Soya Lecithin (30%) phospholipid: edge activator ratio, X2= time will be need to sonication, X3= RPM(round per minute). Objective variables were show that close -1, 0 &+1 conforming to the small, medium, and large standards value correspondingly.

The controlled retorts Y1= particle size distribution (PSD) &Y2= %EE and Y3= %DL through constrictions related for optimization of 5-FU entrapped transfersomes are designated. Additional 3-D comeback surface graphology shown that designed for screening on belongings of programmed different factors affecting on the deliberate retorts. 3-D comeback surface graphology are supportive on illumination the association interact to liberated adaptable factors and deliberated adaptable

factors.

 Table 2: Actul experimental value design and expected values actual response.

F. Code	X ₁ ,P/EA	X ₂ ,ST	X _{3,} R	EE(%)	DL(%)	PDI	ZP(mV)
P-1	95:5(1)	20 (0)	60(1)	76.54 ± 0.21	8.015 ± 0.025	0.542±0.121	8.52±0.22
P-2	95:5(1)	30(1)	40 (0)	73.52 ± 0.42	5.085 ± 0.006	0.452±0.452	7.51 ± 0.8
P-3	85:15(-1)	10 (-1)	40 (0)	69.08 ± 0.32	2.017 ± 0.124	0.331±0.521	5.23 ± 0.5
P-4	85:15(-1)	30(1)	40 (0)	72.21 ± 0.84	0.351 ± 0.251	0.152±0.113	7.13 ± 0.2
P-5	90:10(0)	20 (0)	40 (0)	72.26 ± 0.52	3.275 ± 0.25	0.253±0.503	8.78 ± 0.3
P-6	90:10(0)	20 (0)	40 (0)	76.42 ± 0.14	2.017 ± 0.052	0.754±0.124	13.15 ± 0.2
P-7	95:5(1)	20(0)	20 (-1)	79.56 ± 0.12	4.921 ± 0.057	0.157±0.542	11.52 ± 0.6
P-8	85:15(0)	20(0)	40(0)	69.95 ± 0.29	3.163 ± 0.231	0.325±0.542	9.65 ± 0.32
P-9	95:5(1)	10(-1)	40 (0)	68.25 ± 0.24	4.215 ± 0.026	0.252±0.142	13.25 ± 0.5
P-10	85:15(-1)	20(0)	20 (-1)	72.52 ± 0.88	5.152 ± 0.055	0.523±0.521	8.52 ± 0.8
P-11	90:10(0)	20(0)	40 (0)	71.86 ± 0.53	5.053 ± 0.028	0.523±0.521	10.52 ± 0.6
P-12	90:10(0)	30(1)	20 (-1)	75.45 ± 0.85	2.052 ± 0.125	0.412±0.103	11.45 ± 0.5
P-13	85:15(-1)	20(0)	60(1)	75.69 ± 0.56	4.95 ± 0.052	0.54±0.542	7.52 ± 0.52
P-14	90:10(0)	20(0)	40 (0)	72.52 ± 0.25	9.52 ± 0.415	0.595±0.254	10.15 ± 0.8
P-15	90:10(0)	10 (-1)	20 (-1)	77.58 ± 0.26	8.582 ± 0.089	0.458±0.854	11.85 ± 0.5
P-16	90:10(0)	30 (1)	60 (1)	85.05 ± 0.58	8.054 ± 0.152	0.285±0.125	14.50 ±0.8
P-17	90:10(0)	10 (-1)	60(1)	75.56 ± 0.57	0.026 ± 0.025	0.582±0.452	10.85 ± 0.54

P/EA= Phospholipid/Edge activator, ST= Sonication time, R= Revolution per minute

Table 3: Comparative estimation of the experimental value and expected values in the planned below to experimental and expected conditions.

7.94
0.70
8.58

*Biaswas calculatedas [(Expected Value–Experimental Value)/Expectedvalue]×100%.

X-ray diffraction Study

Their crystal-like structure of mixture for 5-FU was clearly peak shown in figure 3. Whereas the transfersome formulation showed curved peak for 5-FU, that was representing the comparative decrease in the diffraction strengths in the transfersomes.

In vitro Drug Release

In our study reports, the cumulative % drug release of the optimized transfersomal formulation, pure drug suspended formulation and marketed available product diffusion drug release in 24 hr. was determined 61.68%, 70.85%, and 74.85%, correspondingly.

Release kinetics study

Release kinetics study performed the optimized formulation of 5-FU entrapping transfersomes was associated to various kinetic models. An obtained outcomes represent that the best fitted model was data within the Higuchian equation (R2 = 0.987). In this model describes the drug release profile from an unsolvable medium in time dependent parameter that were based on Fickian diffusion.

The drug stability studies

The drug stability studies shown that here was an insignificant enhance in the particle size range from 35.45 ± 0.58 to 36.85 ± 3.43 nm through the storage temperature (4°C and 25°C). The early % EE of the elevated transfersomes was originate to be 84.54 ± 0.52 %. Later 6 months storage temperature freeze to room temp. (4oC and 25oC) it was determined to be $81.52\pm0.65\%$ and $78.88\pm0.46\%$ correspondingly.

Nidhi Jain / Afr.J.Bio.Sc. 6(10) (2024)

Table 4:. The outcomes effect described that In vitro drug release profile of 5-FU

Time Interval	Optimized formulation	Marketed product	Pure drug suspension
0	0	0	0
2	9.25 ± 0.58	15.15 ± 0.87	26.52 ± 2.14
4	20.21 ± 1.5	30.84 ± 2.54	38.54 ± 3.08
6	32.21 ± 0.58	45.85 ± 1.52	54.12 ± 0.95
12	47.87 ± 2.18	61.24 ± 078	74.18 ± 0.76
24	61.25 ± 1.85	70.34 ± 1.52	14.18 ± 1.59



Figure 3 X-ray diffraction study of (a) 5-FU, (b) Soya Lacithin (30%) phospholipid, (c) Physical mixture and (d) Transfersome formulation

Ex-vivo skin permeation studies

To the study in skin penetration effect determined in naval transferosomal hydrogel preparation (PDH, MP, OTS and TH) by confocal laser microscopy evaluation performing invitro applied on goat skin. Goat skin was working for the intention to evaluated skin penetration capability of prepared hydrogel since that goat's skin is physically and physiologically like that of human skin. The skin saturations of all the transferosomal formulations (PDH, MP, OTS and TH) were pretreated evaluated to Rhodamine 123. In brief, taken the test samples of hydrogel formulation and the probe covering 0.03% of rhodamine 123 were directed applied unvaryingly and non- conclusively to the skin. The trials were done utilizing Franz dispersion cells with the collector chamber loaded up with phosphate support pH 5.5 adjusted. 24 hrs. Later, the skin was isolated and cleansed with phosphate buffer solution. The skin was then quickly frozen by fluid nitrogen and a skin surface cut opposite rectangular piece was taken from the site of medication application with



Figure 4. In vitro drug release profile of 5-FU entrapped transfersome & marketed formulation and pure drug suspension in skin pH 5.5.



Figure 5: (a) Showing Skin penetration ability of PDG formulations through Confocal solution Laser Microscopy at 0-10 μ m and 0-25 μ m (b) Fig 9F-20: Showing Skin penetration ability of MP formulations through Confocal Laser Microscopy at 0-10 μ m and 0₅25 μ m.

the assistance of a sharp edge. This tissue was prepared and fixed on the sample holder with the assistance of a Tissue frozen medium hydrogel. (Gung,Leica, Germany). The skin catted out vertical sections (dermis to horny layer) at 250µm full depth were cut through the help of cry microtome (Leica, Germany). The frozen region was removed and prepared to test for probe penetration. The frozen skin depth was optically scanned at 15-30nm raises through the Z-pivot of a Leica DMIRE2 confocal laser checking magnifying lens (Germany) connected to a Leica TCS SP2. The full skin thickness was optically examined at 15-30nm additions through the Z-hub of a Leica DMIRE2 confocal laser filtering magnifying instrument. In-vitro drug release propositioned the exploded release of drug was that was accessibility of the free 5-FU in the upper most surface on the transfersome. The sustained and prolong release of the drug was owing to 5-FU and possibly will be the object for the sustained and prolong release of the drug since the internal layer phospholipid phase afterward the initial exploded release. It was showed to be non-irritant to the topically skin and adept to retain drug 5-FU.

CONCLUSION

entrapped soya-lecithin 30% The 5-fluorouracil transferosomal preparation in this broad fields have the great beneficial effect. Results showed that the average particle size, polydispersity index, and zeta potential were 35.58±0.56, 0.285±0.125, and 14.50 ±0.8 respectively, for 5-FU loaded transfersomes. The highest encapsulation efficiency achieved was 85.05 ± 0.58 for P-16 formulation. P-16 formulation compared with marketed preparation on the basis of drug release, it shows 61% release by P-16 and 70% respectively. The stability of transferosome was calculated with a variability of different physicochemical parameter. Various amounts use of transferome, constant temperatures, and different marketed preparation concentrations were all evaluated. This Model shown that F-value of 15.85 indicates the model is prominently. There is just only a 0.08% coincidental changes that an F-value. This big could take place due to noise. This values of "Prob > F" below 0.0500 specify model rapports are greatly. The proved by FTIR studies the spectrum of puree from drug of 5-FU shown as distinctive peaks at 3736 cm-1 for NH stretching. The C=O stretching indicate peaks 1733 cm-1 and -C=C- stretching shown at 1660 cm-1 X-ray diffraction Study of pure drug 5-FU sharp diffraction peak showed at 2θ value of 13.2, 15.5, 16.6, 18 and 23, soya lecithin (30%) phospholipid presented specific peak at 2θ value of 11.3 and 23.5The conclusions shows that the preparation signifies exploded release phase resultant to approximately 10-15% was experiential time within 2 hour due to the drug desorption and transfersomes release from the upper surface. Such that the transferossome have determination continuing and different biomolecules presentations in the upcoming duration, seeing the cost-effective, non-hazareous and uncomplicated methods used for their synthesis.

ACKNOWLEDGMENT

The SAIF and my guide, Oriental University, Indore, as well as IISER Bhopal, supported X-ray diffraction analysis.

REFERENCES

- Sağir T, Huysal M, Durmus Z, Kurt BZ, Senel M, Isık S. Preparation and in vitro evaluation of 5-flourouracil loaded magnetite–zeolite nanocomposite (5-FU-MZNC) for cancer drug delivery applications. Biomedicine & Pharmacotherapy. 2016 Feb 1;77:182-90.
- 2. Cheng M, He B, Wan T, Zhu W, Han J, Zha B, Chen H, Yang F, Li Q, Wang W, Xu H. 5-Fluorouracil nanoparticles inhibit hepatocellular carcinoma via activation of the p53 pathway in the orthotopic transplant mouse model.
- Singh S, Kotla NG, Tomar S, Maddiboyina B, Webster TJ, Sharma D, Sunnapu O. A nanomedicine-promising approach to provide an appropriate colon-targeted drug delivery system for 5-fluorouracil. International Journal of Nanomedicine. 2015 Nov 23:7175-82.
- Okada KI, Hirono S, Kawai M, Miyazawa M, Shimizu A, Kitahata Y, Ueno M, Hayami S, Yamaue H. Phase I Study of Nab–Paclitaxel plus Gemcitabine as Neoadjuvant Therapy for Borderline Resectable Pancreatic Cancer. Anticancer Research. 2017 Feb 1;37(2):853-8.

- Han L, Tang C, Yin C. Dual-targeting and pH/redox-responsive multi-layered nanocomplexes for smart co-delivery of doxorubicin and siRNA. Biomaterials. 2015 Aug 1;60:42-52.
- Xian XS, Park H, Choi MG, Park JM. Cannabinoid receptor agonist as an alternative drug in 5-fluorouracil-resistant gastric cancer cells. Anticancer Research. 2013 Jun 1;33(6):2541-7.
- Patel R, Singh SK, Singh S, Sheth NR, Gendle R. Development and characterization of curcumin loaded transfersome for transdermal delivery. Journal of pharmaceutical sciences and research. 2009 Dec 1;1(4):71.
- Nasr M, Ghorab MK, Abdelazem A. In vitro and in vivo evaluation of cubosomes containing 5-fluorouracil for liver targeting. Acta pharmaceutica sinica B. 2015 Jan 1;5(1):79-88.
- Duangjit S, Opanasopit P, Rojanarata T, Ngawhirunpat T. Characterization and in vitro skin permeation of meloxicamloaded liposomes versus transfersomes. Journal of drug delivery. 2011;2011.
- Sabitha M, Rejinold NS, Nair A, Lakshmanan VK, Nair SV, Jayakumar R. Development and evaluation of 5-fluorouracil loaded chitin nanogels for treatment of skin cancer. Carbohydrate polymers. 2013 Jan 2;91(1):48-57.
- Palem CR, Dudhipala N, Battu SK, Goda S, Repka MA, Yamsani MR. Combined dosage form of pioglitazone and felodipine as mucoadhesive pellets via hot melt extrusion for improved buccal delivery with application of quality by design approach. Journal of Drug Delivery Science and Technology. 2015 Dec 1;30:209-19.
- Sarwa KK, Mazumder B, Rudrapal M, Verma VK. Potential of capsaicin-loaded transfersomes in arthritic rats. Drug delivery. 2015 Jul 4;22(5):638-46.
- Jangdey MS, Gupta A, Saraf S, Saraf S. Development and optimization of apigenin-loaded transfersomal system for skin cancer delivery: in vitro evaluation. Artificial Cells, Nanomedicine, and Biotechnology. 2017 Oct 3;45(7):1452-62.
- Jain SK, Gupta A. Development of Gelucire 43/01 beads of metformin hydrochloride for floating delivery. AAPS PharmSciTech. 2009 Dec;10:1128-36.
- Khan S, Boateng JS, Mitchell J, Trivedi V. Formulation, characterisation and stabilisation of buccal films for paediatric drug delivery of omeprazole. Aaps Pharmscitech. 2015 Aug;16(4):800-10.
- 16. Alvi IA, Madan J, Kaushik D, Sardana S, Pandey RS, Ali A. Comparative study of transfersomes, liposomes, and niosomes for topical delivery of 5-fluorouracil to skin cancer cells: preparation, characterization, in-vitro release, and cytotoxicity analysis. Anti-cancer drugs. 2011 Sep 1;22(8):774-82.
- based quartz crystal microbalance biosensor for sensitive and selective detection of leukemia cells using silver-enhanced gold nanoparticle label. Talanta. 2014;126:130–5. https://doi. org/10.1016/j.talanta.2014.03.056
- Levitas VI, Samani K. Size and mechanics effects in surfaceinduced melting of nanoparticles. Nature Communication. 2011;2(1). https://doi.org/10.1038/ncomms1275.
- Aitken RJ, Chaudhry MQ, Boxall ABA, Hull M. Manufacture and use of nanomaterials: Current status in the UK and global trends. Occupational Medicine. 2006;56: 300–6. https://doi. org/10.1093/occmed/kql051.
- He H, Xie C, Ren J. Nonbleaching f luorescence of gold nanoparticles and its applications in cancer cell imaging. Analytical Chemistry. 2008;80(15):5951–7. https://doi. org/10.1021/ac8005796.
- 21. Naik RR, Stringer SJ, Agarwal G, Jones SE, Stone MO.

Nidhi Jain / Afr.J.Bio.Sc. 6(10) (2024)

Biomimetic synthesis and patterning of silver nanoparticles. Nature Materials. 2002;1(3):169–72. https://doi.org/10.1038/nmat758.

- Modi CD, Bharadia PD, "Transfersomes: New Dominants for Transdermal Drug Delivery", Am. J. PharmTech Res., 2012; 2 (3): 71-91.
- 23. Prajapati ST, Patel CG, Patel CN, "Transfersomes: A Vesicular Carrier System for Transdermal Drug Delivery", Asian Journal of Biochemical and Pharmaceutical Research, 2011; 2 (1): 507-524.
- 24. Kombath RV, Minumula SK, Sockalingam A, Subadhra S, Parre S, Reddy TR, David B, "Critical issues related to transfersomes – novel Vesicular system", Acta Sci. Pol., Technol. Aliment., 2012; 11 (1):67-82.
- 25. Swarnlata S, Gunjan J, Chanchal DK, Shailendra S, "Development of novel herbal cosmetic cream with Curcuma longa extract loaded transfersomes for anti-wrinkle effect", African J Pharm Pharmacol, 2011; 5 (8) : 1054-1062.
- 26. Schatzlein A, Cevc G, "Skin penetration by phospholipids vesicles, Transfersomes as visualized by means of the Confocal Scanning Laser Microscopy, in characterization, metabolism, and novel biological applications" AOCS Press, 1995: 191-209.
- Chaurasiya P, , Ganju E, Upmanyu N, Ray SK, Jain P "Transfersomes: a novel technique for transdermal drug delivery", Journal of Drug Delivery and Therapeutics,2019; 9(1):279-285 http://dx.doi.org/10.22270/jddt.v9i1.2198
- 28. Cevc, G., (1991). Isothermal lipid phase. Transitions Chemistry and Physics of Lipids, 57, 293- 299.
- 29. Cevc, G., Blume G., Schatzlein A. Transferosomes mediated transepidermal delivery improves the regiospecificity and biological activity of corticosteroids in vivo. J. Control. Release. 1997; 45:211.
- Jain S., Jain P. Transferosomes: A novel vesicular carrier for enhanced transdermal delivery: development, characterization and performance evaluation, Drug Dev. Ind. Pharm. 2003, 29: 1013-1026.
- 31. Swarnlata S, Gunjan J, Chanchal DK, Shailendra S, "Development of novel herbal cosmetic cream with Curcuma longa extract loaded transfersomes for anti-wrinkle effect", African J Pharm Pharmacol, 2011; 5 (8) : 1054-1062.
- Kaushik, A., Sunda, M., Transfersomes: the drug loaded ultradeformable vesicles for transdermal drug delivery. IRJP, 2011; 2(11): 40-42.
- Vijaya R, Ruckmani K. In vitro and In vivo characterization of the transdermal delivery of sertraline hydrochloride Films. Daru. 2011; 19: 424–432.
- Mishra D, Garg M, Dubey V, Jain S, Jain NK. Elastic liposomes mediated transdermal delivery of an antihypertensive agent: Propranolol hydrochloride. J Pharm Sci. 2007; 96: 145–155.
- Walve, J.R, Bakliwal S.R, Rane B.R, Pawar S.P. Tansfersomes: a surrogated carrier for transdermal drug delivery system. www.ijabpt.com, 2011. 2: 204-213.
- 36. Sachan, R., Parashar, T, Singh, V., Singh, G., Tyagi, S., Patel, C., Gupta, A, Drug Carrier Transfersomes: A Novel Tool For Transdermal Drug Delivery System. Int. J. Res. Dev. Pharm. L. Sci. 2013, 2(2), 309-316.