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Evaluation Of *Jatropha Gossypifolia* Linn's Leaf Extracts And Its Antimicrobial And Anthelmintic Activity In Wound Healing

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Abstract

The leaves of *Jatropha gossypifolia* Linn. By take on extracts of the leaf prepared with petroleum Ether, methanol, and aqueous medium. Methods: The various experimental models that were used for the evaluation of wound healing, antimicrobial, and anthelmintic activities are as follows: Excision wound model: Wound contraction, epithelization time (days), and area (sq. mm) on complete epithelization were studied. Antimicrobial wound model: Wounds were treated with an antimicrobial agent to decrease the number of microorganisms that caused infection. Anthelmintic worm model Tensile strength (g) measurement of restore incision wound was measured for the incision wound model. Antimicrobial activity was determined make use of the cylindrical plate method on a variety of bacterial and fungal organisms. Pet.ether, methanol, and aqueous leaves extracts of *Jatropha gossypifolia* Linn. Have been found to display notable wound healing activity as compared to the control. This was insistent using an excision wound model. When differentiate to the control group, all three leaves extracts of *Jatropha gossypifolia* Linn. Demonstrated a substantial increase in shatter strength in the result incision wound model. It was find that methanol and aqueous extracts had indestructible antibacterial action at concentrations of 25 and 50 mg/ml, despite the fact that pet. Ether extract appear very weak antibacterial activity against gram positive along with gram negative microorganisms. It was discovered that both concentrations of all thrine extracts possessed a level of antifungal activity that ranged from modest to modest when tested on fungal species. Pet. ether, methanol, and aqueous leaf extracts of *Jatropha gossypifolia* have all demonstrated a low level of action in comparison to the standard medicine.

Keywords–epithelization time, *Jatropha gossypifolia*, antimicrobial agent, fungal organisms

Introduction –

The typical anatomical structure and functional capabilities of live tissues are both altered when a wound is present. Healing a wounded is a dynamic process that starts at the moment that the damage is first caused. In its most basic form, it is a survival strategy that also symbolises an effort to keep normal anatomical structure and function.

There is a wide variety of synthetic medication on the market now that can accelerate the process for healing process. However, in the modern day, a significant portion of the population of the world relies on herbal treatments to treat a wide variety of disorders.

Boils, carbuncles, eczema, and itches are some of the conditions that can be treated locally with *Jatropha gossypifolia* (Euphorbiaceae) leaves, which are used in the traditional system of medicine. One form of emmenagogue involves the use of a decoction made from the bark. Worm infestation and ulcers are two more conditions that benefit from using this herb. 2 The leaves of *Jatropha gossypifolia* contain flavonoids³, which are an important ingredient of the leaves. It has been demonstrated that the flavonoids that are found in plants have the ability to promote wound healing. 4,5 The leaves of *Jatropha gossypifolia* have not been shown to have any antibacterial, anthelmintic, or wound–healing properties, according to a review of the relevant scientific literature. In light of these considerations, the current research was conceived with the objective of investigating the potential effects of several leaf extracts of *Jatropha gossypifolia* on the wound healing process, in addition to their antibacterial and anthelmintic activities.



Objective –The primary goals of the research were to apply scientific techniques of evaluation in order to analyse the wound healing, antibacterial, and anthelmintic properties of leaf extracts from *Jatropha gossypifolia*. As a result, the current study was initiated with the goals that will be discussed below. Extraction from the *Jatropha gossypifolia* leaves after they have been dried. using a variety of different solvents. An initial phytochemical examination of the crude extracts. The LD50 value was determined by using mice. Evaluation of wound healing activities utilising the various experimental models that are listed below.

- A model of a wound caused by excision.
- A model of an incision wound

Antimicrobial activity evaluations include the following:

- Actions that is antibacterial
- Fungicidal and antimycotic action

LITERATURE REVIEW

Ashrub, 0.9–1.8 m in height with palmately 3–

5lobedleaves,easilyrecognisedbythestipitate,yellowviscidglands,whichcovertheleafmargins,petioles and stipules, and by the small red flowers in glandular corymbose cymes.

Hindi-: *Bherenda*

Kan-:Doddaharalu

Mal-:*Simayaranak*

San-: *Raktaeranda*

Tam-:*Attalai*

Beng-:*Lalbherenda*

Chemical Constituents¹

Alanine, arginine, cystine, glycine, leycine, isoleucine, methionone, valine, linoleic, oleic, palmitic acids; proteins (Seeds); apigenin, viterxin, and isovitexin. Ceryl and myricyl alcohols, carnavbyl palmitate, patmitone, and beta-sitosterol are the components that make up this compound. Coumarino-lignans, fraxetin, jatrophone, jatropholones, A and B, 2 and 2 - hydroxy jatrophones, and 2 -hydroxy - 5,6 - iso jatrophone are some of the components of jatrophone.

Medicinal Uses²

Leaves: Locallyapplied inboils,carbuncles, eczemaand itches.

Bark: Adecoctionof the barkis used as emmenagogueTheplantisalsobeneficial in ulcers and worminfestation.

Photographof *Jatropha gossypifolia* (Linn) Plant.

A search of the relevant published material reveals that *Jatropha gossypifolia* has been found to be reported for the following uses:

- The presence of antimicrobial activities in crude fruit extracts of *Jatropha gossypifolia*⁶
- Flavonoids found in the *Jatropha gossypifolia* plant's leaves³.
- An extremely uncommon diterpene derived from *Jatropha gossypifolia*⁷
- The effects of an ethanolic extract from *Jatropha gossypifolia* on rats for lowering blood pressure and relaxing the blood vessels.
- Gossypidien is a lignan that is extracted from the stem of *Jatropha gossypifolia*.
- Cleomiscosin A, a coumarino lignoid from *Jatropha gossypifolia*.
- *Jatropha* seed oils for use in the production of energy ¹¹.
- Gossypifan is a lignan that comes from the plant ***Jatropha gossypifolia***¹².

PHASESOFWOUNDHEALING:

1. The Phase of Inflammation-

In this phase, the immediate response to damage is a constriction of the tiny blood arteries and capillaries in the area surrounding the lesion. This is called the vasoconstriction phase. At the site of the injury, vascular occlusion can take place, which has the effect of reducing or stopping bleeding. After the vasodilation has taken place, this response continues for the immediate 5-10 minutes,^{13, 14} that follow. All components of the local autocoid system are involved in this vasodilation.¹⁵

2. The phase of repair-The processes of repair start fairly immediately after the harm has occurred. At the location of a wound, the first cells to develop in great numbers are called polymorphonuclear (PMN) leucocytes. PMN leucocytes are essential to the healing process of wounds because they

phagocytose bacteria that may be present at the wound site in the event that the lesion becomes infected. In addition to this, it is known that they clean the area around the wound of any dead cells and debris, so preparing it for the regeneration of new cells. Additionally, fibroblasts migrate into the area surrounding the wound and deposit collagen. 16

3. Fibroblastic phase—Shortly after the injury, undifferentiated mesenchymal cells begin to convert into migrating fibroblast, which then move to the wound area. This happens after PMN leucocyte clean the wound area of the debris. Enzymes are secreted by fibroblasts, which are responsible for the conversion of fibrinogen to fibrin. This fibrin both acts as a haemostatic barrier and provides a framework for the other components of wound closure. The ground substance is composed of proteins, polysaccharides, and a variety of other glycoproteins, all of which are secreted by fibroblasts. The matrix's mucopolysaccharides envelop the fibroblasts, rendering them stationary and preventing their movement. It has an effect on the manner in which collagen aggregates and is oriented. The fibroblast is responsible for the production of collagen; in the process, it makes use of hydroxyproline and hydroxylysine. These procedures often take place after the fourth day after the wound has been caused. In most cases, the fibroblastic phase of wound healing lasts between two and four weeks. The formation of capillaries and a slowdown in the rate at which collagen is being synthesised both signify the beginning of the end of the fibroblastic phase. 17

4. The phase of epithelization includes— One of the most important steps in the process of wound healing is epithelial cell migration. It is necessary for the stem cells of the epithelium to move into the wound after they have detached from the borders of the lesion. In a healthy dermis, basal dermal cells will attach to both one another and the underlined basal layer of the dermis. After being mobilised, epithelial cells start to expand and migrate across and down the wound. This process occurs after mobilisation. Epithelial cells that migrate across a wound typically move along the basal lamina or fibrin deposits. This process is known as contact guiding, and it is an important feature in epithelial migration. Transected hair follicles also contribute to the amount of migrating epithelial cells. After epithelial migration, there is an increase in the amount of mytosis that occurs in the epithelium. Recent research suggests that the modulation of mytosis¹⁸ is controlled by a water-soluble, heat-labile chemical termed chalone, which is released near the wound site.

5. Reduction—Wound contraction reduces a full-thickness wound. Centripetal movement of surrounding skin thickness heals open lesions. All evidence leads to cell-mediated contraction. Electron microscopy shows that certain contracting wound fibroblasts resemble smooth muscle cells. These are myofibroblasts. Smooth muscle and fibroblast-like. In vitro pharmacological investigations show granuloma tissue strips contract or relax like smooth muscle strips. Smooth muscle relaxants reduce wound contraction in vivo. 19,20 Myofibroblasts aid wound healing. Observations show that myofibroblasts connect to the wound bed, penicularis, and dermis of the wound edge. 21,22 Once contraction begins, it continues until wound edges meet inhibition or until surrounding skin tension equals or exceeds contraction force. During contraction, wound skin is stretched and thinned. This state is short-lived. This process continues until the stretched skin's entire thickness is restored. intussusceptive growth 21

6. Renovation—This phase has early and late stages. Early wound strength is attributed to fibrin clot development. Epithalization helps heal wounds.wound healing. New capillaries embedded in the wound help strengthen it. 21

14 to 16 days post-wound, wound strength grows dramatically. Increased wound strength under fast fibroblastic and collagen synthesis and deposition. Hydroxyproline, a collagen precursor. 16, 17New collagen fibres are put down in the late stabilising phase. Collagenase digests and removes excess wound cells. After wound collagen stabilises, wound strength increases. Scar collagen affects

tensile strength¹⁴.

Wound repair parameters

TableNo.-1

Sl.No	Attributes	Type of wound	Parameter studied	Material used
1	Physical	Incision (resutured)	Scar Modeling	Planimetry
		Excision	Contraction, epithelization scar remodeling	Planimetry
		Dead space wound	Granuloma dry weight	Weighing
		Chemical granulomas	Granuloma size	Plenthysmogrph
		Burn and Freeze wound	Contraction epithelization & scar remodeling	Planimetry
		Cellular injury by X-radiation ultraviolet light etc.,	Contraction epithelization & scar remodeling	Planimetry
2.	Mechanical	Incision (resutured)	Breaking tensile strength	& Tensiometer
		Poly vinyl sponge cotton pellet & other foreign body induced granulomas	-do-	-do-
	Biochemical	Burn and Freeze wound	Collagen & MPS content	Hydroxyprolin or flexosamin
Sl.No	Attributes	Type of wound	Parameter studied	Material used
3	Biochemical	Incision (resutured)	Colloagen mucopolysaccharide (MPS) Content	& Hyroxyprolin Hexosamine estimation
		Poly vinyl sponge cotton pellet 7 other foreign body induced granulomas	Collagen & MPS content	„
		Burn and Freeze wound	„	„
		Cellular injury by X-radiation & ultra violet, light etc.,	„	„
04	Histological	Excision	Cellular elements fibroblasts & collagen fibres	Microscopy
		Poly vinyl sponge cotton pellet 7 other foreign body induced granulomas	„	„
		Cellular injury by X-radiation & ultra violet light etc.,	Cellular elements collagen fibres	„

HOW DRUGS AFFECT THE RECOVERY OF A WOUND

Wound healing is aided by a number of naturally occurring substances as well (cytokines and growth factors). Wound healing can be stimulated or slowed down by a wide variety of medications. When it comes to wound healing, these chemicals can either stimulate or depress the synthesis of certain growth factors.

Aspirin, phenylbutazone, and indomethacin are just a few examples of medications that may slow the healing process of a wound. Large doses of aspirin have been shown to reduce the tensile strength of wounds in rats. 27

By slowing protein synthesis and lysosomal membrane stability, cortisone and its derivatives also reduce the body's normal inflammatory response. Steroids have been shown to slow wound healing in general. Wound healing in animals is not affected by vitamin A administration, despite evidence that vitamin A stimulates fibroblasts and increases collagen synthesis in the tissue. 32

Wound healing and collagen formation are both slowed by vitamin E's ability to maintain the membrane. Wound healing can be facilitated by vitamin C. Collagen synthesis is dependent on the presence of vitamin C. Collagen molecules are incomplete in the absence of vitamin C, and fibroblasts may be unable to release it. Wound healing is slowed down when vitamin C deficiency is present. 33 Animals with low zinc levels can have their wounds healed when zinc is administered to them. 34 While epithelial and fibroblast cells can move to the site of a lesion, normal proliferation of these cells requires zinc-dependent enzymes such as DNA polymerase and reverse transcriptase. can't get any more. Consequently, epithelization takes longer, and the wound's collagen synthesis also declines. 34 Local administration of cytotoxic medicines has been shown to impair wound healing. 5-fluororacil and meclorothamine are just two examples of this type of medication.

Anti-Microbial Agent Screening Methods-

Methods of screening are included in which solely measure the growth inhibition (bacterio-static). 48

1. Cup plate technique
2. The procedure of serial dilution
3. Ditch plate technique
4. The process of solid dilution
5. Gradient plate technique

Cup plate technique-During this step of the process, the test solution is allowed to come into contact with agar that has previously been inoculated with the test organism. Once the incubation process is complete, zones of inhibition can be seen. The test solution can either be administered in the form of an impregnated disc of filter paper or it can be placed in a small cup that is sealed to the surface of the agar in a well that has been cut out of the agar using a sterile corkborer.

The procedure of serial dilution- The serial dilution approach involves the incorporation of graded doses of test chemicals into broth, followed by the incubation of tubes that have been infected with the organism under investigation. The concentration at which there is no visible sign of growth is referred to as the lowest inhibitory concentration (MIC).

Ditch plate technique-In this method, the test solution is either poured directly into a ditch that has been carved into nutrient agar inside of a petri dish, or it is first mixed with a little amount of nutrient agar before being poured into the ditch. The test organisms, of which there may be as many as six, are scattered throughout the ditch in a random pattern. After that, the plate is allowed to incubate while antibacterial activity is monitored.

Solid dilution method—A dilution of the material being tested is generated using this technique, however rather than using broth, agar is used. After that, the agar medium that is now being evaluated is transferred to a petri dish, after which it is placed in an incubator and monitored for any signs of abnormal growth. This method has the benefit that multiple species can be examined using a single concentration of the test chemical.

Gradient plate technique—Using this method, the amount of medication that is deposited onto an agar plate can be adjusted in an unrestricted manner between 0 and some predetermined maximum value. In order to carry out the procedure, nutritious agar is first allowed to melt, then the solution being evaluated is added to the molten agar, and finally the entire thing is placed into a clean petri dish and allowed to harden into the shape of a wedge. After that, an additional quantity of agar is poured onto the wedge, and the petri dish is laid down flat on the bench while it is let to set. In order to allow for the diffusion of the drug and to dry the surface, the plates are placed in an incubator for the night. The organisms that will be put to the test need to be streaked in a path that runs from the highest concentration to the lowest concentration. This method can be used to test anywhere from one to six different species.

List of materials and equipment's used during experiments

S.N	Nameofthetmaterialandequipment's
1	Agar
2	Anaestheticether
3	Beef extract
4	Calciumchloride
5	Caseinhydrolysateofsoyabeen
6	Ciprofloxacin
7	Dextrose (dehydrated)
8	Distilled water
9	Electronicbalance
10	Ethyl alcohol
11	Glucose
12	Griseofulvin
13	Methanol
14	Peptone
15	Petroleumether
16	Silk thread
17	Sodiumbicarbonate
18	Sodiumchloride
19	Soxhlet apparatus
20	Yeast extract

METHODOLOGY

- Screening for potential phytochemicals at an early stage
- Evaluation of the Immediate Dangers to Humans (LD50)
- Activities related to the healing of wounds
 - Excision(openwound)model
 - Incisionmodel
- Antimicrobial activity

- Antibacterial activity.
- Antifungal activity.
- Activity against parasitic worms.

Screening for potential phytochemicals at an early stage

Animals used– For the purpose of this investigation, albino rats of either sex (wistar strain) weighing 150–200 g and albino mice of the female gender weighing 20–25 g were utilised. They came from Sri Venkateshwara Enterprises in Bangalore, which is where we got them. Ten days were spent in the lab gradually acclimating the animals to the environment there. They were confined in cages made of polypropylene and kept at a temperature of 270 degrees Celsius with a relative humidity of 65 to 10 percent. The light and dark cycle lasted for 12 hours. The rats were given an ad libitum supply of water and a rodent pellet diet that was manufactured by Gold Mohur Lipton India Ltd. The Institutional Animal Ethics Committee gave its blessing for the researchers to carry out the tests on the animals they had in their care (IAEC).

Determination of acute toxicity (LD₅₀)–The female albino mice were used to test the acute toxicity of pet. ether, methanolic, and aqueous extracts of *J. gossypifolia* Leaf. Prior to the experiment, the animals were allowed to go without food or water for a full 24 hours. The fixed dose approach of OECD guideline No. 420; (Annexure–2d) of CPCSEA78 was used for this endeavour. Each test dose was administered to a group of three mice, and the screening dose for wound healing activity was determined to be one tenth of the LD₅₀ cutoff value for the extracts being tested.

Healing procedures for wounds

Studies on wound healing have been conducted, and the parameters, together with their details, are supplied. For the purpose of systemic administration in excision and incision wound models, pet. ether and methanol leaf extracts of *J. gossypifolia* were suspended in 2% tween 80, and aqueous extract was diluted in distilled water. On the other hand, all three extracts were combined into a single ointment at a concentration of ten percent and were applied topically to the excision wound model solely.

Conclusion

- A preliminary screening for phytochemicals was carried out. Different phytoconstituents were found in ether, methanolic, and aqueous extracts of the leaves of *J. gossypifolia*.
- At a dose of 2000 mg/kg b.w., both aqueous and petroleum ether extracts of *J. gossypifolia* leaf were lethal to the mice (all three perished). Extracts were safe to consume at a dosage of 300 mg/kg. 500 milligrammes per kilogramme was the lethal dose for both petroleum ether and aqueous extracts..
- In tests of the substance's acute toxicity, a dose of 2000 mg/kg of *J. gossypifolia* methanol extract was sufficient to kill two-thirds of the mice. Extract at a dose of 300 mg/kg did not cause death. The median lethal dose (LD₅₀) of methanol extract was determined to be 1000 mg/kg.
- When compared to the control groups, the topical application of extracts at a concentration of ten percent in an ointment and oral administration of pet. ether, methanol, and aqueous extracts of *J. gossypifolia* leaf promoted wound contraction on different days.
- The breaking strength of incision wounds was significantly increased by systemic injection of *J. gossypifolia* leaf extracts, specifically pet. ether (382 8.84), methanol (325 7.25), and water (440 8.96), in comparison to the control group (246 6.46).
- The methanol and aqueous extracts at 25 and 50 mg/ml had significant antibacterial activity, whereas the pet. ether extract had only weak antibacterial activity against gram-positive bacteria

such as *Staphylococcus aureus* and *Streptococcus pyrogene*, as well as gram-negative bacteria such as *Escherichia coli* and *Pseudomonas aeruginosa* (gram negative bacteria).

- At concentrations of 25 and 50 mg/ml, all three extracts exhibited low to moderate antifungal efficacy against *Candida albicans* and *Aspergillus flavus*.

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