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# **Sciences**



GC-FID, LC-QToF-MS, NMR, quantum chemical analysis, and toxicological evaluation of lemongrass (*Cymbopogon flexuosus*) essential oil yields in Mayurbhanj district of Odisha state

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# Abstract

The main objective of this research is to characterize the volatile chemical components found in essential oil of lemongrass leaves. Lemongrass cultivation is done in very few districts of Odisha and leaf sample collected from Mayurbhanj district. Essential oil from leaves was extracted by steam distillation process and oil was analyzed by gas chromatography (GC), liquid chromatography mass spectroscopy (LCMS) and nuclear magnetic resonance (NMR). We have reported the phytochemicals, material safety and toxicological effects observed in lemongrass leaves cultivated at Mayurbhanj district of Odisha state. Lemongrass oil chemistry profile reveals 1.4805 refractive index and 0.888 specific gravity at 27°C, exceeding norms. GC analysis reveals phytochemicals such as Camphene (0.834%; Rt. Time: 6.872 min), 6-Methyle-5-heptene-2-One (0.452%; Rt. Time: 7.251 min), 4-Nonanone (1.082%; Rt. Time: 9.755 min), and Citronellol (0.104%, Rt. Time: 15.341 min). Using LCMS analysis, 40 phenolic compounds were discovered. Proton-induced chemical shift on carbon displays signals at  $\delta 1.484$ , 2.000, 7.187, and 9.747 ppm, corresponding to methyl, methylene, aromatic, and aldehyde groups. Carbonyl group is present in the  $\delta$ 1.484-1.967 range. Quantum chemical structure analysis of citral isomers revealed highest and lowest electron charge occupancy. Not previously reported, lemongrass leaves from Mayurbhanj had the highest citral content. We also stressed hazard identification, and toxicological studies showed LD 50 was 4400 mg/Kg in oral rat treatment. Not previously reported, lemongrass oil was stable at room temperature and had a flash point of 71°C. This research also expects new farming methods to boost essential oil, citral yields, and toxic levels.

**Keywords:** *Cymbopogon flexuosus*; gas-liquid chromatography; nuclear magnetic resonance; citra; essential oil.

#### **1** Introduction

Lemongrass is the common name for the plant *Cymbopogon flexuosus*, which is a member of the Gramineae family [1]. Citral, a cyclic monoterpene, is responsible for the prefix lemon's characteristic lemon-like aroma. Numerous essential oils, tannins, flavonoids, and alkaloids are among the phytoconstituents found in lemongrass [2]. Numerous components' secondary active

Article History Volume 6, Issue 13, 2024 Received: 18June 2024 Accepted: 02July 2024 doi:10.48047/AFJBS.6.13.2024. 2008-2031 metabolites have also been linked to the diverse pharmacological actions of this plant. From the Greek word "kymbe-pogon" which translates as "boat-beard," gets the name Cymbopogon [3]. The plant's essential oil is utilized in culinary herbs, medicinal tea, aromatherapy, and treatments for skin conditions. It is recognized as a source of traditional remedies [4]. In several regions of the world, C. flexuosus is used to treat rheumatism, fevers, menstrual abnormalities, and other joint difficulties in addition to digestive problems [5]. This species is a member of the Gramineae family, which consists around 8,000 plant species and about 500 genera [6]. Lemon grass is a tufted perennial grass that may reach a height of 1 meter and has several stiff, green stems that emerge from roots that are short and rhizomatous. Practitioners of folk medicine have continued to treat illnesses and crippling diseases mostly with whole herbs and extracts [7]. It is used in the food and perfume sectors as a good source of citral [8]. It serves as the raw material to produce ionones, which create vitamin A. Due to its extensive usage in the cosmetics, food, pharmaceutical, agricultural, and taste sectors, it grows throughout subtropical and tropical parts of the world [9]. The subtropics and tropics are where Cymbopogon grasses are widely cultivated. Cymbopogon flexuosus smells like lemons because it contains more aldehydes than other plants [10]. It has two geometric isomers: neral (Citral- $\beta$ ) and geranial (Citral- $\alpha$ ). One of the elements that affects the creation of fragrant lemongrass is the usage of exceptional seeds. Farmers' revenues will rise as production levels of fragrant lemongrass change [11]. According to research, growers of lemongrass who used premium varietals made more money than those who used local seeds. In addition to using excellent seeds, the best and most efficient labor will yield the greatest advantage. Through farmer organizations or community leaders, it is necessary to promote the use of better seeds by supplying seed sources, encouraging community involvement, and offering pertinent technical assistance.

#### Agro-climatic zones of Odisha

In India, there are fifteen main agroclimatic zones. Odisha, a state on the eastern coast of India, belongs to the Easter Plateau and Hilly Region agroclimatic zone VII. However, twelve agroclimatic zones can be further subdivided into the state at the regional level based on climate, temperature, relief, and rainfall [12]. The details of agro-climatic zones of Odisha given below:



Figure 1: Physiography of different agro-climatic zones in Odisha.

	Table 1:	Details of	agro-climatic	zones with	climate, sc	oil profile	e and rain	fall [13].
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SI. No	Agro- climatic Zone	Agricultural Districts	Broad Soil groups	Different (Normal)	parameters	of climate	Climate
				Mean	Mean	Mean	
				annual	maximum	minimum	
				rain	summer	winter	
				fall	temp(°C)	temp(°C)	
				(mm)			
1.	North	Sundargarh,		1600			Hot & moist
	Western	parts of	Red,		38.0	15.0	sub-humid
	Plateau	Deogarh,	Brown				
		Sambalpur	Forest,				
		&	Red &				
		Jharsuguda	Yellow,				

#### Krishnendu Adhikary /Afr.J.Bio.Sc. 6(13)(2024).2008-2031

			Mixed Red				
			& Black				
2.	North	Mayurbhanj,		1534	36.6	11.1	Hot & moist
	Central	Majorparts of	Lateritic,				sub-humid
	Plateau	Keonjhar,	Red &				
		(except	Yellow,				
		Anandapur	Mixed Red				
		& Ghasipura	& Black				
		block)					
3.	North	Balasore,		1568	36.0	14.8	
	Eastern	Bhadrak, parts	Red,				Moist
	Coastal	of Jajpur &	Lateritic,				sub-
	Plain	Hatdihi block	Deltalc				humid
		of Keonjhar	alluvial,				
			Coastal				
			alluvial				
			&Saline				
4.		Kendrapara,	Saline,	1577	39.0	11.5	
	East &	Khurda,	Lateritic,				Hot &
	South	Jagatsinghpur,	Alluvial,				Humid
	Eastern	part of Cuttack,	Red				
	Coastal	Puri, Nayagarh	&Mixed				
	Plain	& part of	red&				
		Ganjam	Black				
5.		Phulbani,		1597	37.0	10.4	Hot &
	North	Rayagada,	Brown				moist, sub-
	Eastern	Gajapati, part of	forest,				humid
	Ghat	Ganjam &	Lateritic				
		small patches	Alluvial,				
		of Koraput	Red,				
			Mixed Red				
_			& Black				
6.	Eastern Ghat	Major parts	5 1	1522	34.1	7.5	Warm &
	High Land	of Koraput,	Red,				humid
		Nabarangpu	Mixed Red				
		r	& Black,				
			& reliow				

#### Krishnendu Adhikary /Afr.J.Bio.Sc. 6(13)(2024).2008-2031

7.	South	Malkangiri &	Red,	1710	34.1	13.2	Warm &
	Eastern	part of Keonjhar	Lateritic,				humid
	Ghat		Black				
8.		Kalahan	Red,	1352	37.8	11.9	Hot & moist
	Western	di &	Mixed Red				sub-humid
	Undulating	Nuapada	& Black				
	Zone		and Black				
9.		Bargarh,		1614	40.0	12.4	Hot & moist
	Western	Bolangir,	Red &				sub-humid
	Central Table	Boudh,	Yellow,				
	Land	Sonepur, parts	Red &				
		of Sambalpur	Black,				
		& Jharsuguda	Black,				
			Brown				
			forest,				
			Lateritic				
10.				1421			
	Mid central	Angul, Dhen	Alluvial,		38.7	14.0	Hot & moist
	table land	kanal, parts of	red,				sub-humid
		Cuttack &	lateritic,				
		Jajpur	mixed				
			red& black				

#### 2 Materials and method

#### Plant material collection and preparation

Lemongrass sample (Sample ID: LG24CUTM2024) has been collected from the cultivating field in Baripada block (Co-ordinates: Latitude- 21.998709<sup>0</sup>; Longitude- 87.011419<sup>0</sup>) of Mayurbhanj district of Odisha state. The slips of leaves are cut into 70-80 cm and dried in sunlight for 24 hours.

#### Oil extraction by steam distillation

The steam distillation unit is equipped with two vessel/drum with 500 Kg ×2 nos. which is equal to 1 ton capacity. Vessel is manufactured with 304 stainless steel (SS) with 3mm thickness. Vapour sealing is done with water. A boiler unit (Water tube) is attached to each vessel for warm water supply [14]. Boiler (Water tube) outer shell and fire box is made with mild steel (MS), top cover and chimney is made up of MS or Galvanized iron (GI). Boiler quality tubes are used and

glass tube water gauge with auto release pressure gauge is attached with a drain valve 1" ball valve. SS 304 quality separator is used to separate oil from water. MS shell base attached with SS ball valve 80 mm x 2 nos. A connecting pipe with SS 304 73 mm outer diameter connects boiler to shell. 1" mm GI pipe used to supply water from condenser to boiler. Two hot water outlet valve with 1"mm brass ball valve is attached to shell grid which is made with SS 304 (1550 dia  $\times$  2 nos.). On the top of each vessels are attached to a cone shaped steam collector which is directly connected to a condenser and further the condenser is connected to an oil collecting tube. Lemongrass essential oil is collected through steam distillation method, each drum is filled with 500 Kg dried glass and oil is collected through separator [15].



Figure 2: Diagrams showed the steam distillation unit from different views.

# Physical and chemical properties

Physical and chemical properties are measured with appearance and smell, Specific Gravity was measure by Bottle with Stopper - 25 ml and refractive index was measured by Digital refractometer model DR602.

# Gas chromatography (GC)

Modifications were made to the approach described by Uraku (2015) and Thenmozhi and Rajan (2015) for the GC analysis [16, 17]. Environmental Monitoring Services, Auroville, conducted the analysis. Using a flame ionizing detector (FID), the GC-FID (Model: Thermo Scientific Trace 1110 Series) was used to perform the GC analysis. The apparatus is equipped with a dual micro channel plate ion detector, low acceleration ion-transfer system, reflection type TOF analyser, EI ion source, and rotary pump ventilation facilities. The apparatus has a capillary column (SGE Analytical Science - Model: BP-1) measuring 25 meters in length, 0.32 millimeters in diameter, and 0.5 micrometers in thickness. The carrier gas was helium, which

was used at a split ratio of 1:45 with an injection volume of 1 ml and a flow rate of 1 ml/min. Oil sample was made as 25 ml sample in 0.5 ml hexane. The programming temperature for the injector line was set at 180°C and 250°C, respectively. The oven was set to start at 90 degrees Celsius (isothermal for two minutes), increase by 8 degrees Celsius per minute to 250 degrees Celsius, grow by 8 degrees Celsius per minute (isothermal for three minutes) to reach 250 degrees Celsius, then increase by 5°C per minute to 280 degrees Celsius, culminating in five minutes of isothermal operation at 280°C. The GC-FID ran for 37 minutes in total [18].

# Liquid chromatography mass spectroscopy (LCMS)

Electrospray ionization mass spectrometry (ESI-MS) was used to identify the chemical components of the essential oil of lemongrass. High-resolution mass spectroscopy (HRMS) (Waters; Xebo-G2-XS-QTof) was used for the study. An ESI-equipped Q-TOF mass spectrometer was interfaced with the HPLC. During MS scanning, the full-scan mode was configured with a 200°C source temperature and a mass range of m/z 50 to 2000. A maximum flow velocity of 5  $\mu$ l/min was employed to provide the methanol solvent. The positively charged ion mode was used to get the MS spectra. The nebulizing pressure (N<sub>2</sub>) was 25 psi, the boiling point of the drying gas (N<sub>2</sub>) was 350°C, and the gas flow rate was 6 ml/min. Before analysis, about 0.5 g of extracts from the samples were mixed with methanol and screened through a 0.22  $\mu$ m nylon filter. The mass fragmentations were determined using the organic chemical spectrum-database found in MASS Lynx 4.1 (Waters, USA) [19].

## Nuclear magnetic resonance (NMR) spectroscopy

NMR spectroscopy was performed to obtain 1H-NMR spectra of *C. flexuosus* extract, which was recorded using NMR (Bruker Avance III running under TOPSPIN 2.4, Wilmad 535-PP-7 tube) spectrophotometer with operating frequency of 700 MHz at 289 K temperature, the spectra obtained of 10 ppm width. The evaluation of the authenticity and determination of the major constituents of commercial citronella oils were performed by the 1H NMR method developed herein. The analyses were carried out to acquire high-quality spectra; sample run for single scans and chemical shifts were reported in parts per millions (ppm). Stock solution of OMCTS in CDCl<sub>3</sub> (5.59 mg/ml) were prepared to determine repeatability and accuracy/precision of the NMR method. The <sup>1</sup>H spectra were with a single scan in just 15 seconds per sample. To remove the carbon satellites from the <sup>1</sup>H spectra, the signal was acquired in the presence of carbon decoupling [20-23].

# Quantum chemical structure analysis of citral

The quantum chemical analysis of citral isomers,  $\alpha$  and  $\beta$ -citral, was conducted to gain insights into their electronic structure and potential reactivity. This analysis involved computational methods to simulate molecular structures and calculate various electronic properties, providing valuable information on their chemical behavior and interaction. DMol3 in the Material Studio programme (Biovia, France) employing the B3LYP functional and a 6-31G (d,p) basis set was used to accurately calculate the molecular and electronic structure of the oil.

# Other specifications and measurements

With above tests, hazard identification, first aid measures, fire fighting measures, toxicological tests, and regulatory measures are done.

# **3** Results and discussions

Lemongrass sample identification and compound characterization are shown in Table 2 and 3

 Table 2: Composition Identifications lemongrass essential oil.

# **Composition Identifications**

Identity	:	Lemongrass oil (Krishna Variety)
Components	:	Essential oil 100%
CAS Number	:	8007-02-1
EC (EINECS) Number	:	295-161-9
FEMA Number	:	2624

**Table 3:** Physical and chemical parameters with tabulated data.

Analytical test	Specification range	Result
Appearance	Mobile oily liquid bright yellow	Conforms
Odour	Characteristic citral, fresh and	Conforms
	intense	
Relative density at 27°C (g/ml)	0.886-0.896	0.888
Refractive index at 27°C	1.4830–1.4890	1.4805

Optical rotation	-3° to +1°	
Solubility in 70% ethyl alcohol		
1in 2-3 parts	Soluble/ not soluble	Soluble

#### Gas chromatography analysis

As shown by the chromatogram, a total of twelve components with various retention durations were eluted from the GC column and subjected to further analysis using an electron impact MS voyager detector. The retention period and mass spectroscopy library search were used to identify the ingredients. These are the mass spectrographs of the components that have been identified. GC peak regions were used to compute the relative amounts of each component. Gas chromatography analysis detected the presence of certain phytochemicals in the sample. These include Camphene 0.834% with retention time: 6.872 min, 6-Methyle-5-heptene-2-One 0.452%; retention time: 7.251 min, 4-Nonanone 1.082%; retention time: 9.755 min, and Citronellol 0.104%; retention time: 15.341 min (Figure 3, Table 4).



Figure 3: Chromatogram of lemongrass oil.

**Table 4:** Shows retention time in minute, area in mV/s height in mV, height in % and area in % along with name of compounds found in lemongrass essential oil.

Sl. No.	Reten.	Area	Height	Height	Area	Compound Name
	Time	[mV/s]	[mV]	[%]	[%]	
	[min]					
1.	5.019	0.883	0.246	0.1	0.031	
2.	6.331	2.873	0.864	0.3	0.100	
3.	6.528	4.096	1.231	0.4	0.143	
4.	6.872	23.931	6.996	2.1	0.834	Camphene (a)
5.	7.251	12.957	3.467	1.0	0.452	6-Methyl-5-heptene-2-0ne
						(b)
6.	7.488	1.289	0.362	0.1	0.045	
7.	7.651	4.667	0.655	0.2	0.163	
8.	8.797	14.024	3.695	1.1	0.489	
9.	9.149	5.269	1.402	0.4	0.184	
10.	9.755	31.041	7.868	2.4	1.082	4-Nonanone (c)
11.	10.568	2.449	0.524	0.2	0.085	
12.	10.808	30.876	7.251	2.2	1.076	Linalool (d)
13.	10.979	4.967	0.971	0.3	0.173	
14.	12.096	10.024	1.976	0.6	0.349	
15.	12.395	16.597	3.349	1.0	0.578	
16.	12.773	30.306	6.976	2.1	1.056	Isoneral (e)
17.	13.259	2.026	0.488	0.1	0.071	
18.	13.411	52.605	10.374	3.1	1.833	Isogeranial (f)
19.	13.733	3.154	0.475	0.1	0.110	
20.	14.131	2.763	0.591	0.2	0.096	
21.	14.344	3.845	0.718	0.2	0.134	
22.	14.960	1.434	0.288	0.1	0.050	
23.	15.341	2.988	0.393	0.1	0.104	Citronellol (g)
24.	15.885	893.591	95.784	28.8	31.139	Citral $\beta$ (h)
25.	16.547	193.087	16.408	4.9	6.729	Geraniol (i)
26.	17.027	1276.31	122.405	36.8	44.476	Citral α (j)
		0				
27.	17.181	5.275	0.805	0.2	0.184	
28.	18.136	2.746	0.569	0.2	0.096	
29.	19.221	3.526	0.522	0.2	0.123	

30.	19.389	4.199	0.503	0.2	0.146	
31.	20.144	1.692	0.256	0.1	0.059	
32.	20.635	22.611	1.782	0.5	0.788	
33.	21.221	66.340	12.683	3.8	2.312	Geranyl acetate (k)
34.	21.475	6.667	0.707	0.2	0.232	
35.	22.168	3.952	0.440	0.1	0.138	
36.	23.392	27.756	4.834	1.5	0.967	$\beta$ -Caryophyllene (1)
37.	23.677	3.936	0.526	0.2	0.137	
38.	24.661	3.170	0.567	0.2	0.110	
39.	25.611	2.566	0.381	0.1	0.089	
40.	26.205	3.954	0.513	0.2	0.138	
41.	26.517	2.334	0.280	0.1	0.081	
42.	26.771	27.774	5.103	1.5	0.968	
43.	26.952	4.818	0.928	0.3	0.168	
44.	27.048	12.822	1.437	0.4	0.447	
45.	27.955	1.905	0.305	0.1	0.066	
46.	28.816	3.162	0.229	0.1	0.110	
47.	29.301	18.329	2.636	0.8	0.639	
48.	30.179	3.784	0.325	0.1	0.132	
49.	33.053	2.847	0.209	0.1	0.099	
50.	36.709	2.702	0.427	0.1	0.094	
51.	37.387	2.756	0.218	0.1	0.096	
	Total	2869.676	332.939	100.0	100.000	

# LCMS analysis

Scientists have recently been interested in green herbaceous plants after discovering that they contain high concentrations of bioactive substances, particularly phenolic compounds, which have shown promise for health benefits in a number of studies. Phenolic compounds from lemongrass oil (*C. flexuous*) were screened and described using the most recent LC-ESI-QTOF-MS technique. With the use of sophisticated analytical techniques, we are able to identify the majority of the bioactive components found in plant samples. Agilent Mass Hunter, version B.06.00, and the Personal Compound Database Library (PCDA) were used to qualitatively describe the compounds. The data pertaining to the phenolic chemical profile of plants are listed in Table 5. Using LC/MS analysis, 40 phenolic compounds were discovered (Figure 4, Table 5).



Figure 4: LC-QToF-MS analysis of lemongrass essential oil extracted from *C. flexuosus*.

# Table 5: LC-QToF-MS/ MS characterization of phytochemicals extracted from lemongrass

leaves.

Formula	Mass Error	Theoretical	Description	m/z	Retention
	(ppm)	Isotope			time (min)
		Distribution			
C27H48N6O6	-38.92298538	100 - 32.2 -	N-Acetyl-L-leucyl-L-isoleucylglycyl-L-valyl-N-methyl-L-	288.1693	0.236667
		6.25 - 0.9 -	prolinamide		
		0.105			
C31H44N4O5	19.64104697	100 - 35.7 -	Pandamine	288.1693	0.236667
		7.21 - 1.06 -			
		0.124			
C24H40O5	-27.51622527	100 - 26.6 -	(3alpha,5beta,7alpha)-3,7,14-Trihydroxycholan-24-oic acid	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	ursocholic acid	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	beta-Phocaecholate	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	3Beta,7beta,12beta-trihydroxy-5beta-cholan-24-oic acid	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	5a-Cholic acid	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	3a,6a,12a-Trihydroxy-5b-cholan-24-oic acid	373.2625	0.0506
		4.43 - 0.551			
C20H36O6	10.78061116	100 - 22.3 -	8,8a-Deoxyoleandolide	373.2625	0.0506
		3.6 - 0.433			
C24H40O5	-27.51622527	100 - 26.6 -	3a,4b,7a-Trihydroxy-5b-cholanoic acid	373.2625	0.0506
		4.43 - 0.551			
C27H36O3	24.20837509	100 - 29.7 -	Quingestanol acetate	373.2625	0.0506
		4.88 - 0.575			
C24H40O5	-27.51622527	100 - 26.6 -	cholic acid	373.2625	0.0506
		4.43 - 0.551			
C24H36O3	-30.2092888	100 - 26.5 -	3-Ethoxyandrosta-3,5-dien-17beta-ol propanoate	373.2625	0.0506
		3.98 - 0.437			
C24H36O3	-30.2092888	100 - 26.5 -	ANAGESTONE ACETATE	373.2625	0.0506
		3.98 - 0.437			
C26H36N2O2	-3.298080539	100 - 29.3 -	2-[2-(4-Benzyl-1-piperazinyl) ethyl]-2,5,7,8-tetramethyl-6-chromanol	373.2625	0.0506
		4.56 - 0.498			
C24H40O5	-27.51622527	100 - 26.6 -	(3alpha,5beta,7alpha,8xi,9xi,10xi,12alpha,13xi,14xi,17xi,20xi)-	373.2625	0.0506
		4.43 - 0.551	3,7,12-Trihydroxycholan-24-oic acid		
C24H40O5	-27.51622527	100 - 26.6 -	3alpha,7alpha,12beta-trihydroxy-5beta-cholan-24-oic acid	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	Vulpecholate	373.2625	0.0506
		4.43 - 0.551			

C24H40O5	-27.51622527	100 - 26.6 -	1b,3a,7a-Trihydroxy-5b-cholanoic acid	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	2b,3a,7a-Trihydroxy-5b-cholanoic acid	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	(3beta,4beta,5beta,7alpha)-3,4,7-Trihydroxycholan-24-oic acid	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	1b,3a,12a-Trihydroxy-5b-cholanoic acid	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	omega-Muricholate	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	alpha-Muricholate	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	ÃŽÂ <sup>2</sup> -Muricholic Acid	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	(3beta,5beta,6alpha,7alpha)-3,6,7-Trihydroxycholan-24-oic acid	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	(3beta,5beta,6alpha,7beta)-3,6,7-Trihydroxycholan-24-oic acid	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	(3beta,5beta,6beta,7alpha)-3,6,7-Trihydroxycholan-24-oic acid	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	(3beta,5beta,6beta,7beta)-3,6,7-Trihydroxycholan-24-oic acid	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	(3alpha,5alpha,6alpha,7beta)-3,6,7-Trihydroxycholan-24-oic acid	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	(3alpha,5alpha,6beta,7beta)-3,6,7-Trihydroxycholan-24-oic acid	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	(2beta,3alpha,5beta,12alpha)-2,3,12-Trihydroxycholan-24-oic acid	373.2625	0.0506
		4.43 - 0.551			
C24H40O5	-27.51622527	100 - 26.6 -	3a,4b,12a-Trihydroxy-5b-cholanoic acid	373.2625	0.0506
~		4.43 - 0.551			0.0506
C24H40O5	-27.51622527	100 - 26.6 -	(3beta,4beta,5beta,12alpha)-3,4,12-1rihydroxycholan-24-oic acid	373.2625	0.0506
CA 177 10 C -	25.51(22525	4.43 - 0.551			0.0506
C24H40O5	-27.51622527	100 - 26.6 -	(3alpha,5beta,6beta,12alpha)-3,6,12-1rihydroxycholan-24-oic acid	373.2625	0.0506
C2 411 40 0 5	27.51(22527	4.43 - 0.551		272 2625	0.0506
C24H40O5	-27.51622527	100 - 26.6 -	(3beta,5beta,6alpha,12alpha)-3,6,12-1rihydroxycholan-24-oic acid	3/3.2625	0.0506
C2 411 40.05	27.51(22527	4.43 - 0.551		272 2625	0.0506
C24H40O5	-27.51622527	100 - 20.0 -	(3beta, 5beta, 6beta, 12aipna)-3, 6, 12-1rinydroxycnolan-24-olc acid	373.2625	0.0506
C24114005	27.51(22527	4.45 - 0.551	2h 7a 10a Taibudaann fh' abalanais said	272 2625	0.0506
024114005	-27.31022327	100 - 20.0 -	50, / a, 12a- 11 III yoloxy-50-cholanoic acio	5/5.2025	0.0506
C24U4005	77 51677577	4.43 - 0.331 100 - 26 6	3bata 7alaba 19bata Tribudrovy 5bata abalan 24 aig Asid	272 7675	0.0504
024114000	-27.31022327	100 - 20.0 -	שלים אין	575.2023	0.0300
C24H4005	-27 51622527	100 - 26 6	3a 7h 12h Tribydroxy, 5h cholanoic soid	373 7675	0.0506
024114005	-27.31022327	100 - 20.0 -	5a, rb, r2b- minyuroxy-5b-cholahole aciu	575.2023	0.0300
		4.45 - 0.331			

#### NMR analysis

<sup>1</sup>H-NMR is effective technique for metabolites study in plant extract, as it analyses all the metabolites present in extract. Figure 4 illustrated the <sup>1</sup>H-NMR spectrum of *C. flexuosus* extract. Chemical shift obtained from <sup>1</sup>H-NMR spectra can be due to proton on carbon or proton on Oxygen/ Nitrogen. Chemical shift procured from proton on carbon is shown by signal at  $\delta$ 1.484, 2.000, 7.187 and 9.747 ppm which correspond to methyl, methylene and aromatic and aldehyde groups, respectively. The value range between  $\delta$ 1.484–1.967 shows the presence of carbonyl group. Single range between  $\delta$ 4.023–7.416 due to the shift for proton on Oxygen/ Nitrogen which account for the presence of alcohol in the extract. Occurrence of allylic group in extract, is due to the shift acquired between  $\delta$ 1.619–2.046. The Peak Fabrics obtained from NMR profile illustrated that *C. flexuosus* extract comprising of mixture of terpenoids and flavonoids.



**Figure 5:** <sup>1</sup>H-NMR spectrum of *C. flexuosus* extract. Chemical shift obtained from <sup>1</sup>H-NMR spectra can be due to proton on carbon or proton on Oxygen/ Nitrogen.

# Quantum chemical structure analysis of citral

Employing the B3LYP function with a 6-31G (d,p) basis set, quantum chemical calculations were carried out using DMol3 in the Material Studio software (Biovia, France). This analysis provided insights into the electronic structure of citral- $\alpha$  and  $\beta$  isomers derived from lemongrass

oil through steam distillation, which have been shown in Figure 6. The obtained results revealed that the charges of the citral alpha and beta structures for the highest occupied molecular orbital (HOMO) are -5.289 eV and -5.331 eV, respectively. Whereas the charges of the citral alpha and beta structures for the lowest occupied molecular orbital (LUMO) are -2.457 eV and -2.511 eV, respectively. These electronic structure details are visually presented in Figure 6.



**Figure 6:** Highest occupied molecular orbital (HOMO) and lowest occupied molecular orbital (LUMO) chemical structure of citral- $\alpha$  and citral- $\beta$  in lemongrass essential oil (LGEO) extracted by steam distillation (SD) method.

#### Hazard classification

Risk signs, risk symbols, R phrases, and/or S phrases need to be used to designate the kind of danger. Hazardous compositions and compounds are associated with two numbers and two phrases, respectively, called R and R number. S number and S phrase: they are terms and numbers corresponding to the safe management of risky preparations. There are various combinations of Risk Phrases (R) and Safety Phrases (S) as well.

Krishnendu Adhikary /Afr.J.Bio.Sc. 6(13)(2024).2008-2031





**Figure 6:** Label elements GHS signal word DANGER hazard pictograms. Hazard determining components of labeling for citral (GHS07), geraniol (GHS05).

**Table 6:** Hazardous information for safety measures and toxic impact.

Hazards Identification
Flam. Liq.4 H227: Combustible liquid
Skin Irrit.2 H315: Causes skin irritation
Skin Sens.1 H317: Might result in an adverse skin response
Eye Damage H318: Causes severe eye damage
Aquatic Acute H402: Harmful to aquatic organisms
Aquatic Chronic H411: Long-lasting toxic impacts on aquatic life

**Table 7:** Regulatory information for first aid and fire-fighting measures.

R phrases		
R38	:	Irritating to skin
R36	:	Irritating to eyes
R43	:	May cause sensitization by skin contact
Safety Phrases		
S7	:	Keep container tightly closed.
S16	:	Keep away from source of ignition
S24/25	:	Avoid contact with skin and eyes.
FDA no	:	182.2
FEMA no	:	2831
HS Code	:	33012933

#### First aid and Fire-fighting measures

If oil accidentally gets in your eyes, take off your contact lenses if needed. Spend fifteen minutes flushing with a lot of water or an eye wash solution. If there has been skin contact, take off any contaminated clothes. Use plenty of water to flush and wash with soap. Apply an emollient to the affected skin. In the event that oil is accidentally consumed, dilute it with water or milk, call the poison control center, or get medical help right away. Avoid breathing in vapors in a disgusting way if you must inhale. If there was a significant inhalation, remove to fresh air. As quickly as you can, take the sufferer to a secure location. Loosen belts, waistbands, ties, and collars that are too tight. Give oxygen if breathing is difficult. Put the person via mouth-to-mouth resuscitation if they are not breathing. LGEO is purely non-flammable, its flash point>100 Deg by close cup technique. Extinguishing media can be used like dry chemical, carbon dioxide, foam. It was recommended to not use water jet. Additionally, it was mandated that firefighters use positive pressure, self-contained breathing apparatus as part of particular firefighting protocols. Water spray cools containers that are in direct flame contact. Apply extinguishing agent straight into the fire's base. There was no unexpected fire or explosive risks found.

# Stability and reactivity

Under typical circumstances, LGEO is stable. It was advised to avoid using materials that were incompatible with powerful oxidizing agents, as well as heated labor and ignition sources close or on empty containers. A few other situations that were to be avoided included heat, flame, and contact with sparks, as well as potentially dangerous decomposition products such CO, CO<sub>2</sub>, smoke, and unknown chemical compounds that could arise during combustion. It has been noted that while byproducts are produced during burning, dangerous polymerization does not take place.

#### **Toxicological information**

LGEO may be irritating to eyes, skin, and mucous membranes. Acute toxicity level in oral administration in rat was 4400 mg/Kg (LD<sub>50</sub>) in primary stage and there was a secondary dosage which was greater than 2000 mg/Kg in rat. According to Plata-Rudea et al., Lemongrass topically applied was toxic against *S. granarius* adults ( $LD_{50} = 4.03 \ \mu g \cdot insect^{-1}$ ) and mortality increased in a dose-dependent manner, as also reported in other pests [19]. According to Costa et al., LGEO did not present an extensive toxic effect in rodents, since the  $LD_{50}$  in mice was around 3500 mg/Kg, which is a much higher dose than those usually taken as an infusion by humans [24]. According to Xavier et al., LGEO and citral would be safe even at the limit dose level of 2000 mg/Kg body weight after single dose oral administration to female Sprague Dawley rats

and hence the  $LD_{50}$  could be proclaimed as more than 2000 mg/Kg, enabling to be labelled as category 5/unclassified in hazard category of Globally Harmonized System for classification of chemical [25]. There was no data available on chronic effects on humans. Other toxic effects on humans like LGEO can be hazardous in case of skin contact (irritant, permeator), of inhalation. Furthermore, LGRO was not listed as a carcinogen [26-31]. There are no published environmental criteria for this complicated composition. Garbage recycling methods are designed to ensure that no product leftovers remain. Discard in accordance with municipal, state, and Government requirements. Surface pollution of soil, ground, and surface water is prevented by taking preventative measures.

#### **4** Conclusion

The whole study i.e., plant material collection, extraction and chemistry profile analysis of lemongrass essential was done at Mayurbhanj district of Odisha state which was not reported before. The chemistry profile of the lemongrass oil shows refractive index (at 27°C) was 1.4805 (Normal range: 1.4830-1.4890) and specific gravity (at 27°C) was 0.888 (Normal range: 0.886-0.896) which was not reported before, along with this GC analysis shows the photochemical like: Camphene (0.834%; Rt. Time: 6.872 min), 6-Methyle-5-heptene-2-One (0.452%; Rt. Time: 7.251 min), 4-Nonanone (1.082%; Rt. Time: 9.755 min); Citronellol (0.104; Rt. Time: 15.341 min), Citral-β (31.139 %; Rt. Time: 15.885), Geraniol (6.729; Rt. Time: 16.547 min), Citral-α (44.476; Rt. Time: 17.027 min), Geranyl acetate (2.312; Rt. Time: 21.221 min), β-Caryophyllene (0.967, Rt. Time: 23.392). LCMS characterization reveals 40 new phenolic compounds which were not reported before. NMR spectroscopy showed the carbon atoms experience protoninduced chemical shift, resulting in signals at  $\delta$ 1.484, 2.000, 7.187, and 9.747 ppm. These signals correspond to methyl, methylene, aromatic, and aldehyde groups, respectively. The δ1.484-1.967 range encompasses the presence of a carbonyl group. Quantum chemical structure analysis revealed that the charges of the citral alpha and beta structures for the highest occupied molecular orbital (HOMO) are -5.289 eV and -5.331 eV, respectively. Whereas the charges of the citral alpha and beta structures for the lowest occupied molecular orbital (LUMO) are -2.457 eV and -2.511 eV, respectively. Among all these the amount of citral is in high concentration present is lemongrass leaves cultivated in Mayurbhanj area, which was not reported before. After all, we have also emphasized on hazard identifications and toxicological findings show toxic dose or LD<sub>50</sub> was 4400 mg/Kg in oral administration of rat. Lemongrass oil was stable under room temperature and had a flash point of 71°C which was not reported before. Moreover,

modern agricultural advancement and process may increase the yield of essential oil and citral also which is future perspective of this study.

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Informed consent: Each and every author involved in this study offered their informed consent.

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# References

- Dangol S, Poudel DK, Ojha PK, Maharjan S, Poudel A, Satyal R, et al. Essential oil composition analysis of *Cymbopogon* species from Eastern Nepal by GC-MS and chiral GC-MS, and antimicrobial activity of some major compounds. *Molecules* 2023, 28, 543.
- Ali A, Wee Pheng T, Mustafa MA. Application of lemongrass oil in vapour phase for the effective control of anthracnose of 'Sekaki' papaya. J. Appl. Microbiol. 2015, 118, 1456-1464.
- Mao GF, Mo XC, Fouad H, Abbas G, Mo JC. Attraction behaviour of Anagrus nilaparvatae to remote lemongrass (*Cymbopogon distans*) oil and its volatile compounds. *Nat. Prod. Res.* 2018, *32*, 514-520.
- Sattary M, Amini J, Hallaj R. Antifungal activity of the lemongrass and clove oil encapsulated in mesoporous silica nanoparticles against wheat's take-all disease. *Pestic. Biochem. Physiol.* 2020, *170*, 104696.
- 5. Kabotso DEK, Neglo D, Kwashie P, Agbo IA, Abaye DA. GC/MS composition and resistance modulatory inhibitory activities of three extracts of lemongrass: Citral

modulates the activities of five antibiotics at sub-inhibitory concentrations on methicillinresistant *Staphylococcus aureus*. *Chem. Biodivers*. 2022, *19*, e202200296.

- Agnish S, Sharma AD, Kaur I. Nanoemulsions (O/W) containing *Cymbopogon* pendulus essential oil: development, characterization, stability study, and evaluation of in vitro anti-bacterial, anti-inflammatory, anti-diabetic activities. *Bionanoscience* 2022, *12*, 540-554.
- Abdelsamad A, Ahmed K, Al-magboul A, Fadul E. Antimicrobial activity of essential oils and extracts of oleo-gum resins from *Boswellia papyrifera* (Tarak tarak) grown in some parts of the Sudan. *Arab. J. Med. Arom. Plants* 2020, *6*, 22-35.
- Adhikary K, Banerjee P, Barman S, Bandyopadhyay B, Bagchi D. Nutritional aspects, chemistry profile, extraction techniques of lemongrass essential oil and it's physiological benefits. *J. Am. Nutr. Assoc.* 2024, *43*, 183-200.
- Manikandan S, Bhambal AM, Ratchambiga KS, Nithiela M, Swatheka JK, Sridarshini B. Comparative evaluation of the effect of 0.2% chlorhexidine, 2% lemongrass oil, and 2% tea tree oil mouth rinse on salivary pH: An *in vivo* study. *J. Pharm. Bioallied. Sci.* 2021, *13*, 757-760.
- Avila-Sosa R, Palou E, Jiménez Munguía MT, Nevárez-Moorillón GV, Navarro Cruz AR, López-Malo A. Antifungal activity by vapor contact of essential oils added to amaranth, chitosan, or starch edible films. *Int. J. Food Microbiol.* 2012, *153*, 66-72.
- Bapatla KG, Singh AD, Sengottaiyan V, Korada RR, Yeddula S. Impact of climate change on Helicoverpa armigera voltinism in different agro-climatic zones of India. J. *Therm. Biol.* 2022, 106, 103229.
- Behera SS, Ojha CSP, Prasad KSH, Dash SS. Yield, water, and carbon footprint of rainfed rice production under the lens of mid-century climate change: a case study in the eastern coastal agro-climatic zone, Odisha, India. *Environ. Monit. Assess.* 2023, 195, 544.
- Shah AH, Khan AU, Pan L, Amin A, Chandio AA. Reflections of pro-poor growth across agro-climatic zones for farming and non-farming communities: Evidence from Punjab, Pakistan. *Int. J. Environ. Res. Public Health* 2022, *19*, 5516.
- 14. Santoro GF, Cardoso MG, Guimarães LG, Freire JM, Soares MJ. Anti-proliferative effect of the essential oil of *Cymbopogon citratus* (DC) Stapf (lemongrass) on intracellular amastigotes, bloodstream trypomastigotes and culture epimastigotes of *Trypanosoma cruzi* (Protozoa: Kinetoplastida). *Parasitology* 2007,*134*, 1649-1656.
- 15. Donato R, Sacco C, Pini G, Bilia AR. Antifungal activity of different essential oils against Malassezia pathogenic species. *J. Ethnopharmacol.* 2020, *249*, 112376.

- 16. Uraku AJ. Determination of chemical composition of *Cymbopogon citrates* leaves by gas chromatography-mass scpectrometry (GC-MS). *Res. J. Phytochem.* 2015, *9*, 175-187.
- 17. Thenmozhi S, Rajan S. GC-MS analysis of bioactive compounds in *Psidium guajava* leaves. J. Pharmacog. Phytochem. 2015, 3, 162-166.
- 18. Nolvachai Y, Marriott PJ. GC for flavonoids analysis: Past, current, and prospective trends. J. Sep. Sci. 2013, 36, 20-36.
- Kim C, Park J, Lee H, Hwang DY, Park SH, Lee H. Evaluation of the EtOAc extract of lemongrass (*Cymbopogon citratus*) as a potential skincare cosmetic material for acne vulgaris. J. Microbiol. Biotechnol. 2022, 32, 594-601.
- 20. Otify AM, Serag A, Porzel A, Ludger AW, Mohamed AF. NMR metabolome-based classification of *Cymbopogon* species: a prospect for phyto-equivalency of its different accessions using chemometric tools. *Food Anal. Methods* 2022, *15*, 2095–2106.
- 21. Basera P, Lavania M, Agnihotri A, Lal B. Analytical investigation of *Cymbopogon citratus* and exploiting the potential of developed silver nanoparticle against the dominating species of pathogenic bacteria. *Front. Microbiol.* 2019, *10*, 282.
- 22. Le QU, Lay HL, Wu MC. The isolation, structural characterization, and anticancer activity from the aerial parts of *Cymbopogon flexuosus*. J. Food Biochem. 2019, 43, e12718.
- 23. Gikuru M, Samuel M, Phyllis M, Josiah G. Lemongrass (*Cymbopogon flexuosus*) agronomic traits, oil yield and oil quality under different agro-ecological zones. J. Agri. Food Res. 2022, 10, 100422.
- 24. Plata-Rueda A, Rolim GDS, Wilcken CF, Zanuncio JC, Serrão JE, Martínez LC. Acute toxicity and sublethal effects of lemongrass essential oil and their components against the Granary Weevil, *Sitophilus granarius*. *Insects* 2020, *11*, 379.
- 25. Costa CA, Bidinotto LT, Takahira RK, Salvadori DM, Barbisan LF, Costa M. Cholesterol reduction and lack of genotoxic or toxic effects in mice after repeated 21-day oral intake of lemongrass (*Cymbopogon citratus*) essential oil. *Food Chem. Toxicol.* 2011, 49, 2268-2272.
- 26. Abo Ghanima MM, Swelum AA, Shukry M, Ibrahim SA, Abd El-Hack ME, Khafaga AF, et al. Impacts of tea tree or lemongrass essential oils supplementation on growth, immunity, carcass traits, and blood biochemical parameters of broilers reared under different stocking densities. *Poult. Sci.* 2021, *100*, 101443.
- 27. Lulekal E, Tesfaye S, Gebrechristos S, Dires K, Zenebe T, Zegeye N, et al. Phytochemical analysis and evaluation of skin irritation, acute and sub-acute toxicity

of *Cymbopogon citratus* essential oil in mice and rabbits. *Toxicol. Rep.* 2019, *6*, 1289-1294.

- Bidinotto LT, Costa CA, Costa M, Rodrigues MA, Barbisan LF. Modifying effects of lemongrass essential oil on specific tissue response to the carcinogen N-methyl-Nnitrosurea in female BALB/c mice. J. Med. Food 2012, 15, 161-168.
- 29. Kim SI, Yoon JS, Baeck SJ, Lee SH, Ahn YJ, Kwon HW. Toxicity and synergic repellency of plant essential oil mixtures with vanillin against *Aedes aegypti* (Diptera: Culicidae). J. Med. Entomol. 2012, 49, 876-885.
- 30. Tak JH, Isman MB. Metabolism of citral, the major constituent of lemongrass oil, in the cabbage looper, *Trichoplusia ni*, and effects of enzyme inhibitors on toxicity and metabolism. *Pestic. Biochem. Physiol.* 2016, *133*, 20-25.
- 31. Mukherjee T, Mohanty S, Kaur J, Das M, Adhikary K, Chatterjee P, et al. Exploring Small-Molecule Inhibitors Targeting MAPK Pathway Components: Focus on ERK, MEK1, and MEK2 Kinases in Cancer Treatment. *Chem. Bio. Letters* 2024, *11*, 659–9.