

<https://doi.org/10.48047/AFJBS.6.6.2024.5863-5878>



African Journal of Biological Sciences

Journal homepage: <http://www.afjbs.com>



Research Paper

Open Access

Computational Screening and Biochemical Analysis of *Ricinus Communis* Linn Root

Abhilasha Mittal^{1*}, Amit Subhashchand Lunkand², Mithilesh Singh¹,
Gulshan Singh Rathod¹, Mekha Monsi¹, Rahul Yadav¹, Abhishek Dubey¹ and Kunal¹

¹Department of Pharmaceutical Chemistry NIMS Institute of Pharmacy, NIMS University,
Jaipur, Rajasthan, India.

²Department of Pharmaceutical Chemistry Sitabai Thite College of Pharmacy, Shirpur,
Savitribai Phule University, Pune

***Corresponding Author:**

**Department of Pharmaceutical Chemistry
NIMS Institute of Pharmacy, NIMS University, Jaipur, Rajasthan, India.
abhilashamittal24@gmail.com**

Article Info

Volume 6, Issue 6, June 2024

Received: 23 April 2024

Accepted: 26 May 2024

Published: 20 June 2024

[doi: 10.48047/AFJBS.6.6.2024.5863-5878](https://doi.org/10.48047/AFJBS.6.6.2024.5863-5878)**ABSTRACT:**

The goal of the current investigation was to computational screening and biochemical evaluation of *Ricinus communis* Linn's root. The phytochemicals from the root of *Ricinus communis* were extracted using methanol and water. Via phytochemical screening with LC-MS, it was possible to identify the greater number of phytoconstituents in the *ricinus communis* linn root extracts. Potential sources of novel drugs and therapeutic leads included the plant's active phytoconstituents. The study's findings indicated that the methanolic extract of *Ricinus communis* root includes compounds with anticancer properties and some pharmacological active moieties. Esters, bioflavonoids and phenolic compounds have been estimated by chemical analysis and these were further identified with the help of LC-MS analysis. Thus, the raw root extracts of *Ricinus communis* may serve as fresh sources for the creation of novel plant-based medicines for the treatment of various illnesses. Since the plant already contains phenolic compounds, esters, and flavonoids, and the potential toxicity of a few specific components were investigated. It can be employed for the approaching research because it has been found that it is less dangerous and active.

KEYWORDS: Esters, bioflavonoids and phenolic compounds, LC-MS analysis, computational screening and biochemically evaluation.

INTRODUCTION

People from all around the world have been using herbal medicine to enhance medical regimens for millennia. The growing prevalence of adverse medication responses and the emergence of microbial resistance to currently available antimicrobials have led to an exponential increase in the importance of plant-based medicines for treating diseases. Moreover, there is a great chance of obtaining new chemicals¹. Castor oil plant, scientifically named *Ricinus communis* Linn, is a type of flowering plant belonging to the *Euphorbiaceae* family. This soft-wooded

shrub grows widely in tropical climates². The roots have few rootlets, are practically smooth, and are light in weight. However, they do have longitudinal wrinkles³.

Because the plant's leaves, roots, and seed oil have been shown to be hepatoprotective, hypoglycemic, laxative, diuretic, and antibacterial, they have been utilized in Indian medicine to treat inflammation and liver problems. Roots can be used as a strong purgative and as a toothache paste when taken as a decoction or paste, respectively⁴. It has been found that distinct sections of *Ricinus communis* Linn. contain different phytoconstituents. The aerial portions of *Ricinus communis* contain alkaloids. The presence of ricinin (0.55%) and N Demethyl ricinin (0.016%), two alkaloids found in plant leaves⁵.

When compared to an aqueous extract, the methanolic extract of *Ricinus communis* Linn. shown greater anti-microbial activity. The dermatophytic and pathogenic bacterial strains *Streptococcus progenies*, *Staphylococcus aureus*, and *Klebsiella pneumonia* were all successfully combated by *Ricinus communis*. Using the well diffusion method, the various solvent extracts of *Ricinus communis* roots exhibited antibacterial efficacy against pathogenic microbes. For instance: *Proteus vulgaris*, *Salmonella typhimurium*, *Pseudomonas aeruginosa*, *Escherichia coli*, and *Staphylococcus aureus*.⁶ Because of its anti-allergic and mast cellstabilizing properties, *ricinus communis* root extract is useful in the treatment of asthma. Whereby flavonoids have bronchodilator and smooth muscle relaxant properties, and saponin has M. cell stabilizing properties⁷. Numerous investigations have demonstrated the antihistaminic properties of the *ricinus communis* root extract (ethanol/methanol).⁸

The lipid-lowering properties of the root and other portions of the *Ricinus communis* plant have been the subject of numerous investigations. According to findings, extracts from the roots of *Ricinus communis* may have hypolipidemic effects by lowering levels of triglycerides, lowdensity lipoprotein (LDL) cholesterol, and total cholesterol. An increase in high-density lipoprotein (HDL) cholesterol, which is thought to be advantageous for cardiovascular health, has also been observed in several studies⁹. Numerous chemicals found in the root of *Ricinus communis* have insecticidal effects. Among the most well-known substances is the poisonous protein called ricin, which is present in the seeds. But compared to the seeds, the roots have less ricin in them. In insects, ricin causes cell death by acting as a cytotoxin and interfering with the synthesis of proteins.¹⁰

Research has indicated that *Ricinus communis* root extracts can be useful in controlling a variety of insect pests. For instance, studies have shown that root extracts have insecticidal properties against leafhoppers, aphids, mosquitoes, and whiteflies. It has been demonstrated that these extracts cause death in the targeted insect species as well as feeding inhibition and decreased egg laying.¹¹

The antiepileptic potential of *Ricinus communis* methanolic root extract was studied in mouse models by Raju G.M. et al.¹². In order to investigate interactions with the active site utilizing PDB IDs 4MS4 (GABA agonists), 3WFG (glutamate antagonist), 5HCV (aspartate antagonist), and 6J8G (sodium channel antagonist), molecular docking studies were conducted by using Schrodinger software. Tounou et al.¹³ assessed the feasibility of employing crude extracts from *Ricinus communis* leaves, roots, and seed kernels along with oil emulsion as a means of managing the diamond back moth, *Plutella xylostella* (L.) (Lepidoptera: Plutellidae). Jena and Gupta¹⁴ proposed that the oil and the root of *Ricinus communis* were purgatives, effective against fever, rheumatism, inflammatory disorders, costiveness, and flatulence. The majority of compound medications used in rheumatic and neuralgic illnesses involve the root; seeds extracted from husks and germs and boiled in milk and water yield a decoction useful in rheumatism. According to Doshi A. K., et al.¹⁵, the root of both wild and farmed forms of *Ricinus communis* Linn; Euphorbiaceae, often known as Eranda in the Ayurvedic medical system, is used as an analgesic and anti-inflammatory medication. In Swiss albino mice,

gentisic acid (GA) was found to have protective benefits against cyclophosphamide (CP) induced hepatotoxicity and genotoxicity, according to Nafees S., *et al.*¹⁶. The anti-inflammatory and free radical scavenging properties of the methanolic extract of the root of *Ricinus communis* (RCM) (*Euphorbiaceae*) Linn. were investigated in Wistar albino rats by Ilavarasan R., *et al.*. The study's findings showed that in both acute and chronic inflammatory models in rats, the methanolic extract of *Ricinus communis* root exhibited strong anti-inflammatory action. The presence of phytochemicals with diverse biological activities, such as flavonoids, alkaloids, and tannins, in the plant extract could be the cause of the observed pharmacological activity¹⁷.

Compared to alternative analytical methods, LCMS has a number of benefits. Because mass spectrometric detection and chromatographic separation are combined, it also provides selectivity and specificity.¹⁸ Due to the molecular level of analysis; the main benefits of this technology are its sensitivity, specificity, and precision. It is also possible to decipher the analytes' structural features.¹⁹

The current study set out to biochemically analyze and computationally screen the root of *Ricinus communis* Linn. using methanol and water, the phytochemicals from the *ricinus communis* root were extracted. The more phytoconstituents in the *ricinus communis* linn root extracts were found by phytochemical screening utilizing LC-MS.

MATERIALS AND METHODS

This part deals with the material and methods used for extraction, phytochemical screening and LCMS analysis of extracts of root of *Ricinus communis*.

Collection and authentication of plant:

The roots of the plant *Ricinus communis* were acquired from an online herbal store named www.indianjadibooti.com and were authenticated from Dr. Gaurav Dubey, Department of Pharmacognosy, NIMS Institute of Pharmacy **Storage:**

Dried roots of *Ricinus communis* plant was preserved in tightly closed air tight containers and stored in a suitable cool and dry place.

Chemicals used:

All the chemicals used for the study were analytical grade purchased from R.S. Enterprises, Jaipur, Rajasthan.

Extraction¹⁸⁻²²:

Preparation of Methanolic Extract

At a temperature of $45 \pm 5^\circ\text{C}$, 20 gm of powdered *Ricinus communis* roots were extracted using 150 ml of methanol by a soxhlation process. This approach permits a continuous extraction process in order to achieve efficient extraction. Using a water bath, the produced organic extracts were dried and evaporated. This extraction method is cost-effective in terms of time, energy, and monetary outlays. To be used later, the extracts were stored in a refrigerator.

Preparation of Aqueous Extract

20 gm powdered material of roots of *Ricinus communis* were dried and extracted with 100 ml of water by soxhlation technique at a temperature of $60-80^\circ\text{C}$. The extracts were kept in a refrigerator for further use.

Preliminary Phytochemical Screening¹⁹⁻²⁴:

Phytochemical examinations were carried out for both the extracts of roots of *Ricinus communis* as per the standard methods as mentioned below.

Alkaloid:

Each extract was diluted in a little amount of hydrochloric acid before being filtered. To check for the presence of alkaloids, the filtrate was utilized.

Mayer's test:

Potassium mercuric iodide, the Mayer's reagent, was applied to the filtrates. Alkaloids were present when a yellow-colored precipitate formed.

Wagener's test:

Iodine in potassium iodide, Wagener's reagent, was applied to filtrates. When a brown or reddish precipitate formed, alkaloids were present.

Dragendorff's test:

Potassium bismuth iodide, the dragendorff's reagent, was applied to filtrates. The presence of alkaloids was shown by the formation of crimson precipitate.

Hager's test:

Saturated picric acid solution, Hager's reagent, was applied to the filtrates. The presence of alkaloids is verified by the precipitate's yellow coloration.

Flavonoids:

Alkaline reagent test: A few drops of sodium hydroxide solution were applied to the extracts. When diluted acid is added, the strong yellow color that formed before becoming colorless indicates the presence of flavonoids.

Lead acetate test:

A few drops of lead acetate solution were applied to the extracts. The presence of flavonoids was shown by the formation of a yellow-colored precipitate.

Tannins:

Ferric chloride test: In a test tube, 0.5 ml of extract and 10 ml of water were heated. After adding a few drops of 0.1% ferric chloride, the coloration was checked for brownish green or blue-black. This suggested that tannins were present.

Phenols

Test for Ferric Chloride:

Three to four drops of ferric chloride solution were applied to the extracts. The occurrence of bluish black coloration signified the existence of phenols.

Saponins:

Froth Test:

After diluting the extracts with 20 cc of distilled water, they were agitated for 15 minutes in a graduated cylinder. The presence of saponins was suggested by the formation of a 1 cm layer of foam. **Foam Test:**

Two milliliters of water were mixed with 0.5 grams of extract. Saponins were present if the foam generated lasted for 10 minutes or longer.

Glycosides:

Keller- Killiani test:

After boiling 0.5 ml of the extract in 5 ml of distilled water, 2 ml of glacial acetic acid with 1 drop of 0.5% ferric chloride solution was added. This was combined with 1 milliliter of sulfuric acid concentration. The presence of cardiac glycoside was suggested by the creation of a violet ring beneath the brown ring and a greenish ring slightly above the brown ring in the acetic layer, which subsequently spread throughout this layer.

Molisch's reagent test:

The extracts were combined thoroughly with two to three drops of the Molisch reagent. A few cautious drops of strong sulfuric acid were added to this. The presence of glycosides was shown by the formation of a reddish-purple colored ring at the intersection of two layers. **Terpenoids:**

Chloroform test:

After combining the extract with two milliliters of chloroform, three milliliters of concentrated H₂SO₄ were carefully added to create a layer. The interface takes on a reddish-brown coloration that indicates terpenoids are present and working well. **Carbohydrate: Fehling's reagent test:**

Two milliliters of the produced extract were placed in a sterile test tube, and then two milliliters each of Fehling's solutions A and B were added. For roughly ten minutes, the prepared solution was maintained in a bath of boiling water. The presence of carbohydrates was verified by the development of red precipitate.

Benedict's reagent test:

1 milliliter of the extract and 2 milliliters of Benedict's reagent were combined, and the mixture was cooked for 3 to 5 minutes in a boiling water bath. The presence of carbohydrates is confirmed by the formation of a brick-red-colored cuprous oxide precipitate.

Molisch's reagent test:

A tiny quantity of the extract needs to be combined thoroughly in a test tube with two to three drops of Molisch's reagent. To help a layer form and prevent mixing, a few drops of strong sulfuric acid were poured drop-wise along the test tube walls. An encouraging sign for carbohydrates is the emergence of a purple ring at the layer created by the concentrated acid.

Steroids:**Chloroform test:**

After dissolving 2 milliliters of the extract in 2 milliliters of chloroform and treating it with strong acetic and sulfuric acids, the extract developed a greenish hue, which suggested the presence of steroids.

Fixed oils:**Spot test:**

The test material was rubbed between the filter paper folds in this experiment. The emergence of a transparent area indicates that lipids are present in the sample.

Anthraquinones:**Bontrager's test:**

Ten minutes were spent for boiling of one gram of extract in five to ten milliliters of diluted HCl over a water bath. After filtering the mixture, an equal volume of ammonia solution and CCl₄ or benzene were added to the filtrate extract. Following shaking, the presence of the anthraquinone moiety was revealed by the emergence of pink to red color.

Quantification of Bioactive Compounds**Analysis of total phenolic content:^{20, 21}**

With minor adjustments, the literature approach was used to calculate the extracts' total phenolic contents. A calibration curve was created by combining a 1 mL ethanol solution of gallic acid (0.025-0.400 mg/mL), 5 mL of tenfold-diluted Folin-Ciocalteu reagent, and 4 mL of sodium carbonate (0.7 M). At 765 nm, absorbance data were recorded, and a standard curve was created. The aforesaid reagents were combined with 1 mL of *Ricinus communis* root extract (5 g/L), and the absorbance was measured after 30 minutes to ascertain the total phenolic contents. All determinations were carried out in triplicate. By comparison with a gallic acid (GA) standard, an equation was obtained from the standard curve of GA and used to determine the phenolic compounds in the extracts of roots of *Ricinus communis* (milligram of the GA equivalent, mg GAE).

The flavonoid content analysis²²

The colorimetric approach was utilized to ascertain the extracts' flavonoid concentration. To sum up, 30 minutes were spent at room temperature (RT) after 0.5 mL of the extract was combined with methanol, 10% aluminum chloride (AlCl₃), and 1 M potassium acetate. Utilizing a quercetin solution calibration curve and a spectrophotometer, the absorbance of the reaction mixture was determined at 415 nm. Micrograms of quercetin equivalent (QE) are used to represent the flavonoid content.

LCMS Analysis of the root extract of *R. communis*:

The Waters nano Acquity HSS T3, 1.8 μm, 100 μm × 100 mm column was used in the autosampler-equipped UHPLC system. 500 μL min⁻¹ was the flow rate for the mobile phases, which were water 0.1 % formic acid (A) and 90% acetonitrile in water with 0.1 % formic acid (B). Following a 15-minute maintenance period, the LC conditions were as follows: 5% during 0–3 min, a linear increase from 5% to 20% during 3–25 min, 20–40% during 25–40 min, and 40–50% during 40–55 min. A Thermo Electron LTQ-Orbitrap XL mass spectrometer equipped with a nano-electro-spray ion source (Thermo Fisher Scientific, Bremen, Germany) and operated under Xcalibur 2.1 version software, was used in positive ionization mode for the MS analysis using data-dependent automatic switching between MS and MS/MS acquisition modes.

RESULTS & DISCUSSION

Preliminary Phytochemical Analysis

The preliminary phytochemicals were identified in the methanolic and aqueous extract of roots of *R. communis* using various chemical tests. The presences of phytochemicals are shown in table 1.

Table 1: Preliminary Phytochemical Analysis of Methanolic & Aqueous Extract of *R. communis* roots

Phytochemicals	Test	Methanolic Extract		Aqueous Extract	
		Observation	Inference	Observation	Inference
Carbohydrates	Fehling's test	Brick red ppt	+	Brick red ppt	+
Flavonoids	Sulfuric acid test	Orange tored color	+	Orange tored color	+
Glycoside	Keller-Killani test	Reddish brown color appears	+	Reddish brown color appears	+
Saponin	Foam test	Froth appears	-	Thick persistent Foam observed	+
Tannin	5% FeCl ₃ test	Deep blue-black color	+	Deep blue-black color	+
Alkaloid	Dragendroff's test	Orange-brown ppt	+	No ppt formed	-
Steroid	Salkowski reaction test	Chloroform layer appear red and acid layer shows Greenish yellow color	+	No layer appeared	-

Protein	Million's test for protein	White ppt	+	No White ppt formed	-
Terpenoid	With acetic anhydride and sulphuric acid with chloroform	Deep red coloration	+	Orange coloration	-

+ Sign indicates presence and – sign indicates the absence of the phytochemical constituents. For aqueous extract (AE) and methanol (ME), the percentage yield was 5% and 6%, respectively. The methanolic extract of *R. communis* roots revealed the presence of carbohydrates, flavonoids, glycosides, tannins, alkaloids, steroids, protein, and terpenoids; however, saponin was not detected. The aqueous extract of *R. communis* roots was shown to include carbohydrates, flavonoids, glycosides, saponin, and tannins; however, alkaloids, steroids, protein, and terpenoids were found to be negatively associated with the extract.

Chemical Composition and Bioactive Compounds in RC

Bioactive compounds ²³

In terms of gallic acid equivalent, the total phenol levels were determined by the Folin-Ciocalteu reagent (standard curve equation: $y = 0.005x + 0.083$, $R^2 = 0.954$). Aqueous and methanolic extracts of *R. communis* roots showed a range of total phenols from 130 to 134 mg g⁻¹ and 46 to 49 mg/g, respectively. Aqueous and methanolic extracts of *R. communis* roots showed the presence of flavonoid and it was 9.22 mg quercetin dihydrate equivalent (QE)/g DW (AE) and 8.24 mg quercetin dihydrate equivalent (QE)/g DW (ME).

The reported values were compared with earlier reported data and results showed that the total phenolic content of the aqueous leaf extract of *Ricinus communis* L. was 48.39 mg gallic acid equivalent (GAE) per gram dry weight (DW). The flavonoid content was 9.76 mg quercetin dihydrate equivalent (QE)/g DW ^{24 & 25}. Whole phenolic concentration. Phenols are essential components of plants that are found in all plant species. The relationship between TPC and the extracts' overall antioxidant activity may be better understood with the use of the quantitative and qualitative analysis of the major phenolics in plants ²⁴. Individuals' mutagenesis and carcinogenesis are inhibited by polyphenolic compounds. TPC Chaharedhi *et al* ²⁵ were calculated using the Folin-Ciocalteu method, and the results were expressed as gallic acid equivalents (mg gallic acid per gram of dry mass). This approach was chosen for measuring phenolics due to its speed and intensity. Excessive levels of free radicals contribute to aging. They are the cause of many illnesses. The absence of antioxidant activity is the cause. Plant-based antioxidants are thought to be superior to synthetic antioxidants. Antioxidants with small molecular masses exist. They are thought to be effective in preventing oxidative stress ²⁶. The purpose of the *Ricinus communis* leaf extract was to analyze the total flavonoid content. The extracts of benzene, ethanol, and methanol were taken. A range of 17 to 28 µg/ml of catechin equivalent activity was observed ²⁷. Analysis was done on *Ricinus communis* leaf extracts. The aqueous extracts, methanol, and petroleum ether were utilized. They demonstrated 23-37 µg/ml of catechin comparable activity ²⁸. Both the aqueous and methanol exhibited 32 µg/ml. The activity of the remaining extracts was lower than this. It is thought that *Ricinus communis* is a useful medicinal plant. Its antioxidant activity is good. Gallic acid equivalents in aqueous were 131 mg/mL, n-butanol was 127 mg/mL, ethyl acetate was 117 mg/mL, and control was 155 mg/mL. Extracts from *Ricinus communis* roots were found to provide less than the control, but they nevertheless shown good equivalent activity ²².

Total phenolics (mg of GAE*/g DW**) = 130 to 134 mg g⁻¹ (AE) and 46 to 49 mg/g (ME).

Total flavonoids (mg of QE***/g DW**) = 9.22 (AE) AND 8.24 (ME).

*GAE: Gallic acid equivalent.

**DW: Dry weight.

***QE: Quercetin equivalent.

LCMS Analysis of Methanolic Extract

Figures 1 and 4 display the LC-MS chromatogram of the *Ricinus communis* methanolic extract, whereas figures 2 & 3 and 5-10 display the mass spectrum of the chemicals that were identified. It was noted that various retention times were used to generate the various peaks. By employing the molecular weight, the compounds are clarified using the standard reference graphs. Chromatogram of Methanolic extract of *R. Communis Linn* (Positive and negative ions) with the highest peak at that specific retention time and all the compounds are listed in tables 2 & 3, figures 1 & 4.

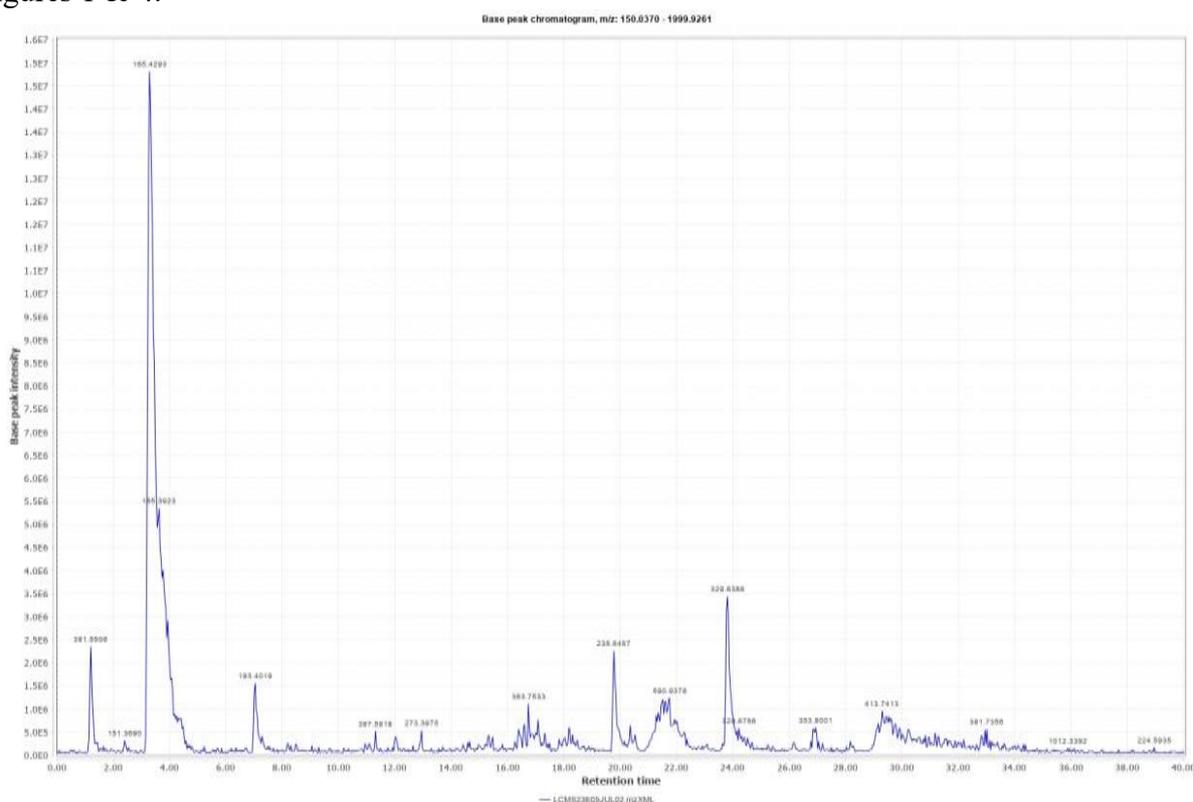


Figure 1: Chromatogram of Methanolic extract of *R. Communis Linn* (Positive ion)

Table 2: List of Compounds Reported from LCMS Analysis of Methanolic extract of *R. communis Linn* (Positive ion)

R. Time (min)	Score	Compound Name	Type of Ion	Formula	Exact Mass	Observed Mass	Mass Difference
1.21	0.731	Zeatin-9-glucoside	Positive	C ₁₆ H ₂₃ N ₅ O ₆	381.164	381.5506	-0.3866
3.77	0.909	D-Carnitine hydrochloride salt	Positive	C ₇ H ₁₅ NO ₃	161.105	165.4293	-4.3243
3.93	0.982	L-Carnitine	Positive	C ₇ H ₁₆ NO ₃	162.113	165.4293	-3.3163

4.08	0.987	Alpha-Methyl-DL-histidine dihydrochloride	Positive	C ₇ H ₁₁ N ₃ O ₂	169.085	165.4293	3.6557
7.05	0.988	Caffeine, Anhydrous	[M+H] ⁺	C ₈ H ₁₀ N ₄ O ₂	194.19	193.4019	0.7881
16.74	0.83	Guanosine-5'-monophosphate	Positive	C ₁₀ H ₁₄ N ₅ O ₈ P	363.057	363.7533	-0.6963
19.78	0.613	1-Isothiocyanato-8-(methylsulfinyl)octane	Positive	C ₁₀ H ₁₉ NO ₂ S ₂	233.09	235.5457	-2.4557
21.52	0.21	Uridine-5'-diphospho-Nacetylglucosamine sodium salt	Positive	C ₁₇ H ₂₇ N ₃ O ₁₇ P ₂	607.081	678.9996	-71.9186
23.77	0.634	Adenosine-3',5'-cyclic monophosphate	[M+H] ⁺	C ₁₀ H ₁₂ N ₅ O ₆ P	329.21	329.6016	-0.3916
23.80	0.606	Malvidin Chloride	Positive	C ₁₇ H ₁₅ O ₇	331.081	329.6386	1.4424
23.84	0.626	Malvidin Chloride	Positive	C ₁₇ H ₁₅ O ₇	331.081	329.6016	1.4794
29.30	0.608	Solasodine	Positive	C ₂₇ H ₄₃ NO ₂	413.329	413.7413	-0.4123

All the peaks of chromatogram (**fig. 1**) of methanolic extract of *R. Communis Linn* (Positive ion) were observed and the highest peak was found out at the retention time of 4.08 min for the compound of Alpha-Methyl-DL-histidine dihydrochloride with mass of 169.085 followed by RT at 1.21min, 19.78 min and 23.77min.

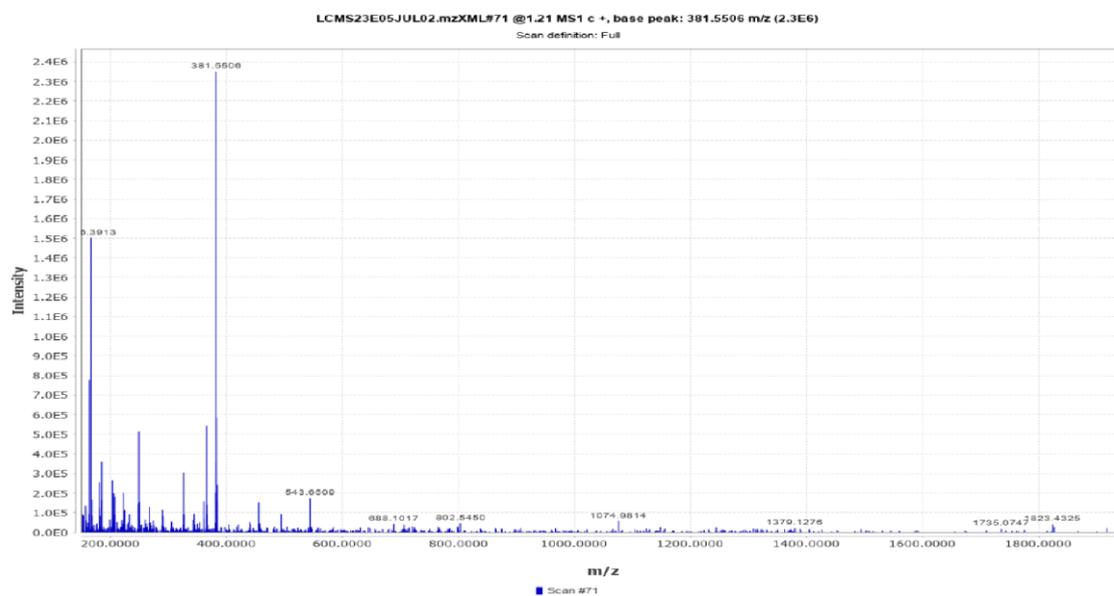


Figure 2: Mass spectrum at the retention time 1.21 (base peak 381.55m/z)

16.5 5	0.94 1	2'-Deoxyadenosine-5'monophosphate	negative	C ₁₀ H ₁₄ N ₅ O ₆ P	331.068	325.716 5	5.35
21.0 6	0.97 5	6-Phosphogluconic acid Barium salt hydrate	negative	C ₆ H ₁₃ O ₁₀ P	276.024	277.726 6	-1.7
22.6 3	0.91 6	D-Glucosamine-6-phosphate sodium salt	negative	C ₆ H ₁₄ NO ₈ P	259.045	253.639 1	5.41
23.3 1	0.97 5	6-Phosphogluconic acid Barium salt hydrate	negative	C ₆ H ₁₃ O ₁₀ P	276.024	279.724 6	-3.7
23.3 8	0.97 4	Gamma-Linolenic acid	[M-H]-	C ₁₈ H ₃₀ O ₂	278.43	279.761 6	-1.33
26.0 4	0.97 4	2'-Deoxyinosine	negative	C ₁₀ H ₁₂ N ₄ O ₄	252.085	255.674 1	-3.59
26.5 5	0.89 5	Luteolin	negative	C ₁₅ H ₁₀ O ₆	286.047	281.722 6	4.32
26.6 9	0.87 6	Gamma-Linolenic acid	[M-H]-	C ₁₈ H ₃₀ O ₂	278.43	281.796 6	-3.37
32.5 6	0.92 9	Acacetin	negative	C ₁₆ H ₁₂ O ₅	284.068	283.757 7	0.31

All the peaks of chromatogram (fig. 4) of methanolic extract of *R. Communis Linn* (negative ion) were observed and the highest peak was found out at the retention time of 26.55 min for the compound of Luteolin with mass of 281.722 followed by RT at 23.31min, and 23.38min.

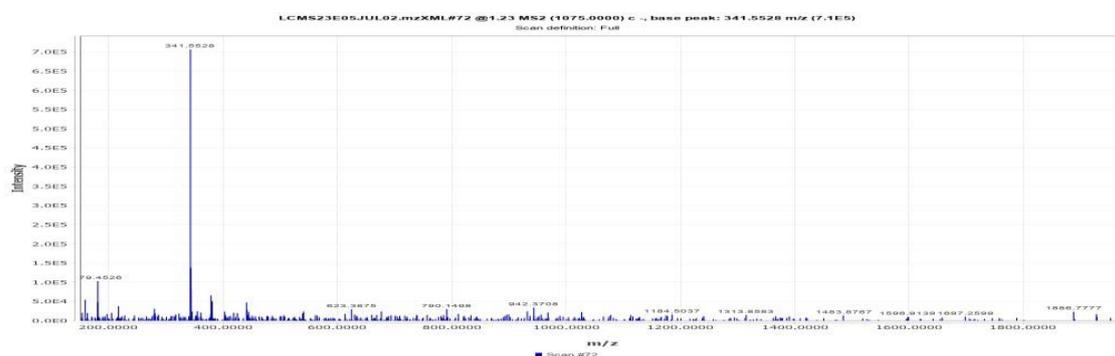


Figure 5: Mass spectrum at the retention time 1.23

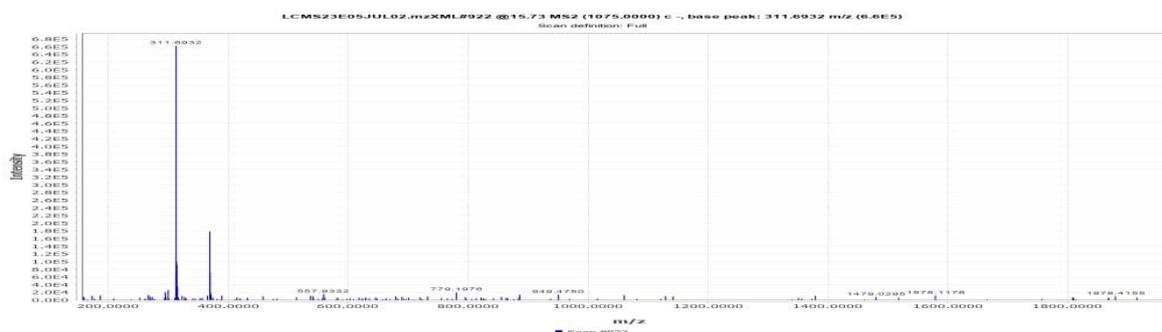


Figure 6: Mass spectrum at the retention time 15.73

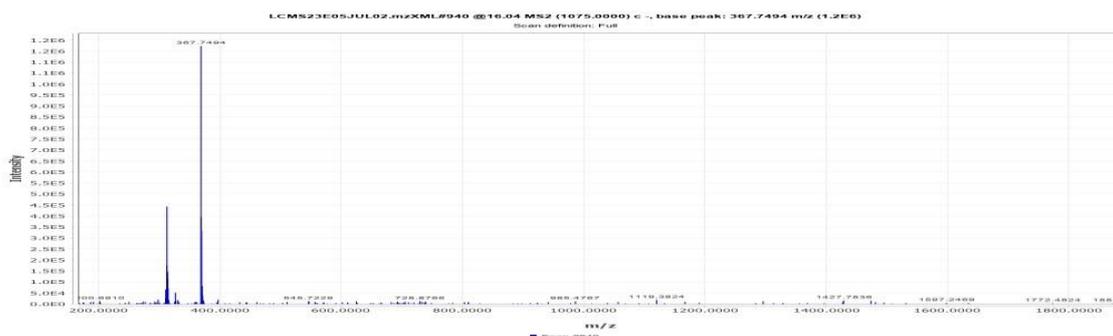


Figure 7: Mass spectrum at the retention time 16.04

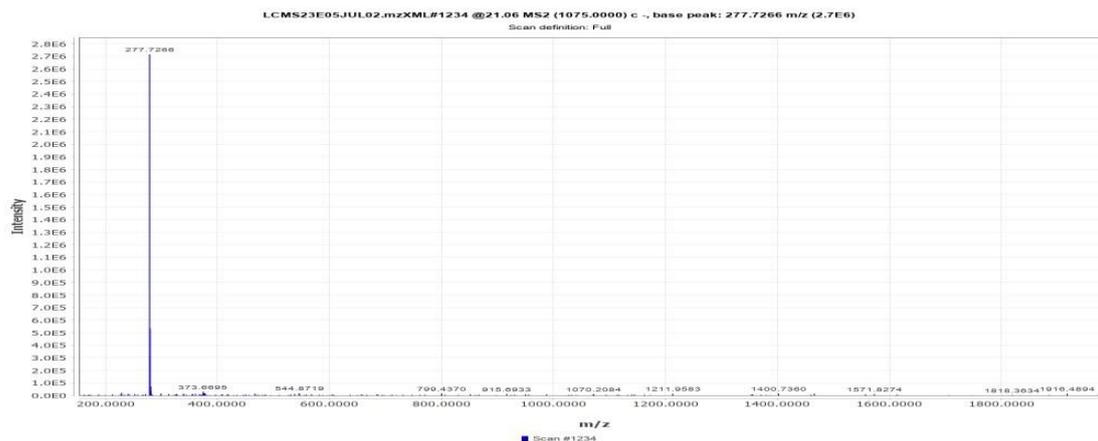


Figure 8: Mass spectrum at the retention time 21.06

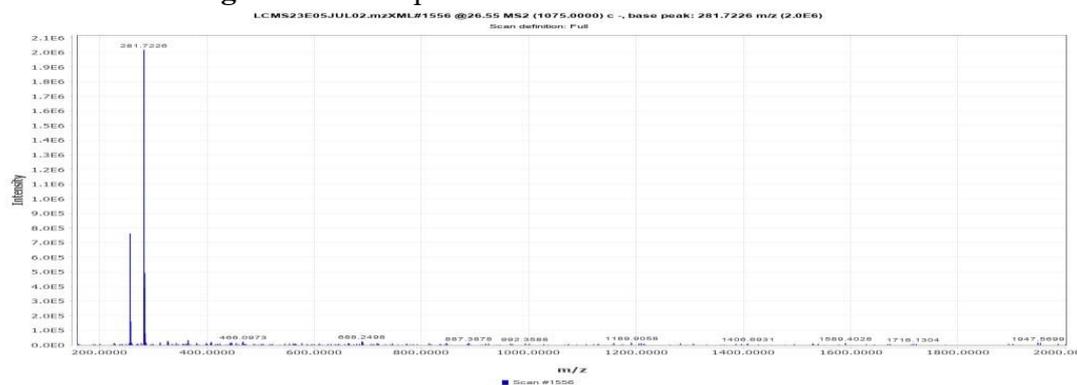


Figure 9: Mass spectrum at the retention time 26.55

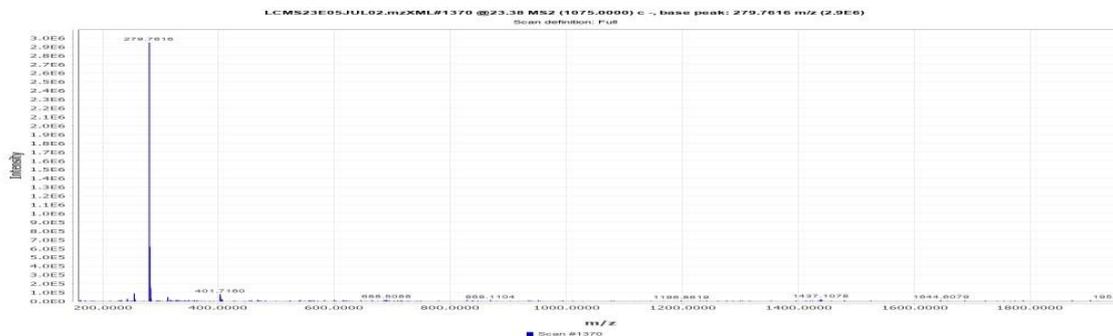


Figure 10: Mass spectrum at the retention time 26.69

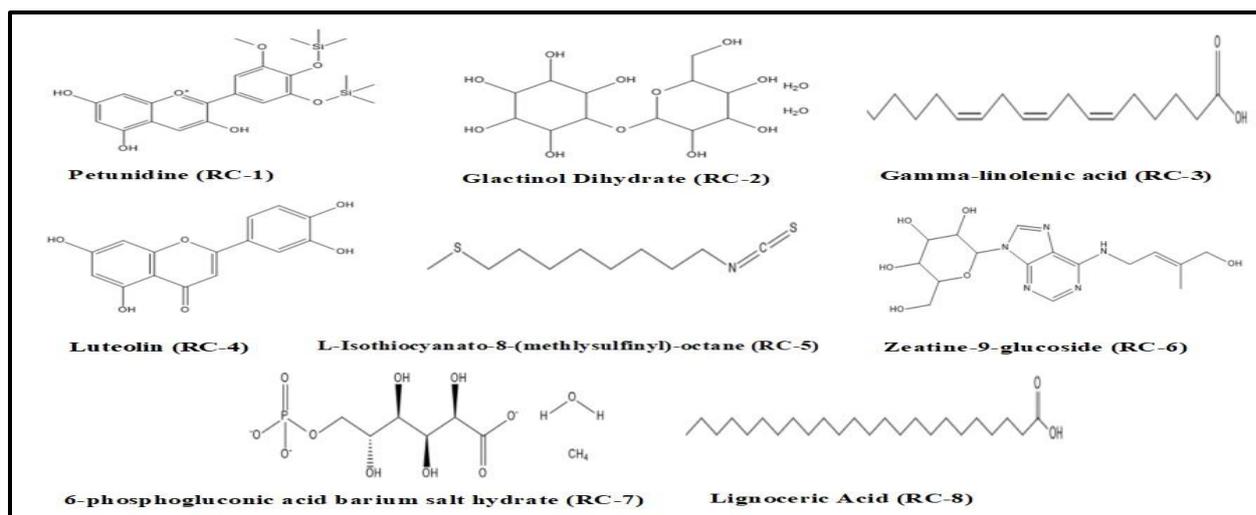


Figure 11: Structures of some isolated compounds

Since the plant already contains flavonoids, esters, and phenolic chemicals, we conducted toxicological investigations on a few selected components. We investigate the anticancer activity based on this. We discovered that the molecule is less hazardous and active; therefore we can use it in our future research.

Computational study of the following compounds

2D and 3D structure of computational study of the 2 compounds of Luteolin and Zeatin-9glucoside were conducted and it was mentioned in fig. 12 and 13.

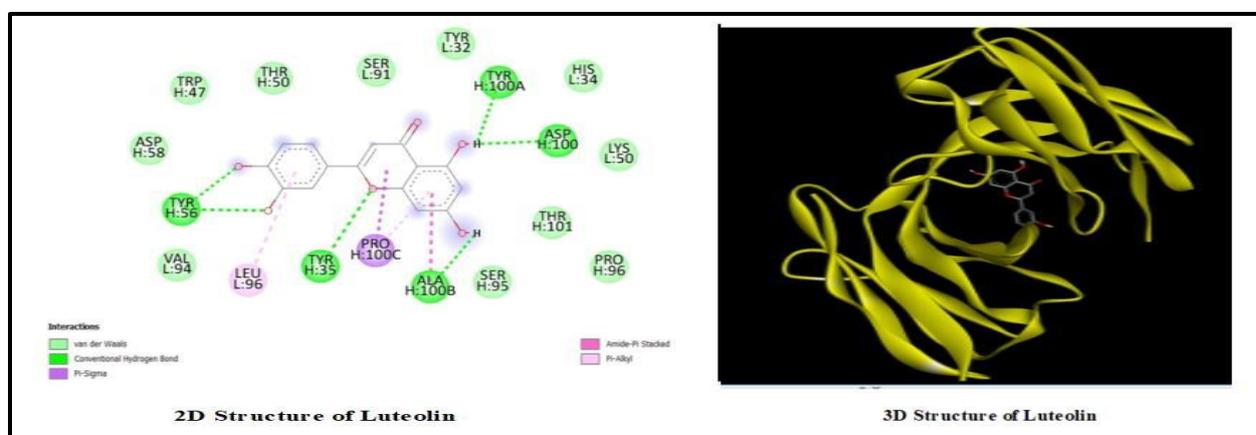


Figure 12: 2D Structure of Luteolin & 3D Structure of Luteolin

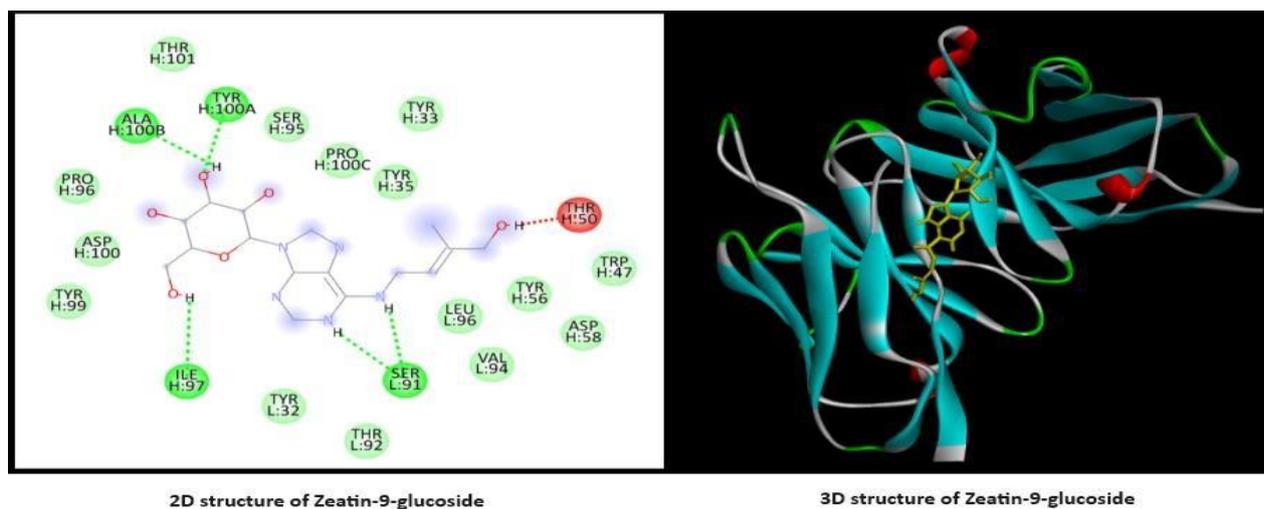


Figure 13: 2D Structure of Zeatin-9-glucoside & 3D Structure of Zeatin-9-glucoside

CONCLUSION

The plant's active phyto-constituents offered a potential source for novel medicinal and pharmaceutical possibilities. The study's findings demonstrated the presence of pharmacologically active compounds with anticancer potential in a methanolic extract of *Ricinus communis* root. The existence of esters, bioflavonoids, and phenolic compounds has been revealed by the LC-MS analysis. As a result, *Ricinus communis* root crude extracts may provide new opportunities for the development of novel plant-based therapies for the treatment of a variety of illnesses.

REFERENCES:

- Elkousy RH, Said ZN, Abd El-Baseer MA. Antiviral activity of castor oil plant (*Ricinus communis*) leaf extracts. *Journal of ethno-pharmacology*. 2021; 271:113878.
- Kumar, M., 2017. A review on phytochemical constituents and pharmacological activities of *Ricinus communis* L. *Plant. International Journal of Pharmacognosy and Phytochemical Research* 9 (4).
- Government of India, Ministry of Health and Family Welfare, Department of Ayush. The Ayurvedic Pharmacopeia of India 2007;Part- I: Vol. III: 34- 35.
- Hussain A, Aslam B, Muhammad F, Faisal MN. In vitro Antioxidant Activity and In vivo Antiinflammatory Effect of *Ricinus communis* (L.) and *Withania somnifera* (L.) Hydroalcoholic Extracts in Rats. *Brazilian Archives of Biology and Technology*. 2022 Jan 5;64.
- Kang S.S., Cordell A., Soejarto D.D., Fong H.H.S., "Alkaloids and flavonoids from *Ricinus communis*." *J. Nat. Prod.* 1985, 48 (1), 155–156.
- Abhishek mathur ; satish K. Verma, sajad yousuf, santosh K. Singh, Gbks prasad and V. K. Dua; Antimicrobial potential of roots of *riccinus communis* against pathogenic microorganisms; international journal of pharma and bio sciences, vol 2/issue 1/ janmar 2011.
- Dnyaneshwar J Taur et al. *Asian Pacific Journal of Tropical Biomedicine* (2011) S13-S16.
- Dnyaneshwar J. Taur; *Lat. Am. J. Pharm.* 30 (6): 1226-8 (2011).
- Matthew O. O., Olusola L., Matthew A. O., "Preliminary Study of Hypoglycaemic and Hypolipidemic Activity of Aqueous Root Extract of *Ricinus communis* in Alloxan-Induced Diabetic Rats." *J. Phys Pharm Adv*, 2012, 2(10): 354-359.

- Tounou A.K, Mawussi G, Amadou S, Agboka K, Gumedzoe Y. M. D and Sanda K., Bioinsecticidal effects of plant extracts and oil emulsions of *Ricinus communis* L. (Malpighiales: Euphorbiaceae) on the diamondback, *Plutella xylostella* L. (Lepidoptera: Plutellidae) under laboratory and semi-field conditions., *Journal of Applied Biosciences*; 43: 2899 – 2914; ISSN 1997–5902.
- Soto KM, Luzardo-Ocampo I, López-Romero JM, Mendoza S, Loarca-Piña G, Rivera-Muñoz EM, et al. Gold Nanoparticles Synthesized with Common Mullein (*Verbascum thapsus*) and Castor Bean (*Ricinus communis*) Ethanolic Extracts Displayed Anti-proliferative Effects and Induced Caspase 3 Activity in Human HT29 and SW480 Cancer Cells. *Pharmaceutics*. 2022 Sep 28;14(10):2069.
- Raju MG, Yadav EV, Reddy VNVLS, Nicholas M. Pharmacological and in silico Evaluations of methanolic flower extract of *Tagetes Patula* as anti-depressant and anxiolytic. *Bulletin Environmental Pharmacological Life Sciences*. 2021; 10(3):29-35.
- Tounou Agbéko Kodjo, Mawussi Gbénonchi, Amadou Sadate, Agboka Komi, Gumedzoe Yaovi Mawuena Dieudonné and Sanda Komla, Bio-insecticidal effects of plant extracts and oil emulsions of *Ricinus communis* L. (Malpighiales: Euphorbiaceae) on the diamondback, *Plutella xylostella* L. (Lepidoptera: Plutellidae) under laboratory and semifield conditions, *Journal of Applied Biosciences* 43: 2899 – 2914
- Jitendra Jena, Ashish Gupta, *Ricinus communis* linn: A phyto-pharmacological review, January 2012, *International Journal of Pharmacy and Pharmaceutical Sciences* 4(4):2529.
- Doshi Krunal A, Acharya Rabinarayan, Ravishankar B, Nariya Mukesh B, Anti-inflammatory activity of wild and cultivated varieties of eranda (*ricinus communis* linn.) Root, *Int. J. Ayur. Pharma Research*, 2014; 2(4): 107-113
- Sana Nafees, Shiekh Tanveer Ahmad, Wani Arjumand, Summya Rashid, Nemat Ali, Sarwat Sultana, Modulatory effects of gentisic acid against genotoxicity and hepatotoxicity induced by cyclophosphamide in Swiss albino mice, *J Pharm Pharmacol*, 2012 Feb;64(2):259-67. doi: 10.1111/j.2042-7158.2011.01393.x. Epub 2011 Dec 7.
- Raju Ilavarasan, Moni Mallika, Subramanian Venkataraman, Anti-inflammatory and free radical scavenging activity of *Ricinus communis* root extract, *J Ethnopharmacol*, . 2006 Feb 20; 103 (3):478-80. doi: 10.1016/j.jep.2005.07.029. Epub 2005 Nov 28.
- Want EJ, Cravatt BF, Siuzdak G. The Expanding Role of Mass Spectrometry in Metabolite Profiling and Characterization. *ChemBioChem*. 2005 Nov 4;6(11):1941–51.
- Stobiecki M, Kachlicki P, Wojakowska A, Marczak Ł. Application of LC/MS systems to structural characterization of flavonoid glycoconjugates. *Phytochemistry Letters*. 2015 Mar; 11: 358–67.
- Wolfe K, Wu X, Liu RH. Antioxidant activity of apple peels. *Journal of Agriculture and Food Chemistry*. 2003; 51: 609-614.
- Ojezele Matthew Obaineh, Abatan Matthew Oluwole and Onifade Abdulfatah Adekunle, Phytochemistry, Phenolic Contents and Free, Radical Scavenging of *Bauhinia thonningii* kurtz and *Ricinus communis* L Extracts as Possible Contribution to Medicinal Effects, *British Journal of Pharmaceutical Research(BJPR)*, 7(1): 9-15, 2015; Article no.BJPR.2015.086, DOI: 10.9734/BJPR/2015/13476
- Faheem Ahmed, Moshin Iqbal. Antioxidant activity of *Ricinus Communis*. *Organic & Medicinal Chem IJ*. 2018; 5(3): 555667. DOI: 10.19080/OMCIJ.2018.05.555667
- Jamshed Iqbal, Sumera Zaib, Umar Farooq, Afsar Khan, IrumBibi, and Saba Suleman, Antioxidant, Antimicrobial, and Free Radical Scavenging Potential of Aerial Parts of *Periploca aphylla* and *Ricinus communis*, *ISRN Pharmacology*, Volume 2012, Article ID 563267, 6 pages, doi:10.5402/2012/563267

- Demiray S, ME Pintado, PML Castro (2009) Evaluation of phenolic profiles and antioxidant activities of Turkish medicinal plants: *Tilia Argentena*, *Crataegi Folium* leaves and *Polygonum Bistorta* roots. *World Acad Sci Eng Technol* 3(6): 312-322.
- Chahardehi AM, D Ibrahim, SF Suleman (2009), Antioxidant activity and polyphenolic contents of some medicinal plants in Urticaceae family. *J Bio Sci* 3: 25-29.
- Hossain MA, MD Shah (2015) A study on total phenols content and antioxidant activity of essential oil and different solvent extracts of endemic plant *Merremia borneensis*, *Arab J Chem* 8(1): 66-71.
- Rao N, S Mittal, Sudhaanshu, E Menghaani (2013) Assessment of phytochemical screening, antioxidant and antibacterial potential of methanolic extracts of the *Ricinus communis*, *Asia J Pharm Tech* 3(1): 20-25.
- Chakarborthi GS (2008) Antioxidant activity of the successive extracts of *Ricinus communis* leaves. *J Envi Res Develop* 3(2): 537-539.