



## African Journal of Biological Sciences



### A REVIEW ON THE EVALUATION AND PHARMACOLOGICAL EFFECTS OF SWERTIA CHIRATA ON VARIOUS ANTIMICROBIAL RESISTANT STRAINS

1. \*Deepa Singh

\* Research Scholar, Lords University Alwar, India, 301028.

Assistant Professor, Biyani Institute of Pharmaceutical Sciences, Jaipur, India, 303706.

2. Dr. Nitin mittal

Professor, Department of Pharmaceutical Science, Lords University Alwar, India, 301028.

3. Dr. Mukesh Gupta

Professor, Department of Pharmaceutical Science, Lords University Alwar, India, 301028.

4. Dr. Shikha Sharma

Professor, Department of Pharmaceutical Science, Lords University Alwar, India, 301028

#### **ABSTRACT**

The global rise in antibiotic resistance has made it necessary to look for alternative treatments because it has compromised the efficacy of conventional antimicrobial therapy. While certain bacteria develop antibiotic resistance spontaneously, antibiotic overuse and the emergence of novel resistant forms through mutation are the main causes of antibiotic resistance in other bacteria. Plants have long been the primary source of medications and complementary therapies for the treatment of illnesses. Valuable secondary metabolites, including flavonoids, terpenoids, quinones, tannins, alkaloids, and polyphenols, are abundant in plants. Plant secondary metabolites are the subject of numerous investigations as a possible source for the development of antibiotics. They can act through a variety of processes and possess the necessary structural qualities. In addition to examining phytochemicals from various classes that have been shown to have antimicrobial activity against resistant bacteria, either on their own or in conjunction with conventional antibiotics, this review examines the antibiotic resistance mechanisms developed by multidrug-resistant bacteria. The current investigation sought to ascertain *Swertiachirata*'s antibacterial efficacy against a number of antibiotic-resistant bacterial pathogens, including *Escherichia coli*, *Pseudomonas aeruginosa*, *Bacillus* species, and *Staphylococcus aureus*. At a Minimum Inhibitory Concentration (MIC), *Swertiachirata* demonstrated strong antimicrobial efficacy against a range of antimicrobial resistant bacteria. It might therefore be evolved into a superb broad-spectrum antibacterial agent.

To sum up, the results are encouraging and indicate that *Swertia chirata* may be useful in treating a range of microbial infections.

**Keywords:** Antimicrobial resistance, *Swertiachirata*, medicinal plant, MIC

Article History

Volume 6, Issue 5, 2024

Received: 01 May 2024

Accepted: 09 May 2024

doi:10.33472/AFJBS.6.5.2024.3274-3287

## 1. INTRODUCTION

A significant increase in the frequency of bacterial infections that cause antibiotic-associated multidrug and antimicrobial resistance has occurred in the last few decades. The misuse, abuse, and overuse of antimicrobial medications are the main reasons why bacteria develop resistance genes. A resistance gene may be acquired by the bacterium as a result of horizontal gene transfer or bacterial DNA changes, in addition to these other factors, which could result in antibiotic resistance (WHO, 2018). Antimicrobial resistance is a complex and significant issue that impacts global health. To combat it, a multidisciplinary approach including partners from all health sectors, including the scientific community and public health authorities, is required. Resistant infections are now the third most common cause of mortality worldwide, according to a 2016 WHO report. Current figures show that antimicrobial resistance (AMR) costs the European healthcare system approximately EUR 1.5 billion annually and results in over 25,000 fatalities each year in Europe (European Commission, 2017). AMR has made it more difficult to manage major procedures, such as caesarean sections, cancer chemotherapy, organ transplants, and diabetes (WHO, 2018). According to estimates, drug-resistant diseases would kill 10 million more people annually than cancer by 2050. If appropriate measures are not taken to address this threat, the world economy would suffer a \$100 trillion USD loss.

In February 2017, the World Health Organization (WHO) published a list of bacteria that have been connected to human diseases. The list made it clear how urgently new antibiotics are needed to tackle infections that are hard to treat. In healthcare facilities that have been designated as critical for antimicrobial resistance (AMR), the pathogens *Acinetobacter baumannii*, *Pseudomonas aeruginosa*, carbapenem-resistant Enterobacteriaceae, and Methicillin-resistant *Staphylococcus aureus* (methicillin-resistant) are particularly dangerous. Antimicrobial resistance (AMR) was the subject of a UK parliament debate in July 2014, during which the prime minister announced an extensive examination of AMR in an effort to identify new treatment approaches. It has also been taken into account in the "UK Five Year Antimicrobial Resistance Strategy 2013 to 2018" (Department of Health, 2013). It is essential to find alternative drugs to treat infectious infections.

Especially in the past ten years, more thorough studies on natural remedies have been conducted. It has been shown that plants are an excellent source of natural substances that support human health. Currently, phytochemicals are being continuously used for medicinal purposes in many countries. According to the World Health Organization (WHO), medicinal plants are the best source of a broad variety of drugs. The use of crude extracts of plant components and phytochemicals with proven antibacterial properties may be crucial in medical therapy. Many plants have been used for their antibacterial qualities, which are derived from the secondary metabolites the plants produce. The active components in these products, which include phenolic and tannin compounds present in essential oils, are widely known. Plants generate a wide variety of secondary metabolites, which are used as direct precursors or lead chemicals in the pharmaceutical industry. Plant extracts that target locations other than antibiotics are expected to be useful against drug-resistant bacteria.

As secondary metabolites, bioactive molecules are typically accumulated by all plant cells. However, the concentration of these substances varies depending on the part of the plant, the season, the climate, and the growth stage. The leaf is one of the plant parts that has the highest concentration of these compounds, which is why most people utilize it medicinally. A few of the active components stop the growth of disease-causing microorganisms, either alone or in combination. The most important of these plant bioactive substances include flavonoids, phenolic compounds, tannins, and alkaloids. Comprehending the correlation between phytoconstituents and plant bioactivity is imperative for the synthesis of molecules possessing specific activities intended for the management of diverse ailments, encompassing chronic conditions. Plants must undergo this form of preliminary phytochemical screening to

discover and produce novel therapeutic compounds with greater efficacy, considering the significance of the previously described context. Other research groups throughout the world have also reported on these types of investigations. Therefore, the current study investigates the antibacterial activity of medicinal plants, particularly *Swertiachirata*, against gram-negative *Escherichia coli* and gram-positive *Staphylococcus aureus* bacteria.

## **1.2 Mechanism of Microbial Resistance**

Numerous theories have been proposed to explain bacterial resistance to conventional antimicrobial treatments, and around 20,000 resistance genes have been found in bacteria. The 1950s saw the discovery of the first antibiotic resistance involving *E. coli*, *Shigella*, and *Salmonella* species. It took two decades to identify this growing issue, as multiple examples of penicillin, tetracycline, and chloramphenicol were documented in the 1970s. Clinical trials were rarely conducted to evaluate the hypothesized causes of bacterial resistance. It is unclear whether multiple bacteria share a common pathway to generate resistance or if each microbe has its own mechanism.

While certain bacteria may develop antibiotic resistance spontaneously, antibiotic overuse and the emergence of novel resistant strains are the main ways in which bacteria gain antibiotic resistance. Antibacterial resistance can be caused by a variety of mechanisms, such as bacterial efflux pumps that speed up the release of antibiotics, changes in the amount of time it takes for medication to diffuse inside bacteria, structural changes in bacterial porins that reduce permeability to antibiotic influx, hydrolytic enzymes that break down antibacterial agents, and modifications to antibiotic binding sites. Bacteria can develop resistance to antibiotics by combining multiple drugs or by resisting just one. In order to determine potential targets for future effective medicine, it is imperative to comprehend the processes of resistance.

## **1.3 Botanical description of genus *Swertia***

The Dutch gardener Emanuel Sweert (1552–1612; also written Swert) is honored by the name *Swertia*. These plants belong to the genus of herbs that are typically annual, biennial, or perennial and range in height from 2.5 cm to 1.5 m. Roots of *Swertia* species can be fibrous or woody. Stems might be terete, striate, angled, winged, simple, or occasionally branched, scapiform, well-developed, ascending, or erect. Most leaves are sessile, petiolate, or opposite; occasionally, they are rosulate, whorled, or alternating. Every *Swertia* L. species has a complete leaf edge. The inflorescences of these plants resemble cymes, and they usually form paniculate or simple thyrses. They can sometimes be solitary or raceme-like, although they are rarely absolutely dichotomous. A pair of sessile, opposing, leaf-like bracts serves as the primary support for pedicellate, four- or five-merous blooms. The calyx and corolla rotate, tubes less than 3 mm in diameter and lobed bases. Each corolla lobe has one or two nectaries, which can be naked, coated in scales or flaps, fringed, fimbriate, or glabrous. The number of corolla lobes is equal to the number of stamens that are attached at the base of the sinuses surrounding the corolla lobes and occasionally ringed by long hairs. The ovary has styles that range in duration from brief to long. The fruit is a flattened or oval capsule with a persistent calyx and corolla that split into two valves; the stigma is divided into two halves. Typically, *Swertia* L. seeds range in size from a few to numerous.

## **1.4 Effect of *Swertiachirata* on antimicrobial-resistant micro-organism**

The chemistry of natural and synthetic medicinal products is very different, with natural products having a wider range of chemical constituents. Natural products are higher in

molecular complexity, scaffold variation, stereochemistry, ring system diversity, and carbohydrate content, and lower in nitrogen, phosphorus, sulfur, and halogen (Schmidt et al., 2008). Natural plant extracts provide a variety of diverse phytochemicals that prevent the development of resistance. Plant extracts can combat diseases through numerous pathways due to the availability of multiple phytochemicals in a single plant. It has been claimed that *Swertiachirata* works well against a variety of infections. *Swertiachirata* contains natural plant extracts and chemicals that, in addition to having several modes of action, can be used in conjunction with conventional antibiotics to boost their antibacterial efficacy. The antibacterial efficacy of the *Swertia* species that are presently being studied varies depending on the type of bacterial strain. As stated by Srinivasan and colleagues (2001). This activity might indicate that *Swertia* species either widespread metabolic poisons or contain compounds that function as broad-spectrum antibiotics. Phytochemical analysis has demonstrated the presence of steroids, flavonoids, tannins, and phenols in *Swertia* species. has demonstrated antimicrobial properties of tannins and flavonoids. The antibacterial activity against certain bacterial strains could be attributed to these phytochemicals. Similar results were obtained from a previous study that looked at various *Swertia* species. Numerous studies have reported the antibacterial activity of different *Swertia* species against various microbial strains, including *Swertiachirata* (Sultana et al., 2007; Ahirwal et al., 2011; Kweera et al., 2011; Ghosh et al., 2012; Roy et al., 2015), *S. ciliata* (Saeed et al., 1998), *S. corymbosa* (Ramesh et al., 2002; Kweera et al., 2015), and *S. Petiolata* (Bader, 2014).

### **1.5 Evaluation and Pharmacological activity of *Swertiachirata***

Numerous pharmacological studies have been initiated as a result of the diverse ethnobotanical applications of *Swertiachirata*. According to earlier studies, the biological activities of the *Swertiachirata* extracts include antibacterial, antifungal, antiviral, anticancer, anti-inflammatory, and other properties like antidiabetic and antioxidant properties (Verma et al., 2008). The pharmacological characteristics of *Swertiachirata* have been assessed using a wide variety of in vitro and in vivo test systems. Evidence-based laboratory research has demonstrated some intriguing pharmacological effects of *Swertiachirata* extracts in aqueous, alcoholic, and methanolic forms. For its antibacterial and antifungal properties, the entire *Swertiachirata* plant has been observed to be utilized. The anti-hepatitis B viral activity of *Swertiachirata* extracts was studied using HepG cell lines.

The entire *Swertiachirata* plant has been reported to possess hypoglycemic and anti-inflammatory properties. Chen et al. (2011) investigated the antioxidant activity of a 70% ethanolic extract of *Swertiachirata* using antioxidant tests such as the beta-carotene assay and reducing power. The results indicated that extracts containing 70% ethanol had a significant DPPH scavenging effect ( $IC_{50} = 267.80 \mu\text{g/mL}$ ).

Evaluation of the biological activities of <i>Swertiachirata</i> .						
Bioactivity evaluated,	Plant part(s) tested,	Test system,	Extracting solvent,	Test Organism/Mo dels,	Resistance for drug,	Toxicity test.
Antibacteria 1	Whole plant	<i>In vitro</i>	Ethanol	<i>Escherichia coli</i> ATCC 26922	Ciprofloxacin	None
				<i>Klebsiella pneumonia</i> ATCC 15380		
				<i>Pseudomonas aeruginosa</i> ATCC 25619		
				<i>Proteus vulgaris</i> ATCC 6380		
Anti-bacterial	Stem	<i>In vitro</i>	Methanol	<i>Bacillus subtilis</i> ATCC 6633	Ceftriaxone, Ceftriaxone sodium, Cefuroxime, Ciprofloxacin, Gentamycine, Levofloxacin, Metronidazole, andTranexamic acid	None
				<i>Enterococcus faecalis</i> (ATCC 14506)		
				<i>Staphylococcus aureus</i> (ATCC 6538)		
				<i>Pseudomonas aeruginosa</i> (ATCC 27853)		
Antibacteria 1	Whole plant	<i>In vitro</i>	Methanol	<i>Bacillus subtilis</i> MTCC 736	Gentamycin	None
				<i>Bacillus polymyxa</i>		
				<i>Staphylococcus aureus</i> MTCC 3160		

				<i>Escherichia coli</i> MTCC 723		
				<i>Salmonella typhi</i> MTCC 3216		
				<i>Vibria cholera</i> MTCC 3906		
				<i>Streptococcus pyogenes</i> MTC C 1927		
				<i>Proteus mirabilis</i> MTC C 1429		
				<i>Providentiaalk alifaciens</i>		
				<i>Pseudomonas aeruginosa</i> MT CC 7837		
Antibacteria 1	Whole plant	<i>In vitro</i>	DCM; Ethanol	<i>Staphylococcus aureus</i>	Kanamycin 30 µg/disc	None
Antibacteria 1	Stem	<i>In vitro</i>	Ethanol	<i>Staphylococcus aureus</i>	Chloramphenicol 30 µg/disc	Brine shrimp assay–positive
				<i>Bacillus subtilis</i>		
				<i>Salmonella typhi</i>		
				<i>Shigella flexeneriae</i>		
				<i>Sarcina lutea</i>		
				<i>Bacillus megaterium</i>		
Antifungal	Whole plant	<i>In vitro</i>	Methanol	<i>Aspegillusniger</i> MTCC 1881	Amphotericin	None
				<i>Aspergillus flavus</i> MTCC 1883,		
				<i>Cladosporium oxysporum</i> MT CC 1777		

Swertiachirata presents a plethora of promising opportunities for both traditional and contemporary treatments. *Swertiachirata* seems to have a wide range of applications as a herbal medicine. An overview of the current use of *Swertiachirata* in ethnobotany, phytochemistry, pharmacological characteristics, safety evaluation, and human conservation status is given in this article. However, as no serious adverse effects or toxicity have been reported to date, more toxicological research is needed. To further substantiate the safety of these diverse plant-derived compounds. Toxicological, mutagenic, and biological activity *in vivo* attributes need to be assessed. Clinical trials are most likely required to determine the effectiveness of employing *Swertiachirata* in medicine. Due to its many applications, demand is always rising on both domestic and foreign markets. The population has drastically decreased as a result of overexploitation and habitat loss. Any proposed research has to be considered in a larger context that encompasses sustainable raw plant supply and conservation methods in order for this critically endangered medicinal plant to be successfully commercialized. Innovative methods that make use of biotechnological interventions—such as cryopreservation, micropropagation, and bioreactors—will be needed for both conservation and increasing commercial production. In order to enable commercial use, more comprehensive research is required in the field of synthetic seed technology, mainly to enhance the frequency of synthetic seed germination and the subsequent growth in soil. Hairy root technology has the potential to be used as a model system in the near future and will give plant biotechnologists powerful tools to improve *Swertiachirata's* advantageous phytochemicals. Despite the development of efficient micropropagation methods, more studies on the biology of seeds and methods to increase the amount of bioactive secondary metabolites in *Swertiachirata* cultures would be helpful for their commercialization. Furthermore, to prevent *Swertiachirata* from being misidentified and perhaps adulterated, quality control methods must be followed.

## 1.6 Biological and pharmacological screening

There are various methods designed to investigate the biological activity of a targeted natural compounds. An optimal procedure must fulfil several criteria: fast, simple, reliable, high sensibility and selectivity, availability and low cost. Bioactivity evaluation for a plant extraction (plant fraction) is usually performed through *in vitro* or/and *in vivo* studies. Most often, *in vitro* studies are focused on the evaluation of specific cell biology (cell count, growth rate, metabolic rate, cell function and protein expression). *In vitro* tests are conducted on various animal or human cell cultures, enzymes, depending on targeted natural compound biological activity. For instance, the bioassays for antitumor activity are conducted on tumor experimental models. Complementary, the immunological activity on normal cell culture should be monitored. The cells will be analyzed by fluorescence microscopy and will be quantified to establish the degree of apoptosis and implicitly the cell viability. Also, the time lapse video microscopy can be used to evaluate the bioactive phytochemicals. The *in vivo* bio tests are applied on animals (mice, rats, pigs, etc.).

### Antimicrobial Resistance (AMR)

Antimicrobial Resistance (AMR) occurs when microorganisms develop resistance to antimicrobial medications such as antivirals and antibiotics. AMR is increasingly becoming a global threat, both economically as well as in terms of health. AMR can lead to the treatments becoming ineffective, making it more difficult to treat infections, and can then cause an increase in the risk of the disease spreading. Particularly, diseases such as tuberculosis, malaria, HIV and Influenza are developing more drug resistant cases, leading to higher healthcare costs and a longer duration of illness. Within the European Union alone, the average yearly additional costs caused by AMR between 2015 and 2030 are estimated to be \$252,215 USD per 100,000 persons per year.

## Factors causing AMR

Several factors have been found to cause AMR. Human use of antimicrobial drugs, particularly antibiotics, is recognized as one of the primary drivers of AMR.

- Antibiotics are among the most familiar of medicines by the public, and the global consumption of antibiotics has been estimated at more than 35 billion daily doses at 2015.
- Several studies have found a positive association between antimicrobial use and AMR.
- Although in the past AMR research has focused on direct public consumption of antimicrobial drugs, drugs use in livestock are an increasingly concerning issue.
- Antibiotics are an integral part of industrial agriculture to ensure healthy livestock and promotion of growth.
- The Food and Agriculture Organization (FAO) expects that two-third of the future growth of antimicrobial use will be linked to animal production. Human consumption of these animals can lead to resistant microorganism transmission between hosts. This can then in turn impact the wider environment through animal and human waste affecting soil and land.

## Other factors causing AMR

- Over-prescription of antibiotics
- Patients not finishing the entire antibiotic course
- Overuse of antibiotics in livestock and fish farming
- Poor infection control in health care settings
- Absence of new antibiotics being discovered

Table 1: Medicinal plants used for the treatment of antimicrobial disease

Sr No.	Botanical Name	Family	Part Used
1	<i>Acacia nilotica</i>	Mimosaceae	Stem
2	<i>Achyranthes aspera</i>	Amaranthaceae	Leaves
3	<i>Acorus calamus</i>	Araceae	Rhizome
4	<i>Aegelemarmelos</i>	Rutaceae	Leaves
5	<i>Aervalanata</i>	Amaranthaceae	WP
6	<i>Ageratum conyzoides</i>	Asteraceae	Leaves



7	<i>Alangiumsalvifolium</i>	Alangiaceae	Fruit
8	<i>Andrographis alata</i>	Acanthaceae	Leaves
9	<i>Andrographis echioides</i>	Acanthaceae	Leaves
10	<i>Andrographis paniculata</i>	Acanthaceae	Leaves
11	<i>Andrographis serpyllifolia</i>	Acanthaceae	Leaves
12	<i>Annona squamosa</i>	Annonaceae	Leaves
13	<i>Aristolochia bracteolate</i>	Aristolochiaceae	Leaves
14	<i>Azadirachtaindica</i>	Meliaceae	Leaves
15	<i>Calotropis gigantean</i>	Asclepiadaceae	Latex
16	<i>Carissa carandas</i>	Apocynaceae	Leaves
17	<i>Curcuma longa</i>	Zingiberaceae	Rhizome
18	<i>Cynodandactylon</i>	Poaceae	Root
19	<i>Euphorbia hirta</i>	Euphorbiaceae	Latex, Leaves
20	<i>Justicia adhotada</i>	Acanthaceae	Leaves
21	<i>Leucas aspera</i>	Lamiaceae	Leaves
22	<i>Mangiferaindica</i>	Anacardiaceae	Bark
23	<i>Mimusopselengi</i>	Sapotaceae	Leaves
24	<i>Plectranthuscoleoides</i>	Lamiaceae	Leaves
25	<i>Psidium guajava</i>	Myrtaceae	Leaves
26	<i>Scantalum album</i>	Santalaceae	Stem
27	<i>Sesbania grandiflora</i>	Fabaceae	Leaves
28	<i>Solanum surattense</i>	Solanaceae	Fruit
29	<i>Sphaeranthus indicus</i>	Asteraceae	Seed
30	<i>Tribulus terrestris</i>	Zygophyllaceae	Whole plant
31	<i>Tridaxprocumbens</i>	Asteraceae	Leaves
32	<i>Vitex negundo</i>	Verbenaceae	Leaves
33	<i>Zingiberofficinale</i>	Zingiberaceae	Rhizome [86-145]

## **CONCLUSION**

The dramatic rise in infections in recent years has led to a chronic issue of antibiotic resistance. The antibacterial efficacy of different *Swertia* species against various bacterial strains varies, according to current research. As stated by Srinivasan and colleagues (2001). This activity might indicate that *Swertia* species either general metabolic poisons or contain broad-spectrum antibiotic chemicals. Phytochemical analysis has demonstrated the presence

of steroids, flavonoids, tannins, and phenols in *Swertia* species. has demonstrated the antimicrobial properties of tannins and flavonoids. The antibacterial activity against certain bacterial strains could be attributed to these phytochemicals. To sum up, a lot of study has been done on *Swertiachirata* in the fields of phytochemistry, biological activity, ethnobotany, taxonomy, and conservation. However, new findings, may increase the therapeutic utility of *Swertiachirata* today and promote its continued use in modern medicine. More biotechnological approaches are required for conservation efforts.

## **REFERENCE**

1. Coates A (2002), Hu Y, Bax R, and Page C. " The future challenges facing the Development of new antimicrobial drugs. Nature Reviews,1: 895-910.
2. L.C. Braga (2005), J.W. Shupp, C. Cummings, M. Jett, J.A. Takahashi, L.S. Carmo Pomegranate extract inhibits *Staphylococcus aureus* growth and subsequent enterotoxin production J. Ethnopharmacol., 96, pp. 335-339
3. Marjorie M.C.(1999) Plant Products as Antimicrobial Agents. Clin. Microbiol. Rev. 12(4):564–582.
4. N. Tomoko (2002), A. Takashi, T. Hiromu, I. Yuka, M. Hiroko, I. Munekazu, et al.(2002) Antibacterial activity of extracts prepared from tropical and subtropical plants on methicillin resistant *Staphylococcus aureus* J Health Sci, 48, pp. 273-276
5. Costa Es (2008), Hiruman – Lima CA, Lima EO, Sucupira GC, Bertolin AO, Loils SF, Andrade FDA, an Vilegas w, Souza- Brito ARM, antimicrobial activity of some medicinal plants of the Cerrado, Brazil." Phytotherapy Research (2008); 22: 705-707.
6. Ateb DA (2003), Erdourul T "Antimicrobial Activities of Various Medicinal and commercial plant extracts. Turkish Journal of Biology 27: 157-162.
7. Nair R. Chanda S (2006) "Activity of some medicinal plants against certain pathogenic bacterial strains. Indian Journal of Pharmacology 38: 142-144.
8. Martinez MJ (1996), Betancourt J, Alonso-Gonzalez N, Jauregui A Screening of some Cuban Medicinal Plants for antimicrobial activity. Journal of Ethnopharmacology 52: 171-174.
9. Nair R. (2007a), Vaghasiya Y, Chanda S. Antibacterial Potency of selected Indian Medicinal Plants. International Journal of Green Pharmacy 1: 37-44.
10. Ndhlala AR (2009), Stafford GI, Finnie JF, Van Staden J "In vitro pharmacological effects of manufactured herbal concoctions used in KwaZulu-Natal, South Africa." Journal of Ethnopharmacology 122: 117-122.
11. Hasler, C. M and J. B. Blumberg. (1999). Symposium on Phytochemicals, Biochemistry and Physiology." Journal of Nutrition, 129: 756 - 757.
12. Lewis K, Ausubel FM (2006) "Prospects for Plant-Derived Antimicrobials. Nat Biotechnol 24:1504–1507.
13. Bonjar S (2004). Evaluation of Antibacterial Properties of some Medicinal Plants used in Iran. J. Ethnopharmacological 94: 301-305.
14. Islam B (2008), Khan SN, Haque I, Alam M, Mushfiq M, Khan AU. Novel Anti-adherence Activity of Mulberry Leaves: Inhibition of *Streptococcus mutans* Biofilms by 1 Deoxynojirimycin isolated from *Morus alba*. J. Antimicrobial Chemother. 62(4): 751-757.
15. Nair R, Chanda S (2007). In vitro antimicrobial activity of *Psidium guajava* L. leaf extracts against clinically important pathogenic microbial strain. Braz. J. Micro. 38: 452-458.
16. Alam K. D. (2009), Ali M. S., Parvin S., Mahjabeen S., Akbar M. A., Ahamed R. In vitro antimicrobial activities of different fractions of *Swertiachirata* ethanolic

- extract. Pak. J. Biol. Sci. 12, 1334–1337. 10.3923/pjbs.1334.1337.
17. Gonzalez C E (1996), Venzon D, Lee S, Mueller B U, Pizzo P A, Walsh T J. Risk factors for fungemia in children infected with human immunodeficiency virus: a case-control study. *Clin Infect Dis* ;23:515–521.
  18. Dean D A, Burchard K W.(1996). Fungal infections in surgical patients. *Am J Surg.* ;171:374–382.
  19. Ng PC. (1994) Systemic fungal infections in neonates. *Arch Dis of Childhood.*;71:F130–F135.
  20. Norrby SR, Nord CE, Finch R.(2005). Lack of development of new antimicrobial drugs: a potential serious threat to public health. *Lancet Infect Dis.* 5:115–119.
  21. Sharma R. (2005), Verma U., Sharma C.L., Kapoor B. Self-medication among urban population of Jammu city. *Indian J. Pharmacol.*37:37–45.
  22. Hussain MA and Gorski MS, (2004) Antimicrobial Activity of *Nerium oleander* Linn. *Asian Journal of Plant Sciences*, 3: 177-180.
  23. Iwu M. W. (1999), Duncan A.R., Okunji C.O. In: *Perspectives on New Crops and New Uses*. Janick A.S., editor. ASHS Press; Alexandria, VA. New antimicrobials of plant origin; pp. 457–462.
  24. Akinyemi KO (2005), Oladapo O, Okwara CE, Ibe CC, Fasura KA “Screening of crude extracts of six medicinal plants used in South – West Nigerian unorthodox medicine for antimethicillin-resistant *S. aureus* activity,” *BMC Comp. Alt. Med*, vol 5(6), pp. 1- 7.
  25. Doughari JH.(2006). Antimicrobial Activity of *Tamarindus indica* Linn. *Tropical J Pharma Res.*5:597–603.
  26. Zaïke LL, (1975). species and herbs: their antimicrobial activity and its determination. *Journal of Food Safety*; 9: 97-118.
  27. ROBBERS, J.M., SPEEDIE and TYLER, V. (1996). *Pharmacognosy and Pharmacobiotechnology*. Williams and Wilkins, Baltimore. P. 1 – 14.
  28. Kumar V. (2014), Singh S. K., Bandopadhyay R., Sharma M. M., Chandra S. In vitro organogenesis secondary metabolite production and heavy metal analysis in *Swertiachirayita*. *Cent. Eur. J. Biol.* 9, 686–698.
  29. Ara H. (2000), Jaiswal U., Jaiswal V. S. Synthetic seed: prospects and limitations. *Curr. Sci.* 78, 1438–1444.
  30. Gantait S. (2015), Kundu S., Ali N., Sahu N. C. Synthetic seed production of medicinal plants: a review on influence of explants, encapsulation agent and matrix. *Acta Physiol. Plant* 37, 98.
  31. Perveen S., Anis M. (2014). Encapsulation of internode regenerated adventitious shoot buds of Indian *Siris* in alginate beads for temporary storage and twofold clonal plant production. *Acta Physiol. Plant.* 36, 2067–2077.
  32. Ara H. (2000), Jaiswal U., Jaiswal V. S. Synthetic seed: prospects and limitations. *Curr. Sci.* 78, 1438–1444.
  33. Gantait S. (2015), Kundu S., Ali N., Sahu N. C. Synthetic seed production of medicinal plants: a review on influence of explants, encapsulation agent and matrix. *Acta Physiol. Plant* 37, 98
  34. Kumar V. (2014), Singh S. K., Bandopadhyay R., Sharma M. M., Chandra S. In vitro organogenesis secondary metabolite production and heavy metal analysis in *Swertiachirayita*. *Cent. Eur. J. Biol.* 9, 686–698.
  35. Sharma N. (2013a), Varshney V. K., Kala R. P., Bisht B., Sharma M. Antioxidant capacity and total phenolic content of *Swertiachirayita* (Roxb. ex Fleming) H. Karst. in Uttarakhand. *Int. J. Pharm. Sci. Rev. Res.* 23, 259–261.
  36. Kumar V. (2014), Singh S. K., Bandopadhyay R., Sharma M. M., and Chandra S. In vitro organogenesis, secondary metabolite production, and heavy metal analysis in *Swertia*.
  37. Cassini A (2019), Högberg LD, Plachouras D, et al. Attributable deaths and disability-

- adjusted life-years caused by infections with antibiotic-resistant bacteria in the EU and the European Economic Area in 2015: a population-level modelling analysis. *Lancet Infect Dis* 19:56–66.
38. O'Neill J. (2016) Tackling drug-resistant infections globally: final report and recommendations. HM Government and Wellcome Trust: UK.
  39. Klein EY (2018), Van Boeckel TP, Martinez EM, et al. Global increase and geographic convergence in antibiotic resistance. *PNAS*. 3463–70.
  40. Costelloe C (2010), Metcalfe C, Lovering A, et al. Effect of antibiotic prescribing in primary care on antimicrobial resistance in individual patients: systematic review and meta-analysis. *BMJ*. 340:c2096.
  41. Mladenovic-Antic S (2016), Kocic B, Velickovic-Radovanovic R, et al. Correlation between antimicrobial consumption and antimicrobial resistance of *Pseudomonas aeruginosa* in a hospital setting: a 10-year study. *J Clin Pharm*. 532–537.
  42. Silbergeld EK (2008), Graham J, Price LB. Industrial food animal production, antimicrobial resistance and human health. *Annu Rev Public Health* 151–69.
  43. Food and Agriculture Organization. Antimicrobial resistance. 2020.
  44. Vijay Kumar (2016), Johannes Van Staden. Review of *Swertia chirayita* (Gentianaceae) as a traditional medicinal plant; *frontiers in Pharmacology*, Volume 6, article 308.
  45. Abdul Aleem (2018), Hifzul Kabir. Review on *Swertia chirata* as traditional uses to its phytochemistry and pharmacological activity, *Journal of Drug Delivery & Therapeutics*. 73-78.
  46. Bhatt, A. (2006), Rawal, R. S., and Dhar, U. Ecological features of a critically rare medicinal plant, *Swertia chirayita*, in Himalaya. *Plant Species Biol*. 49–52.
  47. Schimmer, O. (1996), and Mauthner, H. Polymethoxylated xanthenes from the herb of *Centaurea erythraea* with strong antimutagenic properties in *Salmonella typhimurium*. *Planta Med*. 62, 561–564.
  48. Timsina, B. (2018), Kindlmann, P., Rokaya, M. B., Vrchotová, N., Tříška, J., Horník, Š. & Šýkora, J. Xanthenes Content in *Swertia multicaulis* D. Don from Nepal. *Molecules*, 23(5), 1067.
  49. Ghosh, R. (2012), Rahman, M., Faruki, Z., Rabbee, M.C., Ghosh, D. R. & Hasan, S. M. Phytochemicals and Antimicrobial Behavior of Plant *Swertia chirayita*, A Medicinal Plant from Rangpur, Bangladesh. *International Research Journal of Pharmaceutical and Applied Sciences*, 2(6), 183-186.
  50. Khan, S. W. & Khatoon, S. (2008). Ethnobotanical studies on some useful herbs of Haramosh and Bugrote valleys in Gilgit, northern areas of Pakistan. *Pakistan Journal of Botany*, 40(1), 43-58.
  51. Lv, P. (2010), Wei, L., Du, Y., Yang, H. & Peng, M. Hepatoprotective and toxic characteristics of the whole herb of traditional Tibetan folk medicine *Swertia mussotii* Franch. *Journal of Medicinal Plants Research*, 4(8), 706-709.
  52. Jamwal, A. (2012). Systematic review on xanthenes and other isolates from genus *Swertia*. *International Journal of Pharmaceutical, Chemical and Biological Sciences*, 1, 1464-1482.
  53. Pant, N., Jain, D. C. & Bhakuni, R. S. (2000). Phytochemicals from genus *Swertia* and their biological activities. *Indian Journal of Chemistry*. 39(B), 565-586.
  54. Kumar, V. & Van Staden, J. (2016). A review of *Swertia chirayita* (Gentianaceae) as a traditional medicinal plant. *Frontiers in Pharmacology*, 6, 308.

55. Subedi, I. & Karki, T. B. (2018). Phytochemical and Antimicrobial Screening of native plant Swertiachirayita (Roxb. ex Fleming) karst from Rasuwa district of Nepal. *Journal of Tropical Life Science*, 8(2), 166-171.
56. Mahendran, G. (2015), Manoj, M., Prasad, K. J. R. & Bai, V. N. (2015). Antioxidants, antiproliferative, anti-inflammatory, anti-diabetic and anti-microbial effects of isolated compounds from Swertiacorymbosa (Grieb.) Wight ex CB Clark–An in vitro approach. *Food Science and Human Wellness*, 4(4), 169-179.
57. Samaddar, T. (2013), Chaubey, B., Jha, S. & Jha, T. B. (2013). Determination of swertiamarin and amarogentin content and evaluation of antibacterial activity in Eastern Himalayan species of Swertia L. *Pharmacognosy Communications*, 3(4), 1-7.
58. Colombo, M. L. & Bosisio, E. (1996). Pharmacological activities of Chelidoniummajus L. (Papaveraceae). *Pharmacological Research*, 33(2), 127-134.
59. Khan, L. U. (2017), Khan, R. A., Khan, S., Bano, S. A., Fasim, F. & Uzair, B. Phytochemical screening and assessment of pharmacological properties of Swertiachirayita (Roxb. ex Fleming) root methanolic extract. *International Journal of Pharmacology*, 13(8), 1000-1009.
60. Khanal, S. (2015), Shakya, N., Thapa, K. & Pant, D. R. Phytochemical investigation of crude methanol extracts of different species of Swertia from Nepal. *BMC Research Notes*, 8(1), 1-9.
61. Das, J. (2013), Thapa, S., Pradhan, D., Thorat, S. S. & Talukdar, N. C. Intra-specific genetic diversity, phytochemical analysis and antioxidant activities of a potential Himalayan Swertia (Swertiabimaculata Hook. F. & Thomas.). *Industrial Crops and Products*, 49, 341-347.
62. Phoboo, S. (2010), Bhowmik, P. C., Jha, P. K. & Shetty, K. Anti-diabetic potential of crude extracts of medicinal plants used as substitutes for Swertiachirayita using in vitro assays. *Botanica Orientalis: Journal of Plant Science*, 7, 48-55.
63. Phoboo, S. (2012), Pinto, M. D. S., Barbosa, A. C. L., Sarkar, D., Bhowmik, P. C., Jha, P. K. & Shetty, K. Phenolic- Linked Biochemical Rationale for the Anti- Diabetic Properties of Swertiachirayita (Roxb. ex Flem.) Karst. *Phytotherapy Research*, 27(2), 227-235.
64. Niki, E. (2010). Assessment of antioxidant capacity in vitro and in vivo. *Free Radical Biology and Medicine*, 49(4), 503-515.
65. Nandkarni K. M. (1976). *Indian Materia Medica*, Bombay Popular Prakashan, Vol. I. Bombay: Elsevier; 1184–1186.
66. Kirtikar, KR. and B.D. Basu, (1984) “ *Indian Medicinal Plants* ”Latit Mohan Basu, Leader Road, Allahabad: India, pp.1664-1665.
67. Bhattacharya SK (1976), Reddy PKSP, Ghosal S, Singh AK, Sharma PV “Chemical constituents of gentianaceae XIX: CNS-depressant effects of swertiamarin ”*J. Pharm. Sci.*, Vol .65, pp.1547-1549.
68. Garg DS, (editor-in chief) Dhanvantri-BanaushdhiVisheshAnk., (1965) ,DhanvantriKaryalaya,Vijaygarh, Aligarh :India, Vol.3, p.94.
69. Sharma PV, Dravyaguna-vijnana, (1986) Chaukhambha Bharti Academy, Varanasi:India, Vol. 2.
70. Ahirwal L. (2011), Singh S and Mehta A. Antimicrobial screening of methanol and aqueous extracts of Swertiachirata. *Int J Pharm Pharm Sci*, 3(4):142-146.
71. Khatri, S. (2019). Taxonomy of Genus Swertia L. (Gentianaceae) in Nepal. M. Sc. Dissertation, Central Department of Botany, Tribhuvan University, Kirtipur.
72. Rajbhandari, K. R. (2015), Bhatt, G. D., Chhetri, R. & Rai, S. K. Catalogue of Nepalese flowering plants -supplement-1. Ministry of Forest and Soil Conservation, Department of Plant Resources, National Herbarium and Plant Laboratories, Lalitpur, Nepal.
73. Chandra, D. (2016), Kohli, G., Punetha, V. D., Prasad, K., Bisht, G., Khetwal, K. S. & Pandey, H. K. Phytochemical and ethnomedicinal uses of family Gentianaceae. *Current*

- Research in Chemistry, 8(1-3), 1-9.
74. Rijal, D. P. (2009). Taxonomic study of some medicinally important species of *Swertia* L. (Gentianaceae) in Nepal. *Botanica Orientalis: Journal of Plant Science*, 6, 18- 24.
  75. Li Z, Cheng X, (2005) Wang CJ, Li GL, Xia SZ, Wei FH. Purification of the effective component from *Solanum xanthocarpum* and its effect against *Oncomelania* snails. *Zhongguo Ji ShengChong Xue Yu Ji Sheng Chong Bing Za Zhi* 23, 206-208.
  76. Linda, R. (2002), *Whole Herbs or Standardized Plant Constituents?* San Antonio, Texas. Throug 2012-18.
  77. Madaan R, (2010) Singh B, Kumar S. Pharmacognostic standardization of *Conium maculatum*. *J Pharm Biomed Sci*, 1, 1-5.
  78. Madaus G, (1999). *Lehrbuch der BiologischenHeilmittel*, Band II. Georg Thieme, Leipzig, pp. 1681–1684.
  79. Verma, H. (2008), Patil, P. R., Kolhapure, R. M., and Gopalkrishna, V. Antiviral activity ofthe Indian medicinal plant extract, *Swertiachirata* against herpes simplex viruses: a study byin-vitroand molecular approach.*Indian J. Med.Microbiol.* 26, 322–326.
  80. Pant,N.  
(2000),Jain,D.C.,andBhakuni,R.S.Phytochemicalsfromgenus*Swertia*andtheirbiological activities.*Indian J. Chem.* 39,565–586.
  81. Jiahui Lu1, (2020) Anita Sheldenkar2\* and May Oo Lwin. A decade of antimicrobial resistanceresearch in social science felds: a scientometric review, *Antimicrob Resist Infect Control*9:178, 4-13.
  82. MariyaObanovska, (2017) Giulia Pila. *Penicillin’s Discovery andAntibiotic Resistance: Yale J Biol. Med.*mar,90(1):135-145.