

<https://doi.org/10.48047/AFJBS.6.15.2024.4478-4502>



African Journal of Biological Sciences

Journal homepage: <http://www.afjbs.com>



Research Paper

Open Access

Novel strategies for *Salmonellosis* risk reduction in poultry.

Saima Ansari¹, Vaishali Nirmalkar², Rasika Pawar^{1*}

¹Department of Microbiology, Smt. Chandibai Himathmal Mansukhani College, Ulhasnagar, District: Thane- 421003, Maharashtra, India.

²Department of Botany, K.M.E Society's G.M. Momin Women's College, Bhiwandi, District: Thane- 421302, Maharashtra, India.

*Department of Microbiology, Smt. Chandibai Himathmal Mansukhani College, Ulhasnagar, District: Thane- 421003, Maharashtra, India. Phone: + 91- 9869118328.

E-mail: rasikapawarchm@gmail.com

Volume 6, Issue 15, Sep 2024

Received: 15 July 2024

Accepted: 25 Aug 2024

Published: 05 Sep 2024

doi: [10.48047/AFJBS.6.15.2024.4478-4502](https://doi.org/10.48047/AFJBS.6.15.2024.4478-4502)

ABSTRACT:

Poultry farming is one of the fastest-growing sectors in India. Poultry immunity, health, and production are the variables that challenge the future growth of the poultry industry. Consumer confidence, product quality and safety, types of products, and the emergence and re-emergence of diseases will continue to be significant challenges to the current situation and the strategic future of the industry. Poultry farming faces substantial challenges related to maintaining high levels of immunity and health among birds, directly impacting production and consumer confidence. The industry must address the emergence and re-emergence of diseases to ensure the quality and safety of poultry products for consumers. Foodborne and zoonotic diseases are strictly linked with poultry. Eradication, elimination, or control of foodborne and zoonotic pathogens is extremely important in the poultry industry. *Salmonellosis* is a common foodborne disease affecting humans and animals. *Salmonella's* incidence in poultry significantly impacts the economy and growth of poultry farming. In this article, we shed light on the importance of exploring alternative interventions, such as probiotics, to combat antibiotic-resistant *Salmonella* infections in poultry, enhancing bird health and consumer safety. Implementing strategic measures to reduce *Salmonella* prevalence improves public health outcomes and contributes to the overall success and resilience of the poultry industry in India. These interventions are primarily used to control antibiotic-resistant *Salmonella* colonization in poultry food. Effective control of foodborne and zoonotic pathogens like *Salmonella* is crucial for the economic sustainability and growth of poultry farming in India.

Keywords: Chicken, *Salmonella*, Probiotics, Poultry.

INTRODUCTION

Ensuring the quality and safety of animal products is a crucial focus for the poultry industry, shaping its strategic future. Disease control, high productivity, high-quality products, and low prices are the goals of current poultry farming. (Hafiz and Atiya, 2020). However, animal-derived bacteria (Sweeney et al., 2018), primarily transmitted via food or via direct contact with animals, have become problematic (Marshall & Levy, 2011). Some food-borne bacteria, such as *Salmonella typhimurium* or *Escherichia coli*, isolated from chicken intestines are threat to food industry (Castellanos et al., 2017).

In 1885, American scientist Daniel E. Salmon isolated intestinal bacteria from the intestines of pigs. In 1900, French bacteriologist Joseph Leon Marcel Lignères proposed naming cholera “*Salmonella*” in honor of Daniel Salmon (El-Saadany et al., 2022). *Salmonella* belongs to the Enterobacteriaceae family. *Salmonella* is a Gram-negative, facultative anaerobic, rod-shaped, non-spore-forming bacterium (Gut et al., 2018). *Salmonella* is catalase-positive and oxidase, negative. It hydrolyzes urea using citrate as the sole carbon source (Vijaya et al., 2018). Contaminated water, soil, and wastewater are sources of *Salmonella* (Li et al., 2013). Animal products contaminated with *Salmonella* are responsible for 3% of foodborne illnesses worldwide, infecting 80 million people and killing 155,000 humans (Abd El-Hack et al., 2021a). Non-typhoidal *Salmonella*, *Salmonella enterica* serovar *Typhimurium*, is commonly found in food source all over the world (Havelaar et al., 2015) and has the most significant impact on human health compared to other contaminated foods (Kirk et al., 2015 Scallan et al., 2015). A recent study by the World Health Organization's Foodborne Disease Burden Epidemiology Reference Group also found *Salmonella enterica* serovar *Typhi* to be the pathogen responsible for most deaths from foodborne illness (Havelaar et al., 2015). A recent literature study investigated the characteristics of some probiotic *Enterococcus* strains (Laukova et al., 2008a; Laukova et al., 2008b) and the production of bacteriocins of these strains thought to prevent *Salmonellosis* (Simonova & Laukova, 2007). Poultry birds do not necessarily get infected by eating food contaminated with *Salmonella*. However, humans get infected by consuming contaminated chicken carcasses with *Salmonella* (Wibisono et al., 2020). This article focuses on the risk of *Salmonellosis* in poultry and the use of probiotics in its treatment.

Indian Scenario

Salmonellosis in India is a prevalent disease affecting humans and animals. *Salmonellosis* in poultry is not endemic but is on the rise and significantly impacts the economy and growth of poultry farming. The incidence of *Salmonellosis* in India is high but has not yet been fully reported due to the limited number of laboratories monitoring the birds (Rajagopal & Meaney, 2013).

Classification of *Salmonella* species

The genus *Salmonella* is split into two species, *Salmonella enterica*, and *Salmonella bongori*, based on differences in temporal analysis. The intestinal *Salmonella* group is divided into six subspecies: *Salmonella enterica subsp. enterica*, *Salmonella enterica subsp. Salamae*, *Salmonella enterica subsp. Arizona*, *Salmonella enterica subsp. Diarizonae*, *Salmonella enterica subsp. Houtenae* and *Salmonella enterica subsp. Indica* (Lan et al. 2009). Kauffman and White's phylogenetic-based subspecies classification provides additional concepts for classifying *Salmonella* into serotypes based on the significant antigenic determinants: somatic (O), capsule (K), and flagella (H). Heat-stable somatic O-antigen is an oligosaccharide component of lipopolysaccharide found in bacterial membranes. Heat-labile H antigens are found on bacterial flagella. Surface K antigen is a heat-sensitive polysaccharide found on the surface of the bacterial capsule and is the most common antigen for *Salmonella* serotypes (S.K.Eng et al., 2015). *Salmonella* species specific to avian hosts include *S. gallinarum* and *S. pullorum*. *Salmonella* infection is usually transmitted from chickens via the fecal-oral route by ingesting contaminated food or water (Ferrari et al., 2019). Contamination occurs in two ways: direct contact with animals and the environment or indirect contact via food (e.g., milk, eggs, and meat) with contaminated products (e.g., stainless steel, milk jugs, and knives) (Kebede et al., 2022).

Diagnosis

Water and feed samples can be collected and tested for the presence of *Salmonella*. Sterile Cotton swabs samples are collected from laying cages, breeder nests, or hatchery machines. Rapid *Salmonella* detection techniques including Widal tests, enzyme immunoassays,

antigen capture assays, DNA probes, and immunofluorescence are routinely used for detection. (www.poultrydvm.com)

***Salmonella* antibiotic resistance**

Certain bacterial species, like lactic acid bacteria, carry resistance genes and might serve as reservoirs of resistant genes for enteropathogens. Namely, tetracycline, vancomycin, and erythromycin resistance genes have been identified. Alternatives like prebiotics, probiotics, and essential amino acids (Abou- (Kassem et al., 2021a; Arif et al., 2021; Alagawany et al., 2021a), polyphenols, biologically synthesized nanoparticles, organic acids, essential oils, (Saad et al., 2021a,b), bioactive peptides (El-Saadony et al., 2021a,b; Saad et al., 2021c), herbal extracts (Abou-Kassem et al., 2021b; El-Saadony et al., 2021c), enzymes (Llamas-Moya et al., 2019), bioactive plant compounds (El-Saadony et al., 2021d; Reda et al., 2021a), and phytochemical compounds (Abdelnour et al., 2020a,b; Ashour et al., 2020, 2021; Abd Elkader et al., 2021; Abdel Moneim et al., 2021; Abd El Hack et al., 2021c,d) are used to improve poultry performance and human health using safe and natural products. Probiotics are generally derived from lactic acid bacteria isolated from fermented dairy products, sausages, and raw meat products, including poultry, beef, and pork (V.T. Nair et al., 2018). Antibiotic-resistant *Salmonella* cause the contamination of the farm environment and water systems (H.Y. Done et al., 2015). The use of fecal waste as manure in agricultural lands can be a reason for the spread of antibiotic resistance species of *Salmonella*. Antibiotic-resistant foodborne pathogens, namely *Salmonella*, *E. coli*, and *Shigella*, have often been recovered from fresh products that are available locally (S. Liu et al., 2017). Using pesticides causes soil contamination with livestock feces, and the spraying or irrigation of contaminated water causes the spread of resistant bacteria to fruits, vegetables, and fresh produce (V.T. Nair et al., 2018).

Antibiotic Alternatives against *Salmonella*

Many interventions have targeted antibiotic-resistant *Salmonella*. However, it is understood that the interventions used against non-antibiotic-resistant bacteria could work equally well against resistant *Salmonella*. Here are a few interventions that could attack antibiotic-resistant *Salmonella*. *Salmonella* is mainly treated with antibiotics, including ciprofloxacin, ceftriaxone, and ampicillin (Obaidat & Stringer, 2019). This antibiotic resistance is

responsible for the failed treatment of *Salmonella* in clinics, resulting in high mortality and morbidity. Overuse of antibiotics is also associated with gut dysbiosis and induces other disorders, such as inflammatory bowel disease or allergies (Schulfer et al., 2018). Consequently, probiotics have been identified as promising solutions in *Salmonella's* preventive and therapeutic treatment. Nowadays, the world is directed to limit the usage of antibiotics. Hence use of probiotics is considered safe as they have many benefits for humans and animals (Hill et al., 2014). Many probiotic strains also inhibit *Salmonellosis* (Adetoye et al., 2018; Pradhan et al., 2019).

Direct-Fed Microbials

The use of probiotics in poultry has steadily increased due to the growing demand for antibiotic-free poultry and their well-documented benefits. By 2018, the probiotics market in poultry had reached 80 million USD, with projections indicating growth to 125 million USD by 2025 at a compound annual growth rate of 7.7% (Ahuja K. et al., 2021). Benefits of probiotics in poultry include enhanced growth and laying performance, improved gut health, boosted immunity, and increased beneficial microbiota. Probiotics for livestock are termed direct-fed microbials or DFM. They are introduced into poultry via diets or water or administered to developing embryos through in ovo feeding technology (Pender et al 2017). DFMs, as feed additives containing beneficial microbes, they complement antibiotic usage, restore gut functions, and enhance animal performance (Bernardeau et al., 2013). They have gained popularity due to their ability to prevent bacterial gut infections and modulate immunity (Lee et al., 2017). In contrast, probiotics offer a wider range of benefits as functional foods, reducing disease risk and promoting overall health (Roberfroid et al., 2002). Commercial probiotic products often contain multiple strains to provide synergistic effects, with commonly used genera including *Bifidobacterium*, *Lactococcus*, *Lactobacillus*, *Bacillus*, *Streptococcus*, and yeast such as *Candida*. Selection criteria for probiotic strains include their tolerance to gastrointestinal conditions, adhesion ability to the gastrointestinal mucosa, and competitive exclusion of pathogens (Gadde et al., 2017). Additionally, probiotics are chosen based on their survival during manufacturing, transportation, storage, and application processes, ensuring viability and desired characteristics (Bajagai et al., 2016). Probiotics such as *B. subtilis*, *Lactobacillus* strains, *Saccharomyces* (probiotic yeast), and *Aspergillus oryzae* have antimicrobial properties against pathogenic bacteria such as *Salmonella* spp. (V.T.Nair et al., 2018).

Impact of Probiotics on Gut

In chickens, *Salmonella* first attaches to the cecal epithelial cells and then spreads to the liver, spleen, and oviduct. In pigs, early *Salmonella* infection disrupts microbiome composition and functionality, principally at the ileum, and in humans, it may affect different sites (Argüello *et al.*, 2018). Consequently, probiotic strains may work at different locations (jejunum, ileum, colon, and cecum) in different hosts. Genetically modified *L. casei* promotes overall bacterial species diversity and increases the abundance of *Lactobacillus* and *Bifidobacterium* in the cecum in mice (Peng *et al.*, 2019). Probiotics act with different mechanisms of action in their host, such as competitive exclusion, the release of bacteriostatic and bactericidal agents, immune modulation, lowering intestinal pH, and improvement of the intestinal mucosal barrier (Alagawany *et al.*, 2021b; El-Saadony *et al.*, 2021).

Probiotics play significant roles as normal commensals. They exert their therapeutic properties against *Salmonella* in four ways (Gut *et al.*, 2018). Firstly, they protect the tight junction in the gut and modulate the host's innate and acquired immunity (Pedicord *et al.*, 2016; Thiemann *et al.*, 2017). Second, they directly compete with *Salmonella* for niches and nutrients (Lam & Monack, 2014). Third, they produce antimicrobial molecules which inhibit *Salmonella* (Garcia-Gutierrez *et al.*, 2019). Fourth, they regulate *Salmonella's* virulence by regulating the expression of virulence genes (Tanner *et al.*, 2016).

Probiotics can be regulators, producers, and residents after being administered. As a modulator, it effectively regulates either the host or the pathogen. *L. casei* modulates host immunity by regulating the expression of intestinal inflammation-related cytokines (Peng *et al.*, 2019, Muiyarakandy & Amalaradjou, 2017). As producers, probiotics produce metabolites that may be signals to stimulate host immunity or substances to inhibit *Salmonella* colonization and growth. For example, *L. pentosus* AT6 and its cell-free culture supernatants inhibit *Salmonella* growth, adhesion, and invasion (Liu *et al.*, 2018). As residents, probiotics can reduce *Salmonella* through physical repellence and colonization resistance (Ubeda *et al.*, 2017).

Selecting the Right Probiotics

Probiotics can be either prokaryotes or eukaryotes. Species belonging to *Bacillus spp.*, *Bifidobacterium spp.*, *Clostridium spp.*, *Escherichia coli Nissle 1917*, and *Lactobacillus spp.*, yeast, are most commonly used (Kanmani et al., 2013). Conventional probiotics, such as *Bifidobacterium spp.* and *Lactobacillus spp.*, harbor the most well-characterized probiotic strains and are widely commercialized. These probiotics reduce more than 90% of caecal *Salmonella* load and prevent invasion of organs (Gut et al., 2018). In contrast to conventional probiotics, several next-generation probiotics have recently been identified, such as *Akkermansia muciniphila*, *Eubacterium hallii*, and *Faecalibacterium prausnitzii* (Almeida et al., 2019).

Genetically modified probiotics have attracted much interest due to their advantages and strengthened effects (Barra et al., 2020). Two approaches are used to construct genetically modified probiotics: mutation and overexpression. Considering the infection stages of *Salmonella*, probiotic mutants with high adaptability may be more suitable for prevention and specifically targeted overproduced probiotics may be more effective in therapy. For example, the expression of microcin H47 in probiotic *E. coli* inhibits *Salmonella* growth (Palmer et al., 2018). Genetically modified probiotics may be the best option for *Salmonella* intervention. Synthetic biology explores diverse biosynthetic pathways and provides versatile engineering toolboxes for probiotic strain improvement (Yadav & Shukla, 2020). These toolboxes consist of genetic circuits, different delivery systems, and many genome-editing tools, which strongly accelerated the development of advanced, genetically modified probiotics (Bober et al., 2018; Ozdemir et al., 2018). However, a newly developed strategy called inducible plasmid self-destruction provides a novel genome-editing tool for simple gene knockout and knock-in in *Lactobacilli* and *Bifidobacteria* (Zuo et al., 2019). All these advances will strengthen the prophylactic and therapeutic activity of genetically modified probiotics on *Salmonella*.

How Should Probiotics Be Formulated?

According to Almeida et al., formulating viable probiotics while enabling cost-effective biomass yield is critical for product development in translational applications (Almeida et

al., 2019). Regarding *Salmonella* prevention, conventional probiotics formulated as foods or drinks containing 10^6 CFU/g or 10^6 CFU/mL viable cells may be acceptable. However, next-generation or genetically modified probiotics are more promising in curing *Salmonella* infections. Probiotics can be formulated as concentrated pills or capsules containing more than 10^9 CFU/g or even more cells (O'Toole et al., 2017).

Overview of some critical probiotics studied for the reduction of *Salmonellosis*

Lactobacillus reuteri

L. reuteri produces lactic acid, ethanol, acetate, hydrogen peroxide, carbon dioxide, reuterin, and retericylin (Yu et al., 2007). All the above biomolecules are implicated in the inhibition of the growth of *E. coli*, *S. typhimurium*, *Helicobacter pylori*, *Staphylococcus epidermidis*, *Staphylococcus aureus*, and Rotavirus (Seo et al., 2010). Additionally, chickens supplemented with 10^8 CFU/mL *L. reuteri* revealed reduced lesion scores in birds infected with *C. perfringens*, suggesting that *L. reuteri* can modulate the innate immune response by regulating inflammatory cytokines and inhibiting *C. perfringens* proliferation (Cao et al., 2012).

***E. coli* Nissle 1917**

Forkus et al reported on technology to reduce *Salmonella enteritidis* in poultry by using *in vitro* experiments and an animal model of 300 turkeys. They engineered the probiotic *E. coli* Nissle 1917 to express and secrete the antimicrobial peptide. *Salmonella* was rapidly cleared from the ceca of the birds administered with the modified probiotic compared to the treatment groups (Forkus B.et al., 2017).

Pediococcus acidilactici

P. acidilactici is a facultative anaerobe (Lin et al., 2008). *Pediococci* suppresses enteric pathogens' growth by producing lactic acid and bacteriocins as pediocins. Dietary supplementation with *Pediococcus acidilactici*, mannan-oligosaccharide, and butyric acid reduces colonization of *S. typhimurium* and improves broiler chickens' growth performance (Jazi et al., 2018).

Enterococcus faecium

E. faecium is a gram-positive, facultative anaerobe, and *E. faecium* secretes bacteriocins (Kang & Lee, 2005). According to Beirao et al., after vaccination, *E. faecium* improves layer chickens' immune and health status with live attenuated *S. Enteritidis* vaccine. (Beirao et al., 2018).

Lactococcus lactis subsp. lactis.

Likewise, Sabo et al., 2020 isolated *L. lactis* subsp. *Lactis* from the chicken cecum and screened for their antagonistic effect towards gut pathogens. The study demonstrated a high potential of the strain to be used as probiotics in poultry feed, a valuable alternative to replace use of antibiotics against *Salmonella heidelberg* (Sabo, S.d.S., et al.,2020).

Bifidobacterium animalis

B. animalis is an anaerobic, Gram-positive, rod-shaped bacterium found naturally in the intestines of rabbits, chickens, and humans (Sanchez et al., 2007). Incorporating *Bifidobacteria* and *Lactobacillus* with broiler feed reduces the colonization of infected *Salmonella enterica* and improves broiler health (Sharkawy et al., 2020).

Lactobacillus fermentum 1.2133

According to Wang et al, 2023, *Lactobacillus fermentum* 1.2133 has antibacterial activity against *Salmonella pullorum* CVCC533. Also, the cell-free supernatant of *Lactobacillus fermentum* 1.2133 has bactericidal effect on the *Salmonella pullorum* CVCC533, a biofilm forming pathogen. It significantly reduced the number of *Salmonella* and aerobic bacteria in the chicken duodenum, ileum, and cecum, including *Escherichia* and *Shigella*, and improved *Lactobacilli* count in the gut (M. Wang et al.,2023).

Synergistic effect of lactic acid bacteria against *Salmonella* infection in chicken

Upadhaya et al, reported the beneficial effects of locally isolated probiotic strains, *Bacillus subtilis* RX7 and *Bacillus methylotrophicus* C14, on egg production, excreta, and intestinal

microbiota of laying birds challenged with *S. gallinarum* KVCC BA 0700722 (Upadhaya et al., 2016).

In vitro and in vivo studies conducted by Shanmugasundaram et al, to study the effects of supplementation on *Salmonella enteritidis* proliferation indicated that *L. reuteri*, *P. acidilactici*, *B. animalis*, and *E. faecium* can colonize the chicken intestine successfully and have the capacity to decrease the proliferation of *S. enteritidis*. They confirmed that the supplementation improves the body weight and feed intake and reduces the colonization of *Salmonella* in the cecal content of broiler chickens. (Shanmugasundaram R. et al., 2019)

Chang et al demonstrated that feed containing probiotic-supplemented basal diet containing multi-strain probiotics, namely *Lactobacillus acidophilus* LAP5, *L. fermentum* P2, *Pediococcus acidilactici* LS, and *L. casei* L21 increased *Lactobacillaceae* and reduced *Enterobacteriaceae* in the ceca. The administration of multi-strain probiotics modulated intestinal microbiota, gene expression of tight junction proteins, and immunomodulatory activity in broiler chickens (Chang et al., 2020)

Kowalska J.D. et al., 2020 fed three strains of *Lactobacillus sp.*: *L.rhamnosus* LOCK 1131, *L.casei* LOCK 1132, and *L. paracasei* LOCK 1133. They possess antagonistic activity towards a broad spectrum of environmental *Salmonella enteritidis* species. (Kowalska J.D. et al., 2020)

Likewise, Raheleh ostovan et al., 2021 used a mixture of *B. subtilis* PY79 and *B. subtilis* ATCC 6633. They used their antimicrobial substances against *S. typhimurium*. They found a significant reduction in the invasive ability of *S. typhimurium* to Caco-2 cells by employing *B. subtilis* probiotics (Raheleh Ostovan et al .,2021). *B. subtilis* KATMIRA1933 is a probiotic that can produce bacteriocins (M.H. Tazehabadi et al.,2021). *B. amyloliquefaciens* B-1895 is another probiotic with antimicrobial properties. It produces a variety of proteolytic enzymes, which have been shown to have activity against the foodborne pathogen *Listeria monocytogenes* (Algburi et al., 2020b). Studies reported that when used as probiotics in poultry, they have different impacts on rooster sperm production, egg production, hatching egg quality, egg hatchability, etc. (Mazanko et al., 2018). According to M.H. Tazehabadi et al., a mixed strain of *B. subtilis* KATMIRA1933 and *B. amyloliquefaciens* B-1895 inhibit biofilm formation of several *Salmonella* serovars. They not only act as a beneficial feed

additive for poultry to increase their physiological parameters, but also as an influential antimicrobial producers and prophylactic agents against *Salmonella* (M.H. Tazehabadi et al.,2021)

Table 1: List of probiotics organisms used in poultry.

Sr.no.	Probiotic Strains	Effects in poultry	References
1.	<i>Lactobacillus reuteri</i>	<i>L.reuteri</i> inhibits the growth of <i>E.coli</i> , and rotavirus	Yu et al., 2007; Seo et al., 2010; Cao et al., 2012
2.	<i>Escherichia coli</i> Nissle 1917	Reduce contamination in the ceca of birds	Forkus et al., 2017
3.	<i>Pediococci acidilacti</i>	Reduces colonization of <i>S. Typhimurium</i> , and improves growth performance in broiler chickens.	Lin et al., 2008; Jazi et al., 2018
4.	<i>Enterococcus faecium</i>	Improve gut microbiota.	Kang & Lee, 2005; Beirao et al., 2018
5.	<i>Lactococcus lactis</i> subsp. lactis	Alternative to antibiotics, it reduces pathogen load	Sabo et al., 2020
6.	<i>Bifidobacterium animalis</i>	Improves broiler chicken condition by reducing gut microbiota	Sanchez et al., 2007; Sharkawy et al., 2020
7.	<i>Lactobacillus fermentum</i> 1.2133	It significantly reduces <i>Salmonella</i> proliferation the chicken duodenum, ileum and cecum	M. Wang et al., 2023
8.	<i>Bacillus subtilis</i> RX7 and <i>Bacillus</i>	Improve egg production, modifies intestinal microbiota	Upadhaya et al., 2016

	<i>methylophilic</i> C14		
9.	<i>L.reuteri</i> , <i>P.acidilactici</i> , <i>B.animalis</i> and <i>E.faceium</i>	Enhances intestinal health, reduces pathogen load	Shanmugasundaram et al., 2019
10.	<i>Lactobacillus</i> <i>acidophilus</i> LAP5, <i>L.</i> <i>fermentum</i> P2, <i>Lactis</i> <i>demonstrates</i> , and <i>L.</i> <i>casei</i> L21	Reduced <i>Enterobacteriaceae</i> in the Ceca of chicken	Chang et al., 2020
11.	<i>L. rhamnosus</i> LOCK 1131, <i>L. casei</i> LOCK 1132, and <i>L. paracasei</i> LOCK 1133	Shows Antagonistic activity against Broad spectrum of <i>Salmonella enteritidis</i>	Kowalska et al., 2020
12.	<i>B. subtilis</i> PY79 and <i>B.</i> <i>subtilis</i> ATCC 6633	Improve gut health by inhibit invasion of <i>S.Typhimurium</i> on Caco-2 cells	Raheleh Ostovan et al., 2021
13.	<i>B.subtilis</i> KATMIRA1933 and <i>B.amyloliquefaciens</i> B- 1895	Improves egg production, hatching egg quality	M.H. Tazehabadi et al., 2021
14.	<i>S. entitidis</i>	Reduce <i>Salmonella</i> contamination	Juricova et al., 2022

Conclusions and Future Directions

The problem of antibiotic resistance is that antibiotic-resistant clones of several major pathogens, particularly *Salmonella*, have increasingly been found in foods, which include poultry, retail meat products and seafood. All determinants of maximal vital resistance, which include those conferring resistance to β -lactams, extended-spectrum β -lactams,

fluoroquinolones, aminoglycosides, tetracyclines, and chloramphenicol, have been identified in different *Salmonella* serovars isolated from the food supply. It is increasingly clear that antibiotic resistance will remain an obstacle for the foreseeable future. These studies have demonstrated the effectiveness of probiotic manipulation as part of prevention or internal treatment in competition with *Salmonella* infection. There are different mechanisms by which probiotic can additionally exert their results. They include non-immune mechanisms and modulation of mucosal and systemic immune responses. Moreover, one or more probiotic lines in a fermented product can increase the beneficial properties of the contained probiotic traces. One crucial goal of the poultry industry is the reduction of foodborne pathogens, namely *Salmonella spp.*, *Campylobacter spp.*, and *C.perfringens*. Through the introduction of probiotic bacteria in diets; these pathogenic microorganisms, the primary cause of diseases in humans transmitted by contaminated foods can be eliminatd.

REFERENCES

1. Abd El-Hack, M. E., B. A. Alaidaroos, R. M. Farsi, D. E. Abou-Kassem, M. T. El-Saadony, A. M. Saad, M. E. Shafi, N. M. Albaqami, A. E. Taha, and E. A. Ashour. (2021)d. Impacts of supplementing broiler diets with biological curcumin, zinc nanoparticles, and *Bacillus licheniformis* on growth, carcass traits, blood indices, meat quality, and cecal microbial load. *Animals* 11:1878.
2. Abd El-Hack, M. E., M. T. El-Saadony, A. A. Swelum, M. Arif, M. M. Abo Ghanima, M. Shukry, A. Noreldin, A. E. Taha, and K. A. El-Tarabily. (2021)c. Curcumin, the active substance of turmeric: its effects on health and ways to improve its bioavailability. *J. Sci. Food Agric.* 101:5747–5762.
3. Abd Elkader, A. M., S. Labib, T. F. Taha, F. Althobaiti, A. Aldhahrani, H. M. Salem, M. Saad, and F. M. Ibrahim. (2021). Phytogetic compounds from avocado (*Persea americana* L.) extracts; antioxidant activity, amylase inhibitory activity, therapeutic potential of type 2 diabetes. [e-pub ahead of print]. *Saudi J. Biol. Sci.* doi:10.1016/j.sjbs.2021.11.031. Accessed Nov. 24, 2021.

4. Abdel-Moneim, A. M. E., M. T. El-Saadony, A. M. Shehata, A. M. Saad, S. A. Aldhumri, S. M. Ouda, and N. M. Mesalam. (2021). Antioxidant and antimicrobial activities of *Spirulina platensis* extracts and biogenic selenium nanoparticles against selected pathogenic bacteria and fungi [e-pub ahead of print]. *Saudi J. Biol. Sci.*, doi:10.1016/j.sjbs.2021.09.046.
5. Abdelnour, S. A., A. A. Swelum, A. Salama, M. Q. Al-Ghadi, S. Y. A. Qattan, M. E. Abd El-Hack, A. F. Khafaga, A. R. Alhimaidi, B. O. Almutairi, A. A. Ammari, and M.T. El-Saadony. (2020)b. The beneficial impacts of dietary phycocyanin supplementation on growing rabbits under high ambient temperature. *Ital. J. Anim. Sci.* 19:1046–1056.
6. Abdelnour, S. A., M. T. El-Saadony, S. A. M. Saghir, M. E. Abd El-Hack, O. Y. A. Al-charge, N. Al-Gabri, and A. Salama. (2020)a. Mitigating the negative impacts of heat Stress in growing rabbits via dietary prodigiosin supplementation. *Livest. Sci.* 240:104220.
7. Abou-Kassem, D. E., K. M. Mahrose, R. A. El-Samahy, M. E. Shafi, M. T. El-Saadony, M. E. Abd El-Hack, M. Emam, M. El-Sharnouby, A. E. Taha, and E. A. Ashour. (2021)b. Influences of dietary herbal blend and feed restriction on growing rabbits' growth, carcass characteristics, and gut microbiota. [e-pub ahead of print]. *Ital. J. Anim. Sci.* 20:896–910, doi:10.1016/j.sjbs.2021.10.063.
8. Abou-Kassem, D. E., M. M. El-Abasy, M. S. Al-Harbi, S. Abol-Ela, H. M. Salem, A.M. El-Tahan, M. T. El-Saadony, M. E. Abd El-Hack, and E. A. Ashour. (2021)a. Influences of total sulfur amino acids and photoperiod on growth, carcass traits, blood parameters, meat quality and cecal microbial load of broilers. *Saudi J. Biol. Sci.*
9. Ahuja K., Mamtani K. Poultry Probiotic Ingredients Market Size by Product (Lactobacilli, Bifidobacterium, Streptococcus, Bacillus), by Application (Broilers, Layers, Turkeys, Breeders, Chicks & Poults), Regional Outlook, Application Potential, Price Trends, Competitive Market Share & Forecast, 2019–2025.
10. Alagawany, M., M. Madkour, M. T. El-Saadony, and F. M. Reda. (2021)b. *Paenibacillus polymyxa* (LM31) as a new feed additive: antioxidant and antimicrobial activity and its effects on growth, blood biochemistry, and intestinal

- bacterial populations of growing Japanese quail. *Anim. Feed Sci. Technol.* 276:114920.
11. Almeida, D., Machado, D., Andrade, J. C., Mendo, S., Gomes, A.M., and Freitas, A.C. (2019). Evolving trends in next-generation probiotics: a 5W1H perspective. *Crit. Rev. Food Sci. Nutr.* 60, 1783–1796. doi: 10.1080/10408398.2019.1599812
 12. Argüello, H., Estellé, J., Zaldívar-López, S., Jiménez-Marín, Á., Carvajal, A., López-Bascón, M. A., et al. (2018). Early *Salmonella typhimurium* infection in pigs disrupts microbiome composition and functionality, principally at the ileum mucosa. *Sci. Rep.* 8:7788. doi: 10.1038/s41598-018-26083-3
 13. Arif, M., R. S. Baty, E. H. Althubaiti, M. T. Ijaz, M. Fayyaz, M. E. Shafi, N. M. Albaqami, M. Alagawany, M. E. Abd El-Hack, A. E. Taha, H. M. Salem, A. M. El-Tahan, and S. S. Elnesr. (2021). The impact of betaine supplementation in quail diet on growth performance, blood chemistry, and carcass traits. [e-pub ahead of print]. *Saudi J. Biol. Sci.*, doi:10.1016/j.sjbs.2021.11.002. Accessed Nov. 11, 2021.
 14. Asefa Kebede, I., & Duga, T. (2022). Prevalence and Antimicrobial Resistance of *Salmonella* in Poultry Products in Central Ethiopia. *Veterinary Medicine International*, 2022, 8625636. <https://doi.org/10.1155/2022/8625636>.)salmonellosis *Salmonellosis*
 15. Ashour, E. A., M. E. Ab El-Hack, M. E. Shafi, W. Y. Alghamdi, A. E. Taha, A. A. Swelum, M. T. El-Saadony. 2020. Green coffee powder supplementation impacts growth performance, carcass characteristics, blood indices, meat quality, and gut microbial load in broilers agriculture 10:457.
 16. Ashour, E. A., R. M. Farsi, B. A. Alaidaroos, A. M. E. Abdel-Moneim, M. T. El-Saadony, A. O. Osman, E. T. Abou Sayed-Ahmed, N. M. Albaqami, M. E. Shafi, E. Taha, & M. E. Abd El-Hack. (2021). Impacts of dietary supplementation of pyocyanin powder on growth performance, carcass traits, blood chemistry, meat quality and gut microbial activity of broilers. *Ital. J. Anim. Sci.* 20:1357–1372.
 17. Attia, M. M., and H. M. Salem. (2022). Morphological and molecular characterization of *Pseudolynchia canariensis* (Diptera: Hippoboscidae) infesting domestic pigeons. *Int. J. Trop. Insect Sci* 42:733–740.

18. Bajagai Y.S., Klieve A.V., Dart P.J., Bryden W.L.(2016). *Animal Nutrition: Production, Impact and Regulation*. Food and Agriculture Organization of the United Nations; Rome, Italy: Animal Production and Health Div Probiotics.
19. Barra,M., Danino, T., and Garrido, D. (2020). Engineered probiotics for the detection and treatment of inflammatory intestinal diseases. *Front. Bioeng. Biotechnol.*8:265. Doi: 10.3389/fbioe.2020.00265
20. Beir~ao, B. C. B., M. Ingberman, C. J. Favaro, D. Mesa,L. C. Bittencourt, V. B. Fascina, and L. F. Caron.(2018). Effect of an Enterococcus faecium probiotic on specific IgA following live Salmonella Enteritidis vaccination of layer chickens. *Avian Pathol.*47:325–333.
21. Bernardeau M., Vernoux J.-P.(2013) Overview of differences between microbial feed additives and probiotics for food regarding regulation, growth promotion effects, health properties, and consequences for extrapolating farm animal results to humans. *Clin. Microbiol. Infect.*;19:321–330. doi: 10.1111/1469-0691.12130.
22. Bober, J. R., Beisel, C. L., and Nair, N. U. (2018). Synthetic biology approaches to engineer probiotics and members of the human microbiota for biomedical applications. *Annu. Rev. Biomed. Eng.* 20, 277–300. doi: 10.1146/annurev-bioeng-062117-121019
23. Cao, L., X. J. Yang, Z. J. Li, F. F. Sun, X. H. Wu, and J. H. Yao. (2012). Reduced lesions in chickens with Clostridium perfringens-induced necrotic enteritis by Lactobacillus fermentum 1.20291. *Poult. Sci.* 91:3065–3071.
24. Castellanos, L. R., P. Donado-Godoy, M. Leon, V. Clavijo,A. Arevalo, J. F. Bernal, A.J. Timmerman, D. J. Mevius, J. A. Wagenaar, and J. Hordijk. (2017). High heterogeneity of Escherichia coli sequence types harboring ESBL/AmpC genes on IncI1 plasmids in the Colombian poultry chain. *PLoS One* 12: e0170777
25. Chang, C. H., Teng, P. Y., Lee, T. T., & Yu, B. (2020). Effects of multi-strain probiotic supplementation on intestinal microbiota, tight junctions, and inflammation in young broiler chickens challenged with Salmonella enterica subsp. enterica. *Asian-Australasian journal of animal sciences*, 33(11), 1797–1808. <https://doi.org/10.5713/ajas.19.0427>

26. Contamination in broiler birds. *PLoS ONE* 14(10): e0223577. <https://doi.org/10.1371/journal.pone.0223577>
27. Donaldson, G. P., Lee, S.M., & Mazmanian, S. K. (2016). Gut biogeography of the bacterial microbiota. *Nat. Rev. Microbiol.* 14, 20–32. doi: 10.1038/nrmicro3552
28. Done, H.Y.; Venkatesan, A.K.; Halden, R.U. (2015) Does the recent growth of aquaculture create antibiotic resistance threats different from those associated with land animal production in agriculture? *AAPS J.* 17, 513–524.
29. El-Saadany, A. S., El-Barbary, A., El-Salam, A. A., Ahmed, M., and Shreif, E. (2022). Nutritional and physiological evaluation of quercetin as a phytochemical feed additive in laying hens. *Journal of Animal and Feed Sciences*, 31(3), pp.249-257. <https://doi.org/10.22358/jafs/150080/2022>
30. El-Saadony, M. T., A. M. Saad, H. A. Elakkad, A. M. El-Tahan, O. A. Alshahrani, M.S. Alshilawi, H. El-Sayed, S. A. Amin, and A. I. Ahmed. (2021) c. Flavoring and extending the shelf life of cucumber juice with aroma compounds-rich herbal extracts at four °C through controlling chemical and microbial fluctuations. *Saudi J. Biol. Sci.*, doi:10.1016/j.sjbs.2021.08.092. Accessed Sept. 4, 2021
31. El-Saadony, M. T., M. E. Abd El-Hack, A. A. Swelum, S. I. Al-Sultan, W. R. El-Ghareeb, E. O. Hussein, E. O. S. Hussein, H. A. Ba-Awadh, B. A. Akl, and M. M. Nader. (2021)b. Enhancing quality and safety of raw buffalo meat using the bioactive peptides of pea and red kidney beans under refrigeration conditions. *Ital. J. Anim. Sci.* 20:762–776.
32. El-Saadony, M. T., O. S. Khalil, A. Osman, M. S. Alshilawi, A. E. Taha, S. M. Aboelenin, M. Shukry, and A. M. Saad. (2021)a. Bioactive peptides supplemented raw buffalo milk: biological activity, shelf life, and quality properties during cold preservation. *Saudi J. Biol. Sci.* 28:4581–4591.
33. El-Sharkawy, H., A. Tahoun, A. M. Rizk, T. Suzuki, W. Elmonir, E. Nassef, M. Shukry, M. O. Germoush, F. Farrag, M. Bin-Jumah, & A. M. Mahmoud. (2020). Evaluating bifidobacteria and *Lactobacillus* probiotics as an alternative therapy for *Salmonella typhimurium* infection in broiler chickens. *Animals* 10:1023.
34. Eng, S., P. Pusparajah, N. Ab Mutalib, H. Ser, K. Chan, and L. Lee. (2015). *Salmonella*: A review on pathogenesis, epidemiology, and antibiotic resistance. *Front. Life Sci.* 8:284–293.

35. Ferrari, R. G., Rosario, D., Cunha-Neto, A., Mano, S. B., Figueiredo, E., and Conte-Junior, C. A. (2019). Worldwide epidemiology of *Salmonella* serovars in animal-based foods: a meta-analysis. *Appl. Environ. Microbiol.* 85, e00591–e00519. doi: 10.1128/AEM.00591-19
36. Feye, K. M., M. F. A. Baxter, G. Tellez, M. H. Kogut, & S. C. Ricke. (2020). Influential factors on the composition of the conventionally raised broiler gastrointestinal microbiomes. *Poult. Sci.*99:653–659.
37. Forkus B, Ritter S, Vlysidis M, Geldart K, Kaznessis YN. Antimicrobial Probiotics Reduce *Salmonella Enterica* in Turkey's Gastrointestinal Tracts. *Sci Rep.* (2017) Jan 17;7:40695. Doi: 10.1038/srep40695. PMID: 28094807; PMCID: PMC5240571.
38. Gadde U.D., Kim W.H., Oh S.T., Lillehoj H.S. Alternatives to antibiotics for maximizing growth performance and feed efficiency in poultry:(2017) A review. *Anim. Health Res. Rev.*;18:26–45. doi: 10.1017/S1466252316000207.
39. Garcia-Gutierrez, E., Mayer, M. J., Cotter, P. D., and Nrabad, A. (2019). The gut microbiota is a source of novel antimicrobials. *Gut Microbes.* 10, 1–21. doi: 10.1080/19490976.2018.1455790
40. Gut, A. M., Vasiljevic, T., Yeager, T., and Donkor, O. N. (2018). *Salmonella* infection prevention and treatment by antibiotics and probiotic yeasts: a review. *Microbiology* 164, 1327–1344. doi: 10.1099/mic.0.000709
41. Hafez, Yaser, Kotb Attia, Salman Alamery, Abdelhalim Ghazy, Abdullah Al-Doss, Eid Ibrahim, Emad Rashwan, Lamiaa El-Maghraby, Ahmed Awad, and Khaled Abdelaal. (2020). "Beneficial Effects of Biochar and Chitosan on Antioxidative Capacity, Osmolytes Accumulation, and Anatomical Characters of Water-Stressed Barley Plants" *Agronomy* 10, no. 5: 630. <https://doi.org/10.3390/agronomy10050630>
42. Havelaar AH, Kirk MD, Torgerson PR, Gibb HJ, Hald T, Lake RJ, Praet N, Bellinger DC, de Silva NR, Gargouri N, Speybroeck N, Cawthorne A, Mathers C, Stein C, Angulo FJ, Devleeschauwer B; World Health Organization Foodborne Disease Burden Epidemiology Reference Group. World Health Organization Global Estimates and Regional Comparisons of the Burden of Foodborne Disease in 2010. *PLoS Med.* 2015 Dec 3;12(12):1001923.

43. Hill, C., Guarner, F., Reid, G., Gibson, G. R., Merenstein, D. J., Pot, B., et al.(2014). Expert consensus document: The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nat. Rev. Gastroenterol. Hepatol.* 11, 506–514. doi: 10.1038/nrgastro.2014.66
44. Isayas Asefa Kebede, Tefari Duga,(2022) "Prevalence and Antimicrobial Resistance of Salmonella in Poultry Products in Central Ethiopia," *Veterinary Medicine International*, vol. 2022, Article ID 8625636, 7 pages, 2022. <https://doi.org/10.1155/2022/8625636>
45. Jazi, V., A. D. Foroozandeh, M. Toghyani, B. Dastar, R. R. Koochak Saraie, and M. Toghyani. (2018). Effects of *Pediococcus acidilactici*, mannan-oligosaccharide, butyric acid and their combination on growth performance and intestinal health in young broiler chickens challenged with *Salmonella typhimurium*. *Poult.Sci.* 97:2034–2043.
46. Juricova, H.; Matiasovicova, J.; Faldynova, M.; Sebkova, A.; Kubasova, T.; Prikrylova, H.; Karasova, D.; Crhanova, M.; Havlickova, H.; Rychlik, I. (2022)Probiotic Lactobacilli Do Not Protect Chickens against *Salmonella Enteritidis* Infection by Competitive Exclusion in the Intestinal Tract but in Feed, Outside the Chicken Host. *Microorganisms* 2022, 10, 219. <https://doi.org/10.3390/microorganisms10020219>
47. Kang, J. H., and M. S. Lee. (2005). Characterization of a bacteriocin produced by *Enterococcus faecium* GM-1 isolated from an infant. *J. Appl. Microbiol.* 98:1169–1176.
48. Kanmani, P., Satish Kumar, R., Yuvaraj, N., Paari, K. A., Pattukumar, V., and Arul, V. (2013). Probiotics and its functionally valuable products - a review. *Crit. Rev. Food Sci. Nutr.* 53, 641–658. doi: 10.1080/10408398.2011.553752
49. Kirk MD, Pires SM, Black RE, Caipo M, Crump JA, et al. (2015). Correction: World Health Organization Estimates of the Global and Regional Disease Burden of 22 Foodborne Bacterial, Protozoal, and Viral Diseases, 2010: A Data Synthesis. *PLOS Medicine* 12(12): e1001940. <https://doi.org/10.1371/journal.pmed.1001940>

50. Kowalska JD, Nowak A, Śliżewska K, Stańczyk M, Łukasiak M, Dastyh J. Anti-Salmonella Potential of New *Lactobacillus* Strains with the Application in the Poultry Industry. *Pol J Microbiol.* (2020). Sep;69(1):5–18. doi: 10.33073/pjm-2020-001. Epub 2020 Jan 28. PMID: 32189480; PMCID: PMC7256722.
51. Lam, L. H., and Monack, D. M. (2014). Intraspecies competition for niches in the distal gut dictate transmission during persistent Salmonella infection. *PLoS Pathog.* 10:e1004527. doi: 10.1371/journal.ppat.1004527
52. Lan, R., P. R. Reeves, & S. Octavia. (2009). Population structure, origin, and evolution of major Salmonella enterica clones. *Infect. Genet. Evol.* 9:996–1005.
53. Lauková, A., Marciňáková, M., Strompfová, V., Ouwehand, A.C. (2008a). Probiotic potential of enterococci isolated from canine feed. *Folia Microbiologica* 53: 84-88.
54. Lauková, A., Simonová, M., Strompfová, V., Štyriak, I., Ouwehand, A., Várady, M. (2008b). Potential of enterococci isolated from horses. *Anaerobe* 14: 234-236
55. Lee K.-W., Lillehoj H.S. An update on direct-fed microbials in broiler chickens in post-antibiotic era. (2017) *Anim. Prod. Sci.* 2017;57:1575. doi: 10.1071/AN15666.
56. Li, H., H. Wang, J. Y. D’Aoust, and J. Maurer. (2013). Salmonella Species. Pages 225–261 in *Food Microbiology Fundamentals and Frontiers*. M. P. Doyle and R. L. Buchanan, eds. 4th ed. ASM Press, Washington, DC.
57. Lin, Y. P., C. H. Thibodeaux, J. A. Pena, G. D. Ferry, and J. Versalovic. (2008). Probiotic *Lactobacillus reuteri* suppresses proinflammatory cytokines via c-Jun. *Inflamm. Bowel Dis.* 14:1068–1083.
58. Liu, J., Gu, Z., Lu, W., Hu, D., Zhao, X., Huang, H., et al. (2018). *Lactobacillus* applied multiple mechanisms, including pentosus AT6, to mute the lethal effects of salmonella in a mouse model. *Food Funct.* 9, 2787–2795. doi: 10.1039/C7FO01858D7538
59. Llamas-Moya, S., C. P. Girdler, S. M. M. Shalash, A. M. Atta, H. B. Gharib, E. A. Morsy, H. Salim, M. H. H. Awaad, and M. Elmenaway. (2019). Effect of a multicarbohydase containing agalactosidase enzyme on broilers' performance, carcass yield, and humoral immunity fed corn–soybean meal–based diets of varying energy density. *J. Appl. Poult. Res.* 29:142–151.

60. Marshall, B. M., & S. B. Levy. (2011). Food animals and antimicrobials: impacts on human health. *Clin. Microbiol. Rev.* 24:718–733.
61. Min Wang (2023) , *Lactobacillus fermentum* 1.2133 display probiotic potential *in vitro* and protect against *Salmonella pullorum* in chicken of infection, *Letters in Applied Microbiology*, Volume 76, Issue 1, January 2023,ovac041, <https://doi.org/10.1093/lambio/ovac041>
62. Muiyyarikkandy, M. S., and Amalaradjou, M. A. (2017). *Lactobacillus bulgaricus*, *Lactobacillus rhamnosus*, and *Lactobacillus paracasei* attenuate *Salmonella enteritidis*, *Salmonella Heidelberg*, and *Salmonella typhimurium* colonization and virulence gene expression *in vitro*. *Int. J. Mol. Sci.* 18:2381.doi: 10.3390/ijms18112381
63. Nirmala, T.V., Reddy, A.D., Sree, E.K., Subbaiah, K.V., Raju, G.S. and Reddy, R.V.S., (2018). Salmonellosis in poultry: A case report. *Int. J. Curr. Microbiol. Appl. Sci.* 7(2), pp.2347-2349.
64. O'Toole, P. W., Marchesi, J. R., & Hill, C. (2017). Next-generation probiotics: the spectrum from probiotics to live biotherapeutics. *Nat Microbiol.* 2:17057. doi: 10.1038/nmicrobiol.2017.57
65. Obaidat,M.M., and Stringer, A. P. (2019). Prevalence, molecular characterization, and antimicrobial resistance profiles of *Listeria monocytogenes*, *Salmonella enterica*, and *Escherichia coli* O157:H7 on dairy cattle farms in Jordan. *J. Dairy Sci.* 102, 8710–8720. doi: 10.3168/jds.2019-16461
66. Ostovan, R., Pourmontaseri, M., Hosseinzadeh, S., & Shekarforoush, S. S. (2021). Interaction between the probiotic *Bacillus subtilis* and *Salmonella Typhimurium* in Caco-2 cell culture. *Iranian journal of microbiology*, 13(1), 91–97. <https://doi.org/10.18502/ijm.v13i1.5497>
67. Ozdemir, T., Fedorec, A. J. H., Danino, T., and Barnes, C. P. (2018). Synthetic biology and engineered live biotherapeutics: toward increasing system complexity. *Cell Syst.* 7, 5–16. doi: 10.1016/j.cels.2018.06.008
68. Palmer, J. D., Piattelli, E., McCormick, B. A., Silby, M. W., Brigham, C. J., & Bucci,V. (2018). Engineered probiotic for the inhibition of *Salmonella* via tetrathionate-induced production of microcin H47. *ACS Infect. Dis.* 4, 39–45.doi: 10.1021/acsinfecdis.7b0011

69. Pedicord, V. A., Lockhart, A., Rangan, K. J., Craig, J. W., Loschko, J., Rogoz, A., et al. (2016). They exploit a host-commensal interaction to promote intestinal barrier function and enteric pathogen tolerance. *Sci Immunol.* 1: 7732.
70. Peng, M., Tabashsum, Z., Patel, P., Bernhardt, C., Biswas, C., Meng, J., et al. (2019). Prevention of enteric bacterial infections and modulation of gut microbiota with conjugated linoleic acids producing *Lactobacillus* in mice. *Gut Microbes.* (2020) 11, 433-452. doi: 10.1080/19490976.2019.1638724
71. Pender C.M., Kim S., Potter T.D., Ritzi M.M., Young M., Dalloul R.A. (2017) In ovo supplementation of probiotics and its effects on performance and immune-related gene expression in broiler chicks. *Poult. Sci.*;96:1052–1062.
72. Prieto, M.; O’Sullivan, L.; Tan, S.; McLoughlin, P.; Hughes, H.; Gutierrez, M.; Lane, J.; Hickey, R.; Lawlor, P.; Gardiner, G. (2014) In vitro assessment of marine *Bacillus* for use as livestock probiotics. *Mar. Drugs.* 12, 2422–2445.
73. Rajagopal, R. and Mini, M. (2013). Outbreaks of salmonellosis in three different poultry farms of Kerala, India. *Asian Pac J Trop Biomed* 3(6): 496-500
74. Reda, F., M. T. El-Saadony, T. K. El-Rayes, M. Farahat, G. Attia, and M. Alagawany. (2021)a. Dietary effect of licorice (*Glycyrrhiza glabra*) on quail performance, carcass, blood metabolites and intestinal microbiota. *Poult. Sci.* 100:101266.
75. Ricke, S. C., S. I. Lee, S. A. Kim, S. H. Park, & Z. Shi. (2020). Prebiotics and the poultry gastrointestinal tract microbiome. *Poult. Sci.* 99:670–677.
76. Roberfroid M.B. Global view on functional foods: European perspectives. (2002) *Br. J. Nutr.* ;88:133–S138.
77. S.D. Upadhaya, A. Hossiendoust, I.H. Kim, (2016) Probiotics in *Salmonella*-challenged Hy-Line brown layers, *Poultry Science*, Volume 95, Issue 8,, Pages 1894-1897.
78. Saad, A. M., A. S. Mohamed, M. T. El-Saadony, and M. Z. Sitohy. (2021)a. Palatable functional cucumber juices supplemented with polyphenols-rich herbal extracts. *LWT-Food Sci. Technol.* 148:111668.
79. Saad, A. M., M. T. El-Saadony, A. M. El-Tahan, S. Sayed, M. A. Moustafa, A. E. Taha, T. F. Taha, and M. M. Ramadan. (2021)b. Polyphenolic extracts from pomegranate and watermelon wastes as a substrate to fabricate sustainable silver

- nanoparticles with larvicidal effect against *Spodoptera littoralis*. Saudi J. Biol. Sci. 28:5674–5683.
80. Saad, A. M., M. Z. Sitohy, A. I. Ahmed, N. A. Rabie, S. A. Amin, S. M. Aboelenin, M. M. Soliman, and M. T. El-Saadony. (2021)c. Biochemical and functional characterization of kidney bean protein alcalde-hydrolysates and their preservative action on stored chicken meat. *Molecules* 26:4690.
81. Sabo, S.d.S., Mendes, M.A., Araújo, E.d.S. *et al.* (2020) Bioprospecting of probiotics with antimicrobial activities against *Salmonella* Heidelberg that produce B-complex vitamins as potential supplements in poultry nutrition. *Sci Rep* 10, 7235 . <https://doi.org/10.1038/s41598-020-64038-9>
82. Salem, H. M., N. Yehia, S. Al-Otaibi, A. M. El-Shehawi, A. A. M. E. Elrys, M. T. El-Saadony, & M. M. Attia. (2021). The prevalence and intensity of external parasites in domestic pigeons (*Columba et al.*) in Egypt with particular reference to the role of deltamethrin as an insecticidal agent. *Saudi J. Biol. Sci*, doi:10.1016/j.sjbs.2021.10.042. Accessed Oct. 22, 2021.
83. Sanchez, B., M. C. Champomier-Verges, B. Stuer-Lauridsen, P. Ruas-Madiedo, P. Anglade, F. Baraige, C. G. de los Reyes-Gavilan, E. Johansen, M. Zagorec, and A. Margolles. (2007). Adaptation and response of *Bifidobacterium animalis* subsp. *lactis* to bile: a proteomic and physiological approach. *Appl. Environ. Microbiol.* 73:6757–6767.
84. Scallan E, Hoekstra RM, Mahon BE, Jones TF, Griffin PM. An assessment of the human health impact of seven leading foodborne pathogens in the United States using disability-adjusted life years. *Epidemiol Infect.* (2015) Oct;143(13):2795-804. doi: 10.1017/S0950268814003185. Epub 2015 Jan 30. PMID: 25633631; PMCID: PMC9151020.
85. Schulfer, A. F., Battaglia, T., Alvarez, Y., Bijnens, L., Ruiz, V. E., Ho, M., et al. (2018). Intergenerational transfer of antibiotic-perturbed microbiota enhances colitis in susceptible mice. *Nat Microbiol.* 3, 234–242. doi: 10.1038/s41564-017-0075-5
86. Seo, B. J., M. R. Mun, V. J. Rejish Kumar, C. J. Kim, I. Lee, Y. H. Chang, & Y. H. Park. (2010). Bile-tolerant *Lactobacillus reuteri* isolated from pig feces inhibits enteric bacterial pathogens and porcine rotavirus. *Vet. Res. Commun.* 34:323–333.

87. Setta, A., H. M. Salem, M. Elhady, A. El-Hussieny, and A. A. Arafa.(2018). Molecular and genetic characterization of infectious bronchitis viruses isolated from commercial chicken flocks in Egypt between 2014 and 2016. *J. World Poult. Res.* 8:01–08.
88. Shanmugasundaram R, Mortada M, Cosby DE, Singh M, Applegate TJ, Syed B, et al.(2019). Synbiotic supplementation to decrease *Salmonella* colonization in the intestine and carcass
89. Simonová M, Lauková A. Bacteriocin activity of enterococci from rabbits. *Vet Res Commun.* (2007) Feb;31(2):143-52. doi: 10.1007/s11259-006-3411-4. Epub 2007 Jan 9. PMID: 17216320.
90. Soliman, S. M., M. M. Attia, M. S. Al-Harbi, A. M. Saad, M. T. El-Saadony, and H.M. Salem. (2021). Low host specificity of *Hippobosca equina* infestation in different Domestic animals and pigeons. *Saudi J. Biol. Sci.*, doi:10.1016/j.sjbs.2021.11.050. Accessed Nov. 24, 2021.
91. Sweeney, M. T., B. V. Lubbers, S. Schwarz, & J. L. Watts. (2018). Applying definitions for multidrug resistance, extensive drug resistance, and pan drug resistance to clinically significant livestock and companion animal bacterial pathogens. *J. Antimicrob. Chemother.* 73:1460–1463.
92. Tanner, S. A., Chassard, C., Rigozzi, E., Lacroix, C., and Stevens, M. J. (2016). *Bifidobacterium thermophilum* RBL67 impacts the growth and virulence gene expression of *Salmonella enterica* subsp enteric serovar Typhimurium. *BMC Microbiol.* 16:46. doi: 10.1186/s12866-016-0659-x
93. Tazehabadi MH, Algburi A, Popov IV, Ermakov AM, Chistyakov VA, Prazdnova EV, Weeks R, and Chikindas ML (2021). Probiotic Bacilli Inhibit *Salmonella* Biofilm Formation Without Killing Planktonic Cells. *Front. Microbiol.* 12:615328. doi: 10.3389/fmicb.2021.615328
94. Thiemann, S., Smit, N., Roy, U., Lesker, T. R., Gálvez, E., Helmecke, J., et al. (2017). Enhancement of IFN γ production by distinct commensals ameliorates *Salmonella*-induced disease. *Cell Host Microbe.* 21, 682–694. doi: 10.1016/j.chom.2017.05.005

95. Ubeda, C., Djukovic, A., and Isaac, S. (2017). U pr Roles of the intestinal microbiota in pathogen protection. *Clin Transl Immunol.* 6:e128. doi: 10.1038/cti.2017.2
96. V T Nair, D., Venkitanarayanan, K., & Kollanoor Johny, A. (2018). Antibiotic-Resistant *Salmonella* in the Food Supply and the Potential Role of Antibiotic Alternatives for Control. *Foods (Basel, Switzerland)*, 7(10), 167. <https://doi.org/10.3390/foods7100167>.
97. Wibisono FJ, Sumiarto B, Untari T, Effendi MH, Permatasari DA, Witaningrum AM. The Presence of Extended Spectrum Beta-Lactamase (ESBL) Producing *Escherichia coli* On Layer Chicken Farms In Blitar Area, Indonesia. *Biodiversitas.* (2020); 21 (6): 2667-2671. www.poultrydvm.com › condition › in Chickens - PoultryDVM
98. Yadav, M., and Shukla, P. (2020). Efficiently engineered probiotics using synthetic biology approaches: a review. *Biotechnol. Appl. Biochem.* 67, 22–29. doi: 10.1002/bab.1822
99. Yu, B., J. R. Liu, M. Y. Chiou, Y. R. Hsu, & P. W. S. Chiou. (2007). The effects of probiotic *Lactobacillus reuteri* Pg4 strain on intestinal characteristics and performance in broilers. *Asian Australas. J. Anim. Sci.* 20:1243–1251.
100. Zuo, F., Zeng, Z., Hammarström, L., and Marcottem, H. (2019). Inducible plasmid self-destruction (IPSD) assisted genome engineering in *Lactobacilli* and *Bifidobacteria*. *ACS Synth. Biol.* 8, 1723–1729. doi: 10.1021/acssynbio.9b 00114