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## SYNTHESIS AND BIOLOGICAL EVALUATION OF SOME NOVEL DERIVATIVES OF (E)-2-ACETOXY-5-(PHENYLDIAZENYL)BENZOIC ACID DERIVATIVES FOR ANTI-BACTERIAL AND ANTI-FUNGAL ACTIVITIES”

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### ABSTRACT:

Azo compounds, characterized by their distinctive azo moiety and aromatic systems, play pivotal roles across various industries due to their versatile applications. Their prevalence in food, cosmetics, pharmaceuticals, dyes, and textiles underscores their significance in modern chemistry. Notably, azo compounds exhibit diverse biological activities, contributing significantly to medicinal and therapeutic advancements. Recent research explores compounds like Congo Red and Evans Blue for their potential as HIV replication inhibitors, highlighting ongoing efforts in disease treatment. Additionally, their antibacterial and pesticidal properties make them promising candidates for industrial and medical applications. Synthesis of azo compounds offers pathways for exploring novel functionalities and derivatives, further enhancing their utility. Commonly referred to as chromogens, azo compounds find extensive use as synthetic dyes, with broad applications from textiles to photoconductive materials. Their role as chemotherapeutic agents and enzyme inhibitors underscores their broad utility. MIC assays provide valuable insights into their antimicrobial efficacy, aiding in combating infectious diseases. In summary, the multifaceted properties of azo compounds drive ongoing research and innovation, showcasing their significance in both industrial and medicinal realms.

**Keywords:** Azo-moiety, acetyl salicylic acid, anti-bacterial, anti-fungal

### INTRODUCTION

The azo moiety (-N=N-) and two unique or related monocyclic, polycyclic, or heterocyclic aromatic systems distinguish azo compounds from other colours. They have numerous uses in the food, cosmetic, pharmaceutical, and dyeing industries and textile industries as well as

analytical chemistry due to their distinctive biological activities and physico-chemical properties. However, its colouring option continues to be the most popular and widely used one. The most substantial and diverse family of dyes is the azo dye family.<sup>[1,2]</sup>

The biological activity of the azo compounds makes them appropriate for treating textiles with biocide. Azo compounds are used as anti-diabetic, antiseptic, antineoplastic, antibacterial, and anti-tumour medicines and are extensively recognised for their medicinal efficacy<sup>[29]</sup>. They are involved in a wide range of biological functions, such as nitrogen fixation, carcinogenesis, protein synthesis, and DNA and RNA inhibition.<sup>[1,23,32]</sup> It's likely that some of the Schiff bases' alleged biological activity is due to the azo-imide relationship, as azo compounds are helpful in the medical and pharmaceutical industries.<sup>[1,6,7,34]</sup>

Congo Red and Evans blue, two azo chemicals, are currently being researched as HIV viral replication inhibitors. Antibacterial and pesticidal properties are demonstrated by the presence of the azo moiety. A recent study on the use of compounds containing the azo group as antibacterial agents was published. The bulk of azo compounds are created by diazotizing a primary aromatic amine, followed by nucleophilic coupling. As a result, chemicals such as benzoic, phenolic, salicylic, and naphthol undergo diazotization reactions. It is intriguing to examine the production of those salicylic azo compounds and their derivatives in order to research the newer potentials of such compounds because of the range of applications for azo compounds.<sup>[3,11,39]</sup>

In the literature, azo compounds are occasionally referred to as chromogens. The coupling elements amino- and hydroxy-groups are frequently employed<sup>[42,49]</sup>. As a result of ongoing efforts to identify a particular dye for use in a variety of industrially significant materials, including but not restricted to paper, leather, inkjet printers, and aluminium sheet, varied classes of synthetic dyes, including azo dyes, have emerged. Additionally, azo compounds are the most common organic photoconductive compounds and have numerous uses in the photo industry, including photodynamic therapy, photographic, or electro-photographic systems.<sup>[3,8,9]</sup>

Some of the azo dyes can be used as chemotherapeutic agents and have antiseptic and antiprotozoal effects. Compared to anionic dyes, cationic dyes are more active in acidic media and prefer to attack gram-positive bacteria. Scarlet red and diamazon are the two most popular azo dyes used as antiseptics. We are encouraged to synthesize azo compounds because of their biological activity and industrial use as colourants.<sup>[4]</sup>

A white, crystalline substance with a slight acidity called aspirin has analgesic and anti-inflammatory actions.<sup>[24,25,42]</sup> Cancer and cardiovascular disease are also prevented by using it. By preventing the development of thromboxane, a substance that normally binds platelet molecules to form a patch across injured blood vessels, the substance has an antiplatelet effect.<sup>[5,12,28]</sup>

Azo chemicals can also be used to suppress the action of certain enzymes, including chymotrypsin and tyrosinase. Azo compounds, such as azo-oxyresveratrol and 1,3-indandione, as well as their derivatives, have shown strong tyrosinase inhibitory, antifungal, and antibacterial effects. Azo-salicylic acid derivatives showed that the compounds prepared and after diagnosis are not used only as dyes but can also be used in the field of medicines because of its have the

biological activity against different type of bacteria and fungi and also found through the diagnosis that these compounds prepared have high melting point that lead to high expire date.<sup>[10]</sup>

Anti-susceptibility test-The disk diffusion method is among the most flexible susceptibility testing methods in terms of antimicrobial agents that can be tested. The method consists of placing paper disks saturated with antimicrobial agents on a lawn of bacteria seeded on the surface of an agar medium, incubating the plate overnight, and measuring the presence or absence of a zone of inhibition around the disks.<sup>[13,17]</sup>

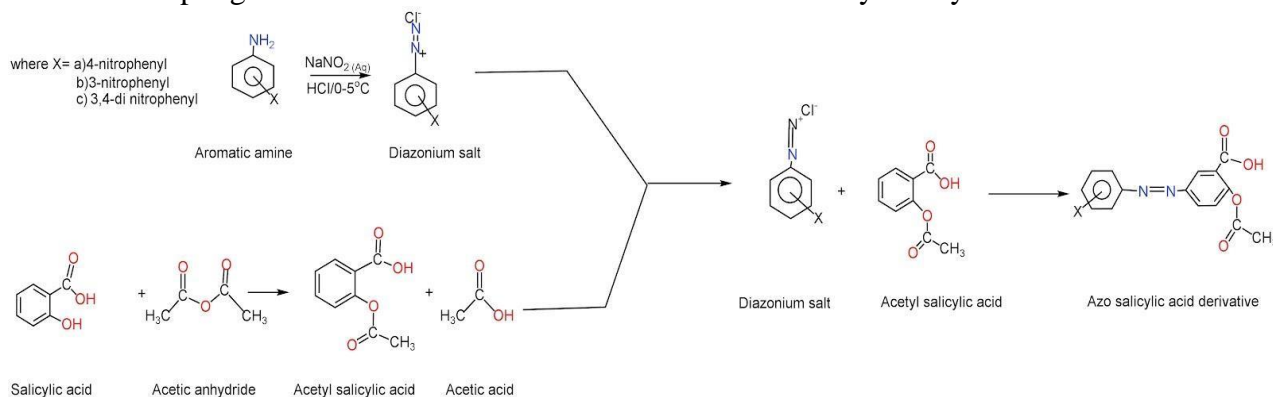
To conduct a Minimum Inhibitory Concentration (MIC) test, one introduces a substantial but imperceptible quantity of microorganisms into the sample, then observes whether the mixture of microorganisms and sample transitions from transparent to turbid. If turbidity occurs, it indicates that microorganisms have proliferated significantly, suggesting that the sample, at that specific dilution, does not inhibit their growth.<sup>[13,14,15]</sup> Clear test wells following incubation may harbor either the initial low-level microorganism inoculation or indicate that the antimicrobial agent has effectively eradicated all microorganisms. Distinguishing between these outcomes visually is not feasible.<sup>[50,51]</sup> Therefore, scientists employ MIC assays primarily to gauge the inhibitory efficacy of an antimicrobial agent rather than its biocidal properties.<sup>[16]</sup>

## MATERIALS AND METHODS

p-Nitroaniline, m-Nitroaniline, 3,4-Dinitroaniline, salicylic acid, acetic anhydride and solvents will be used in the synthesis of azo-salicylic acid derivatives. Thin layer Chromatography will be used to check the purity of the prepared compounds. Gallenkamp equipment will be used to record melting points. Shimadzu spectrometer analysis of the synthesized compounds' FT-IR spectra (KBr). The Bruker-NMR spectrometer was used to determine the proton nuclear magnetic resonance (<sup>1</sup>H-NMR) spectra.<sup>[18]</sup>

### Synthesis of Azo salicylic acid derivatives:

**Scheme:** Coupling reaction of diazonium salt derivatives with acetyl salicylic acid .

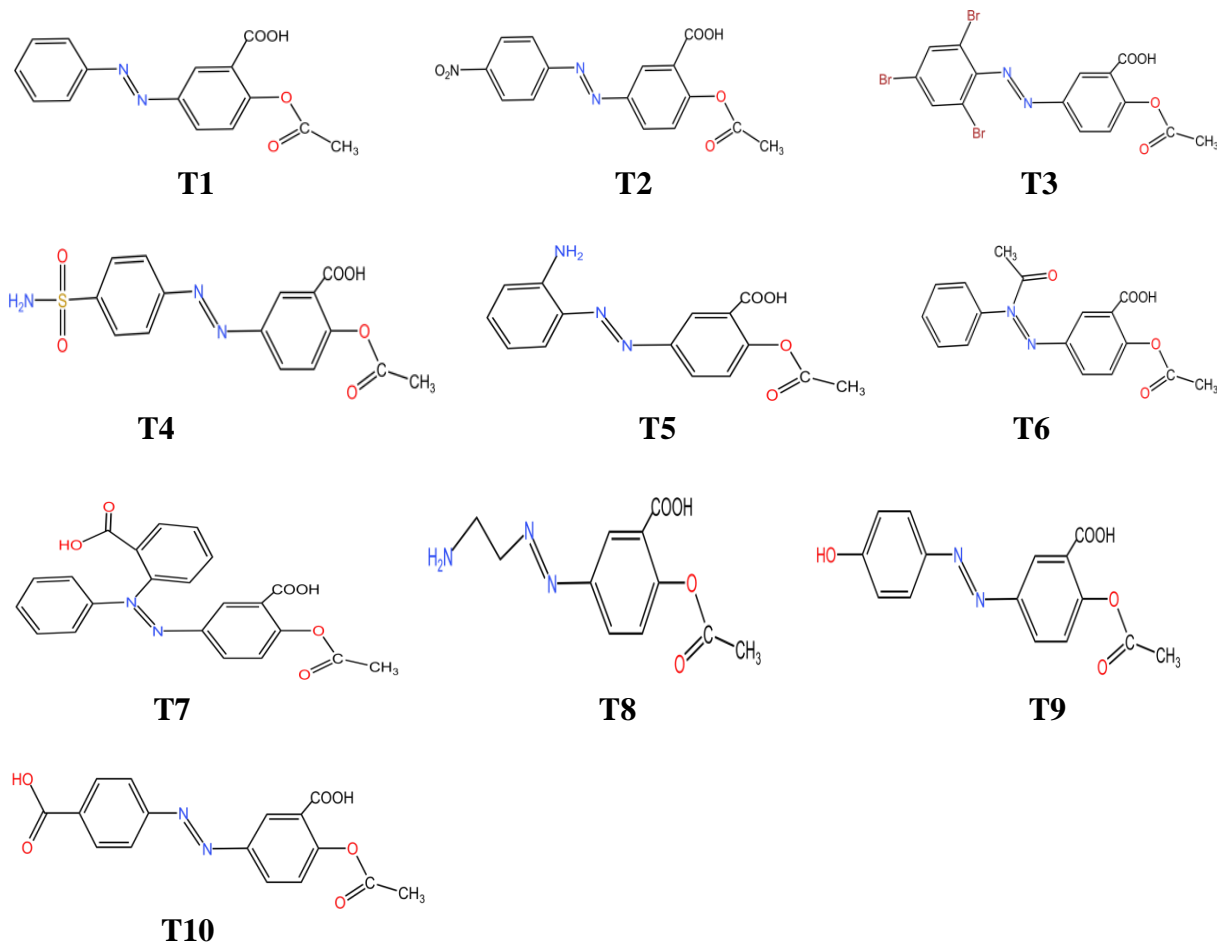


### Step.1 Formation of diazonium salt:

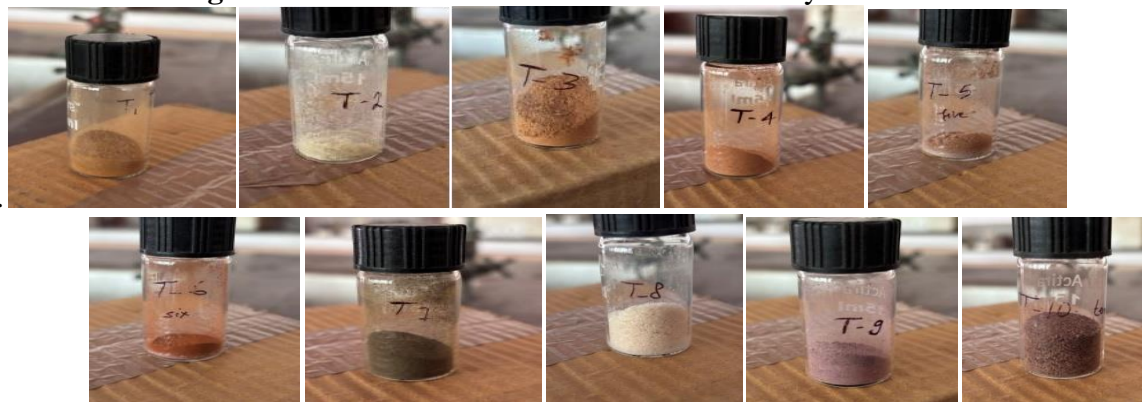
To (5 mmol) aromatic amine solution, water( 1.5ml) and conc.HCl (1.5 ml) is taken in ice-salt bath ( $0^\circ\text{C}$ ) and is kept cooled.The sodium nitrite solution (5.5 mmol) is added slowly by stirring to (1.5 ml) of water. At  $0^\circ\text{C}$ , the mixture is kept for further step. Similar steps were taken to synthesize the other diazonium salts.<sup>[3]</sup>

### Step.2 Coupling reaction with salicylic acid derivative:

To (5.4mmol) salicylic acid derivatives solution and 2.5 M aq. sodium hydroxide (10 ml), the diazonium salt solution was added in portions. For 3–5 hours, the mixture was held at 0–5 C° while being stirred. The liquid was then brought to pH 3 by adding conc. HCl (1.5 ml). The compound precipitated, separated and rinsed with water. With the help of glacial acetic acid, the required product dried and crystallized again.<sup>[3]</sup>



**Fig.1** : Name and chemical structure of azo-salicylic acid derivatives



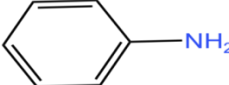
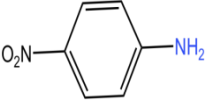
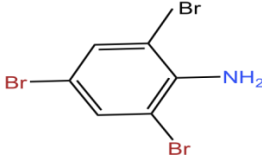
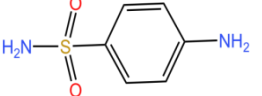
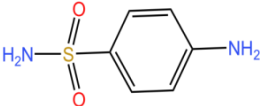
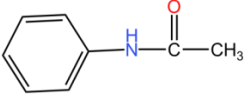
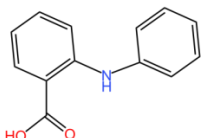
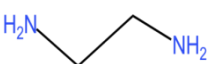
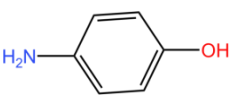
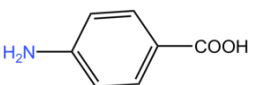
**Fig.2** : The color of synthesized derivatives.

**Table 1:** Physical properties of synthesized derivatives.

| Comp. name | Color              | Melting point | Solubility in water | Solubility in 1-butanol | Solubility in ethanol | Solubility in methanol |
|------------|--------------------|---------------|---------------------|-------------------------|-----------------------|------------------------|
| T1         | cider orange       | 220 °C        | insoluble           | highly Soluble          | highly Soluble        | highly Soluble         |
| T2         | light cider orange | 176 °C        | insoluble           | highly soluble          | highly soluble        | highly soluble         |
| T3         | clay orange        | 200 °C        | insoluble           | highly soluble          | highly soluble        | highly soluble         |
| T4         | rust orange        | 205 °C        | insoluble           | highly soluble          | highly soluble        | highly soluble         |
| T5         | cider orange       | 160 °C        | insoluble           | sparingly soluble       | highly soluble        | highly soluble         |
| T6         | ginger orange      | 180 °C        | insoluble           | highly soluble          | highly soluble        | highly soluble         |
| T7         | spice orange       | 183 °C        | insoluble           | highly soluble          | highly soluble        | highly soluble         |
| T8         | cider orange       | 179 °C        | insoluble           | sparingly soluble       | highly soluble        | highly soluble         |
| T9         | rust orange        | 124 °C        | insoluble           | sparingly soluble       | slightly soluble      | slightly soluble       |
| T10        | clay orange        | 221 °C        | insoluble           | sparingly soluble       | slightly soluble      | slightly soluble       |

**Table 2:** FT-IR absorptions of synthesized azo-salicylic acid derivatives.

| Compound code | MOLECULAR FORMULA | R1 | R2 | Mol. weight | m.p | Yield (%) | R <sub>F</sub> Value |
|---------------|-------------------|----|----|-------------|-----|-----------|----------------------|
|---------------|-------------------|----|----|-------------|-----|-----------|----------------------|

|     | (R)  |   |  |              |        |        |        |
|-----|--|---|--|--------------|--------|--------|--------|
| T1  | C <sub>15</sub> H <sub>12</sub> O <sub>4</sub> N <sub>2</sub>                    |    | C <sub>9</sub> H <sub>8</sub> O <sub>4</sub> | 284.3 g/mol  | 220 °C | 32.7%  | 0.327  |
| T2  | C <sub>15</sub> H <sub>11</sub> O <sub>6</sub> N <sub>3</sub>                    |    | C <sub>9</sub> H <sub>8</sub> O <sub>4</sub> | 329.27 g/mol | 176 °C | 54.61% | 0.272  |
| T3  | C <sub>16</sub> H <sub>12</sub> O <sub>4</sub> Br <sub>3</sub><br>N <sub>2</sub> |    | C <sub>9</sub> H <sub>8</sub> O <sub>4</sub> | 535.99 g/mol | 200 °C | 75.6%  | 0.4375 |
| T4  | C <sub>16</sub> H <sub>16</sub> O <sub>6</sub> N <sub>3</sub> S                  |    | C <sub>9</sub> H <sub>8</sub> O <sub>4</sub> | 378.38 g/mol | 205 °C | 74.4%  | 0.375  |
| T5  | C <sub>16</sub> H <sub>16</sub> O <sub>4</sub> N <sub>3</sub>                    |    | C <sub>9</sub> H <sub>8</sub> O <sub>4</sub> | 314.11 g/mol | 160 °C | 39.31% | 0.685  |
| T6  | C <sub>19</sub> H <sub>21</sub> O <sub>5</sub> N <sub>2</sub>                    |   | C <sub>9</sub> H <sub>8</sub> O <sub>4</sub> | 357.39 g/mol | 180 °C | 43.18% | 0.555  |
| T7  | C <sub>24</sub> H <sub>24</sub> O <sub>6</sub> N <sub>2</sub>                    |  | C <sub>9</sub> H <sub>8</sub> O <sub>4</sub> | 436.46 g/mol | 183 °C | 55.88% | 0.621  |
| T8  | C <sub>12</sub> H <sub>16</sub> O <sub>4</sub> N <sub>3</sub>                    |  | C <sub>9</sub> H <sub>8</sub> O <sub>4</sub> | 266.28 g/mol | 179 °C | 25.73% | 0.59   |
| T9  | C <sub>16</sub> H <sub>15</sub> O <sub>5</sub> N <sub>2</sub>                    |  | C <sub>9</sub> H <sub>8</sub> O <sub>4</sub> | 315.3 g/mol  | 124 °C | 39.58% | 0.635  |
| T10 | C <sub>17</sub> H <sub>15</sub> O <sub>6</sub> N <sub>2</sub>                    |  | C <sub>9</sub> H <sub>8</sub> O <sub>4</sub> | 343.32 g/mol | 221 °C | 45.6%  | 0.596  |

## RESULTS AND DISCUSSION

The new azo dye compounds were synthesized by coupling a diazonium salt with salicylic acid, as illustrated in Scheme (1). The reaction was conducted under ice-cold conditions (0-5°C). The structures assigned were confirmed by consistent results from FTIR, <sup>13</sup>CNMR, LCMS, UV, and Elemental analysis.<sup>[19]</sup>

### COMPOUND T1: (E)-2-acetoxy-5-(phenyldiazenyl)benzoic acid

Yield: % (Cider Orange); M.P: 220-224 °C; R<sub>f</sub>: 0.327(Chloroform : acetone : n-butanol : 88% Formic acid 4:1:1:1); UV(λ<sub>max</sub>): 370nm; IR spectrum(cm<sup>-1</sup>):3775.29 cm<sup>-1</sup>(NH stretching), 3912.56 cm<sup>-1</sup> (OH stretching), 2930.81 cm<sup>-1</sup> ( CH stretching), 2348.90 cm<sup>-1</sup> asymmetrical stretching (CN stretching ), 1845.65 cm<sup>-1</sup> (CO stretching), 836.46 cm<sup>-1</sup> (Aro); <sup>13</sup>CNMR (500MHz, Methanol): 40.94(s),102(s),113, 146.53, 154(s),165.87(s); Mass spectrum (m/z): 284 (M<sup>+</sup> ),224,114; UV(λ<sub>max</sub>): 342nm; CHN calculated for C<sub>15</sub>H<sub>12</sub>O<sub>4</sub>N<sub>2</sub> (284.3): C:63.38% ,H:4.26%,O:22.51% ,N:9.85%

**Compound T2: (E)-2-acetoxy-5-((4-nitrophenyl)diazenyl)benzoic acid**

Yield: % (Light cider Orange); M.P: 176 °C; R<sub>f</sub>: 0.272(Chloroform : acetone : n-butanol : 88% Formic acid 4:1:1:1); UV(λ<sub>max</sub>): 224nm; IR spectrum(cm<sup>-1</sup>):3775.29 cm<sup>-1</sup>(NH stretching), 3912.56 cm<sup>-1</sup> (OH stretching), 2930.81 cm<sup>-1</sup> ( CH stretching), 2423.70 cm<sup>-1</sup> asymmetrical stretching (CN stretching ), 1845.65 cm<sup>-1</sup> (CO stretching), 764 cm<sup>-1</sup> (Aro); <sup>13</sup>CNMR (500MHz, Methanol): 42(s),114,132,138(s),141, 150(d),156(s); Mass spectrum (m/z): 329 (M<sup>+</sup> ),248,102,48; UV(λ<sub>max</sub>): 342nm; CHN calculated for C<sub>15</sub>H<sub>11</sub>O<sub>6</sub>N<sub>3</sub> (329.27): C:54.72% ,H:3.37%,O:29.15% ,N:12.76%

**COMPOUND T3: (E)-2-acetoxy-5-((2,4,6-tribromophenyl)diazenyl)benzoic acid**

Yield: % (Clay Orange); M.P: 200 °C; R<sub>f</sub>: 0.4375(Chloroform : acetone : n-butanol : 88% Formic acid 4:1:1:1); UV(λ<sub>max</sub>): 214nm; IR spectrum(cm<sup>-1</sup>): 3912.75 cm<sup>-1</sup> (OH stretching), 3403.61 cm<sup>-1</sup>(NH stretching), 2860.54 cm<sup>-1</sup> ( CH stretching), 2341.65 cm<sup>-1</sup> asymmetrical stretching (CN stretching ), 1845.65 cm<sup>-1</sup> (CO stretching),756.12 (C-Br stretching )826.3cm<sup>-1</sup> (Aro); <sup>13</sup>CNMR (500MHz, Methanol): 51(s),118, 121,128(s), 137(d),142(d),165(s); Mass spectrum (m/z): 534(M<sup>+</sup> ),224,156,48; UV(λ<sub>max</sub>): 342nm; CHN calculated for C<sub>16</sub>H<sub>12</sub>O<sub>4</sub>Br<sub>3</sub>N<sub>2</sub> (535.99): C:35.85% ,H:2.26%,O:11.94% ,N:5.23,Br:44.72%

**COMPOUND T4: (E)-2-acetoxy-5-((4-sulfamoylphenyl)diazenyl)benzoic acid**

Yield: % (Rust Orange); M.P: 205 °C; R<sub>f</sub>: 0.375(Chloroform : acetone : n-butanol : 88% Formic acid 4:1:1:1); UV(λ<sub>max</sub>): 352nm; IR spectrum(cm<sup>-1</sup>): 3912.75 cm<sup>-1</sup> (OH stretching),3403.61 cm<sup>-1</sup>(NH stretching), 2921.71 cm<sup>-1</sup> ( CH stretching), 2596.69 (SH stretching) , 2356.80cm<sup>-1</sup> asymmetrical stretching (CN stretching ), 1662.34 cm<sup>-1</sup> (CO stretching), 756.77 cm<sup>-1</sup> (Aro); <sup>13</sup>CNMR (500MHz, Methanol): 47(s),115,129,131,138,142(d),166(s); Mass spectrum (m/z): 378 (M<sup>+</sup> ),248,114,48; UV(λ<sub>max</sub>): 342nm; CHN calculated for C<sub>16</sub>H<sub>16</sub>O<sub>6</sub>N<sub>3</sub>S (378.38): C:50.79% ,H:4.26%,O:25.37% ,N:11.11%,S:8.47%

**COMPOUND T5: (E)-2-acetoxy-5-((2-aminophenyl)diazenyl)benzoic acid**

Yield: % (Cider Orange); M.P: 160 °C; R<sub>f</sub>: 0.685(Chloroform : acetone : n-butanol : 88% Formic acid 4:1:1:1); UV(λ<sub>max</sub>): 364nm; IR spectrum(cm<sup>-1</sup>): 3320.22 cm<sup>-1</sup> (OH stretching), 2917.60 cm<sup>-1</sup> ( CH stretching), 1748.09 cm<sup>-1</sup> (CO stretching), 3513.12cm<sup>-1</sup>(NH stretching), 2350.38 cm<sup>-1</sup> asymmetrical stretching (CN stretching ), 528.87 cm<sup>-1</sup> (Aro); <sup>13</sup>CNMR (500MHz, Methanol): 52(s),114,125(d),129(s),139,146,161(s); Mass spectrum (m/z): 314 (M<sup>+</sup> ),224,114; UV(λ<sub>max</sub>): 342nm; CHN calculated for C<sub>16</sub>H<sub>16</sub>O<sub>4</sub>N<sub>3</sub> (314.32): C:61.14% ,H:5.13%,O:20.36% ,N:13.37%

**COMPOUND T6: HO-(Z)-2-acetoxy-5-(2-acetyl-2-phenyl-2λ<sup>4</sup>-diazenyl)benzoic acid**

Yield: % (Ginger Orange); M.P: 180 °C; R<sub>f</sub>: 0.555(Chloroform : acetone : n-butanol : 88% Formic acid 4:1:1:1); UV( $\lambda_{\max}$ ): 205nm; IR spectrum( $\text{cm}^{-1}$ ):3351.12  $\text{cm}^{-1}$ (NH stretching), 3502.12 $\text{cm}^{-1}$  (OH stretching), 2881.3  $\text{cm}^{-1}$  ( CH stretching), 2347.30 $\text{cm}^{-1}$  asymmetrical stretching (CN stretching ), 1810  $\text{cm}^{-1}$  (CO stretching), 882.5  $\text{cm}^{-1}$  (Aro); <sup>13</sup>CNMR (500MHz, Methanol): 44(s),118,121,134(s),136,153(d),162(s); Mass spectrum (m/z): 357 (M<sup>+</sup> ),241,102,48; UV( $\lambda_{\max}$ ): 342nm; CHN calculated for C<sub>19</sub>H<sub>21</sub>O<sub>5</sub>N<sub>2</sub> (357.39): C:63.86% ,H:5.92%,O:22.38% ,N:7.84%

**COMPOUND T7: (2)-2-acetoxy-5-12-12-carboxyphenyl)-2-phenyl-2 $\lambda^4$ -diazenyl)benzoic acid**

Yield: % (Spice Orange); M.P: 183°C; R<sub>f</sub>: 0.621(Chloroform : acetone : n-butanol : 88% Formic acid 4:1:1:1); UV( $\lambda_{\max}$ ): 224nm; IR spectrum( $\text{cm}^{-1}$ ) : 2812.33  $\text{cm}^{-1}$  ( CH stretching), 3697.14 $\text{cm}^{-1}$ (NH stretching), 3596.15 $\text{cm}^{-1}$  (OH stretching), 2323.64 $\text{cm}^{-1}$  asymmetrical stretching (CN stretching ), 1804.21  $\text{cm}^{-1}$  (CO stretching), 821.8 $\text{cm}^{-1}$  (Aro); <sup>13</sup>CNMR (500MHz, Methanol): 51(s),119,125,133(d),138,139(s),148,166(s); Mass spectrum (m/z): 436 (M<sup>+</sup> ),284,114,48; UV( $\lambda_{\max}$ ): 342nm; CHN calculated for C<sub>24</sub>H<sub>24</sub>O<sub>6</sub>N<sub>2</sub> (436.46): C:66.05% ,H:5.54%,O:21.99% ,N:6.42%

**COMPOUND T8: (E)-2-acetoxy-5-((2-aminoethyl)diazenyl)benzoic acid**

Yield: % (Cider Orange); M.P: 179 °C; R<sub>f</sub>: 0.590(Chloroform : acetone : n-butanol : 88% Formic acid 4:1:1:1); UV( $\lambda_{\max}$ ): 305nm; IR spectrum( $\text{cm}^{-1}$ ): 3912.41  $\text{cm}^{-1}$  (OH stretching), 3233.88  $\text{cm}^{-1}$ (NH stretching), 2919.82  $\text{cm}^{-1}$  ( CH stretching), 2303.29  $\text{cm}^{-1}$  asymmetrical stretching (CN stretching ), 1751.37  $\text{cm}^{-1}$  (CO stretching), 882.13  $\text{cm}^{-1}$  (Aro); <sup>13</sup>CNMR (500MHz, Methanol): 55(s),119(d),122(s),136,139,145(d),161(s),165(s); Mass spectrum (m/z): 266 (M<sup>+</sup> ),214,48; UV( $\lambda_{\max}$ ): 342nm; CHN calculated for C<sub>12</sub>H<sub>16</sub>O<sub>4</sub>N<sub>3</sub> (266.28): C:54.13% ,H:6.06%,O:24.03% ,N:15.78%

**COMPOUND T9: (E)-2-acetoxy-5-((4-hydroxyphenyl)diazenyl)benzoic acid**

Yield: % (Rust Orange); M.P: 124 °C; R<sub>f</sub>: 0.635(Chloroform : acetone : n-butanol : 88% Formic acid 4:1:1:1); UV( $\lambda_{\max}$ ): 242nm; IR spectrum( $\text{cm}^{-1}$ ): 37743.81 $\text{cm}^{-1}$  (OH stretching), 3775.29  $\text{cm}^{-1}$ (NH stretching), 2922.13  $\text{cm}^{-1}$  ( CH stretching), 2348.90  $\text{cm}^{-1}$  asymmetrical stretching (CN stretching ), 1664.23 $\text{cm}^{-1}$  (CO stretching), 813.66  $\text{cm}^{-1}$  (Aro); <sup>13</sup>CNMR (500MHz, Methanol): 48(s),112,119(d),127,135,144(d),161(s),164(s); Mass spectrum (m/z): 315 (M<sup>+</sup> ),224,48; UV( $\lambda_{\max}$ ): 342nm; CHN calculated for C<sub>16</sub>H<sub>15</sub>O<sub>5</sub>N<sub>2</sub> (315.3): C:60.95% ,H:4.8%,O:25.37% ,N:8.88%

**COMPOUND T10: (E)-2-acetoxy-5-((4-carboxyphenyl)diazenyl)benzoic acid**

Yield: % (Clay Orange); M.P: 221 °C; R<sub>f</sub>: 0.596(Chloroform : acetone : n-butanol : 88% Formic acid 4:1:1:1); UV( $\lambda_{\max}$ ): 352nm; IR spectrum( $\text{cm}^{-1}$ ): 3720.58  $\text{cm}^{-1}$  (OH stretching), 3238.46  $\text{cm}^{-1}$ (NH stretching), 2921.71  $\text{cm}^{-1}$  ( CH stretching), 2348.90  $\text{cm}^{-1}$  asymmetrical stretching (CN stretching ), 1635.49  $\text{cm}^{-1}$  (CO stretching), 786.46  $\text{cm}^{-1}$  (Aro); <sup>13</sup>CNMR (500MHz, Methanol): 50(s),116(s),118,134,139,142,165,169(s); Mass spectrum (m/z): 344 (M<sup>+</sup> ),248,114; UV( $\lambda_{\max}$ ): 342nm; CHN calculated for C<sub>17</sub>H<sub>15</sub>O<sub>6</sub>N<sub>2</sub> (343.32): C:59.48% ,H:4.4%,O:27.96% ,N:8.16%



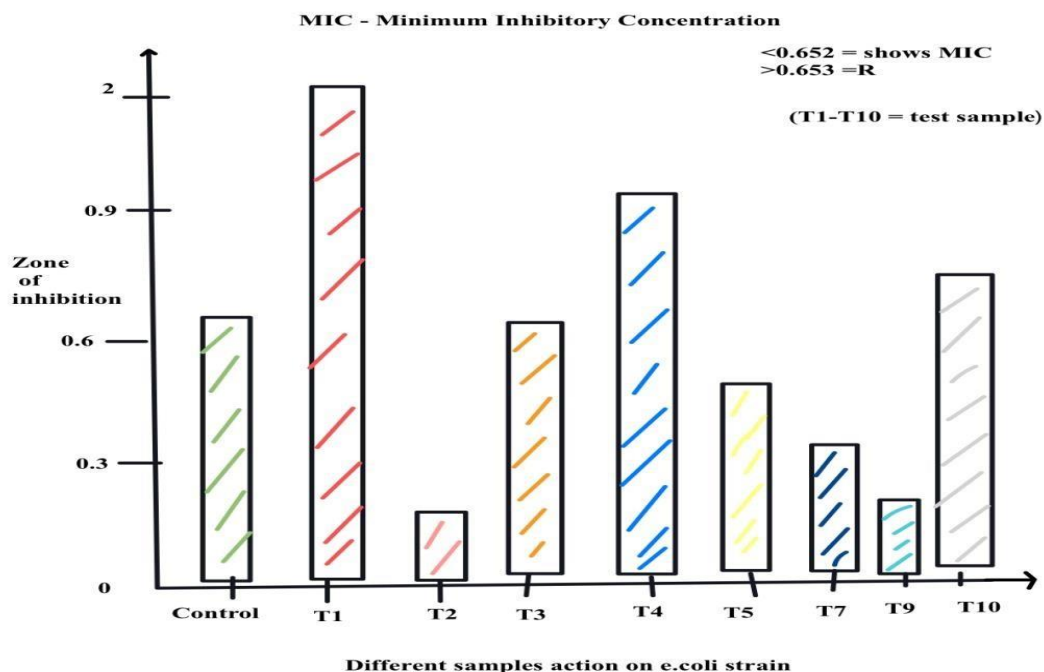
**Table 3:** Biological activity of synthesized azo-salicylic acid derivatives with gentamicin as standard.

| Azo dye compounds | Escherichia Coli | Candida Albicans | klebsiella Sp. |
|-------------------|------------------|------------------|----------------|
| T1                | R                | 13mm             | 13mm           |
| T2                | 16mm             | 24mm             | 17mm           |
| T3                | 16mm             | 28mm             | 24mm           |
| T4                | 11mm             | 15mm             | 11mm           |
| T5                | 10mm             | 10mm             | 12mm           |
| T6                | 12mm             | 12mm             | 13mm           |
| T7                | 14mm             | R                | 17mm           |
| T8                | R                | 12mm             | 32mm           |
| T9                | 18mm             | 15mm             | 12mm           |
| T10               | 10mm             | R                | R              |

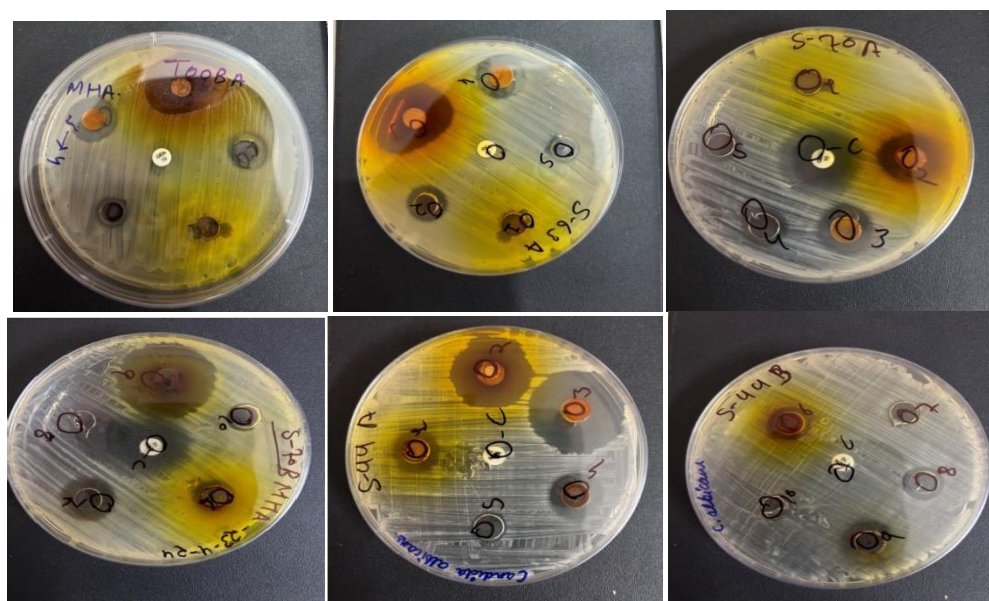
**Table 4: Minimum Inhibitory Concentration**

| S.No. | CONTROL  | TEST(O.D) | MIC |
|-------|----------|-----------|-----|
| T1    | 0.652 nm | 2.990 nm  | R   |
| T2    | 0.652 nm | 0.093 nm  | MIC |
| T3    | 0.652 nm | 0.641 nm  | MIC |
| T4    | 0.652 nm | 1.0889 nm | R   |
| T5    | 0.652 nm | 0.546 nm  | MIC |
| T7    | 0.652 nm | 0.36 nm   | MIC |
| T9    | 0.652 nm | 0.151 nm  | MIC |
| T10   | 0.652 nm | 0.713 nm  | R   |

R= Resistance, MIC=Minimum Inhibitory Concentration, O.D= Optimum Density



**Biological activity** Anti-microbial activity: Anti-bacterial activity will be carried out by Agar diffusion method of prepared azo dye compounds. Biological activity of prepared azo dye compounds were determined with gentamicin as standard, by agar diffusion method, all azo dye compounds were tested and the plates were incubated at 37°C for 24 hours, the inhibition zone measured in (mm). Azo dye compound were evaluated for antibacterial activity against different bacterial strain, (*Staphylococcus aureus*, *Staphylococcus epidermidis*, *Escherichia coli*, *Klebsiella sp.*) and fungi such as (*Candida albicans*).<sup>[3]</sup>



**Fig.6 :** Agar diffusion method of synthesized azo-salicylic acid derivatives.

The results presented in Tables 3 and 4 indicate that several azo dye compounds (T2, T3, and T8) exhibit higher biological activity compared to others, approaching the effectiveness of the standard antibiotic Gentamicin. Conversely, compounds T1 and T10 demonstrate lower biological activity relative to the rest. Specifically, compound T1 shows activity against yeast and gram-negative bacteria such as *Candida Albicans* and *Klebsiella* sp., while compound T10 is effective only against *Escherichia coli*. Compound T7 is resistant to yeast (*Candida Albicans*) but shows biological activity against both gram-negative bacteria. Compounds T2, T3, T5, T7, and T9 demonstrate minimum inhibitory concentrations (MIC).

### CONCLUSION

Based on the antimicrobial evaluation of azo dye compounds through agar diffusion and Minimum Inhibitory Concentration (MIC) assays, significant findings have emerged regarding their biological activity. The compounds T2, T3, and T8 displayed notable antibacterial efficacy, comparable to the standard antibiotic Gentamicin, against a range of bacterial strains including *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Escherichia coli*, and *Klebsiella* species. Moreover, these compounds exhibited varying degrees of effectiveness against the yeast *Candida albicans*, underscoring their potential as broad-spectrum antimicrobial agents. Conversely, compounds T1 and T10 demonstrated lower biological activity, with T1 showing effectiveness primarily against *Candida albicans* and *Klebsiella* species, while T10 exhibited activity solely against *Escherichia coli*. Compound T7 displayed resistance against *Candida albicans* but demonstrated biological activity against gram-negative bacteria. Notably, compounds T2, T3, T5, T7, and T9 revealed specific Minimum Inhibitory Concentrations (MIC), indicating their potency in inhibiting microbial growth.

Overall, the observed antimicrobial properties of these azo dye compounds highlight their potential for diverse industrial and medical applications. Further research into their structure-activity relationships and pharmacological profiles could pave the way for their development as novel therapeutic agents against infectious diseases.

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### REFERENCES

1. Aljamali, N. M. (2015). Review in azo compounds and its biological activity. *Biochem Anal Biochem*, 4(2), 1-4.
2. Clarke, H. T., and W. R. Kirner. 1941. Methyl Red. *Org. Synth. Coll. Vol. 1*: 374. Retrieved January 8, 2009
3. Ibrahim, W. A., Farhan, M. A., & Abdulateef, M. H. Synthesis and evaluation of biological activity of some newsalicylic acid derivatives. *Biochem. Cell. Arch*, 20(9), 3727-3732
4. Salman, H. H., AbOOD, H. S., & Ramadhan, U. H. (2019). Synthesis of Some New Azo Compounds of Salicylic Acid Derivatives and Determine Their In Vitro Anti-Inflammatory Activity. *Oriental Journal of Chemistry*, 35(2), 870
5. Aljamali, N. M. (2015). Review in azo compounds and its biological activity. *Biochem Anal Biochem*, 4(2), 1-4

6. Patil, C. J., & Nehete, C. A. (2015). The azo derivatives of salicylic acid. *International journal of pharmaceutical sciences review and research*, 33(2), 248-256
7. Naseem, H. A., Aziz, T., Ahmad, K., Parveen, S., & Ashfaq, M. (2021). Rational synthesis and characterization of medicinal phenyl diazenyl-3-hydroxy-1h-inden-1-one azo derivatives and their metal complexes. *Journal of molecular structure*, 1227, 129574.
8. Tsui, Y. K., Devaraj, S., & Yen, Y. P. (2012). Azo dyes featuring with nitrobenzoxadiazole (NBD) unit: a new selective chromogenic and fluorogenic sensor for cyanide ion. *Sensors and Actuators B: Chemical*, 161(1), 510-519.
9. Mohammed, G. I., El-Ghamry, H. A., & Saber, A. L. (2021). Rapid, sensitive, and selective copper (II) determination using sensitive chromogenic azo dye based on sulfonamide. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 247, 119103.
10. Pearson DC, Jourdeuil D, Meddings JB. The anti-oxidant properties of 5-aminosalicylic acid. *Free Radic Biol Med*. 1996;21(3):367-73.
11. Piotto, S., Concilio, S., Sessa, L., Diana, R., Torrens, G., Juan, C., ... & Iannelli, P. (2017). Synthesis and antimicrobial studies of new antibacterial azo-compounds active against staphylococcus aureus and listeria monocytogenes. *Molecules*, 22(8), 1372.
12. Gurbel, P. A., Bliden, K. P., DiChiara, J., Newcomer, J., Weng, W., Neerchal, N. K., ... & Tantry, U. S. (2007). Evaluation of dose-related effects of aspirin on platelet function: results from the Aspirin-Induced Platelet Effect (ASPECT) study. *Circulation*, 115(25), 3156-3164.
13. Wiegand, I., Hilpert, K., & Hancock, R. E. (2008). Agar and broth dilution methods to determine the minimal inhibitory concentration (MIC) of antimicrobial substances. *Nature protocols*, 3(2), 163-175.
14. Miller, R. A., Walker, R. D., Carson, J., Coles, M., Coyne, R., Dalsgaard, I., ... & Reimschuessel, R. (2005). Standardization of a broth microdilution susceptibility testing method to determine minimum inhibitory concentrations of aquatic bacteria. *Diseases of aquatic organisms*, 64(3), 211-222.
15. Wilkins, T. D., & Thiel, T. (1973). Modified broth-disk method for testing the antibiotic susceptibility of anaerobic bacteria. *Antimicrobial agents and chemotherapy*, 3(3), 350-356.
16. Mohsenzadeh, M. (2007). Evaluation of antibacterial activity of selected Iranian essential oils against Staphylococcus aureus and Escherichia coli in nutrient broth medium. *Pak J Biol Sci*, 10(20), 3693-3697.
17. Lubber, P., Bartelt, E., Genschow, E., Wagner, J., & Hahn, H. (2003). Comparison of broth microdilution, E test, and agar dilution methods for antibiotic susceptibility testing of Campylobacter jejuni and Campylobacter coli. *Journal of clinical microbiology*, 41(3), 1062-1068.
18. Wadher, S. J., Karande, N. A., Sonawane, S. D., & Yeole, P. G. (2009). Synthesis and biological evaluation of schiff base and 4-thiazolidinones of amino salicylic acid and their derivatives as an antimicrobial agent. *International Journal of ChemTech Research*, 1(4), 1303-1307
19. Gupta, P. K., Gupta, S., Singh, S. D., & Tiwari, S. B. SYNTHESIS, MOLECULAR DOCKING AND EVALUATION STUDIES OF NOVEL 2-(N-PHENYL SUBSTITUTED)-3-ALKYL AMINO-QUINAZOLINE-4 (3H)-ONE DERIVATIVES FOR ANTI-INFLAMMATORY & ANTICONVULSANT ACTIVITIES.

20. El-Ghamaz, N. A., El-Sonbati, A. Z., & Serag, L. S. (2018). Linear and nonlinear optical properties of new azo aminosalicylic acid derivatives. *Journal of Luminescence*, 194, 507-518.
21. Garjani, A., Davaran, S., Rashidi, M., & Malek, N. (2004). Protective effects of some azo derivatives of 5-aminosalicylic acid and their pegylated prodrugs on acetic acid-induced rat colitis. *DARU Journal of Pharmaceutical Sciences*, 12(1), 24-30.
22. Carceller, E., Salas, J., Merlos, M., Giral, M., Ferrando, R., Escamilla, I., ... & Forn, J. (2001). Novel azo derivatives as prodrugs of 5-aminosalicylic acid and amino derivatives with potent platelet activating factor antagonist activity. *Journal of medicinal chemistry*, 44(18), 3001-3013.
23. Awad, I. M. (1991). Synthesis of some new azosulphonamides based on salicylic acid and thiosalicylic acid, and having antibacterial and antifungal activity. *Dyes and pigments*, 17(2), 123-139.
24. Teimouri, A., Chermahini, A. N., & Ghorbani, M. H. (2013). The green synthesis of new azo dyes derived from salicylic acid derivatives catalyzed via baker's yeast and solid acid catalysis. *chemija*, 24(1), 59-66.
25. Djurendić, E., Vujašković, S. D., Sakač, M., Ajduković, J., Gaković, A., Kojić, V., ... & Gaši, K. P. (2011). Synthesis and biological evaluation of some new 2-oxazoline and salicylic acid derivatives. *ARKIVOC: Online Journal of Organic Chemistry*.
26. Hassan, G. S., & Soliman, G. A. (2010). Design, synthesis and anti-ulcerogenic effect of some of furo-salicylic acid derivatives on acetic acid-induced ulcerative colitis. *European journal of medicinal chemistry*, 45(9), 4104-4112.
27. Hassan, G. S., & Soliman, G. A. (2010). Design, synthesis and anti-ulcerogenic effect of some of furo-salicylic acid derivatives on acetic acid-induced ulcerative colitis. *European journal of medicinal chemistry*, 45(9), 4104-4112.
28. Sahoo, J., & Paidesetty, S. K. (2015). Antimicrobial, analgesic, antioxidant and in silico study of synthesized salicylic acid congeners and their structural interpretation. *Egyptian Journal of Basic and Applied Sciences*, 2(4), 268-280.
29. Ekinci, D., Şentürk, M., & Küfrevioğlu, Ö. İ. (2011). Salicylic acid derivatives: synthesis, features and usage as therapeutic tools. *Expert opinion on therapeutic patents*, 21(12), 1831-1841.
30. Mohammed, H. A., & Kadem, K. J. (2022). Synthesis Characterization and Biological Evaluation of Some New Azo-Schiff compounds Derived from 5-aminosalicylic acids. *HIV Nursing*, 22(2), 110-112.
31. Dabbagh, H. A., Teimouri, A., Chermahini, A. N., & Shahraki, M. (2008). DFT and ab initio study of structure of dyes derived from 2-hydroxy and 2, 4-dihydroxy benzoic acids. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 69(2), 449-459.
32. Boumya, W., Laghrib, F., Lahrich, S., Farahi, A., Achak, M., Bakasse, M., & El Mhammedi, M. A. (2018). Electrochemical behavior study of salicylic acid following azo dye formation with 2, 4-dinitrophenylhydrazine: Analytical evaluation. *South African Journal of Chemical Engineering*, 25, 48-53
33. . Abdu-Allah, H. H., El-Shorbaji, A. N., Abdel-Moty, S. G., El-Awady, R., & Abdel-Alim, A. A. (2016). 5-Aminosalicylic acid (5-ASA): a unique anti-inflammatory salicylate. *Med Chem (Los Angeles)*, 6(5), 306-15.

34. El-Sayed, Y., Gaber, M., El-Wakeil, N., Abdelaziz, A., & El-Nagar, A. (2021). Metal complexes of azo mesalamine drug: Synthesis, characterization, and their application as an inhibitor of pathogenic fungi. *Applied Organometallic Chemistry*, 35(8), e6290.
35. Aljaber, A. S., & Bani-Yaseen, A. D. (2019). Computational exploration of the effect of molecular medium on the tautomerization of azo prodrug of 5-aminosalicylic acid. *Journal of Molecular Graphics and Modelling*, 86, 160-169.
36. Mutar, M. A., Kmal, R. K., & Jawad, S. F. (2014). Synthesis and characterization of new biodegradable polyurethanes containing azo derivatives of 5-aminosalicylic acid. *Iraqi National Journal of Chemistry*, 14(55), 275-300.
37. Tonle, I. K., & Kuateb, J. R. (2019). Synthesis, characterization, antimicrobial activities and electrochemical behavior of new phenolic azo dyes from two thienocoumarin amines. *Organic Chemistry*, (part vi), 0-0.
38. Callant, D., & Schacht, E. (1990). Macromolecular prodrugs of 5-aminosalicylic acid, 1. Azo-conjugates. *Die Makromolekulare Chemie: Macromolecular Chemistry and Physics*, 191(3), 529-536.
39. Bawa, R. A., & Alzaraide, E. M. (2005). Antifungal study of two synthesized phenolic azo derivatives. *Organic Chemistry*, 11(1), 4-11.
40. Jasim, E. Q., Muhammad-Ali, M. A., & Almakki, A. (2023). Synthesis, characterization, and antibacterial activity of some mesalazine derivatives. *Science and Technology Indonesia*, 8(3), 338-343.
41. Kadhim, Z. Y., Seewan, A. N., Abd, M. T., & Saud, H. R. (2020). Synthesis, Characterization, Antibacterial Screening and application on the wool fabric of new Bis-azo Compounds derived from 4, 4'-Diaminodiphenylmethane. *International Journal of Pharmaceutical Research*, 12(3), 402-407.
42. Allen, R. L. M. (1971). The chemistry of azo dyes. In *Colour chemistry* (pp. 21-36). Boston, MA: Springer US.
43. Stevenson, S. G., & Resuggan, J. C. L. (1938). A colorimetric test for the detection of para-hydroxybenzoic acid in the presence of salicylic acid. *Analyst*, 63(744), 152-155.
44. Verma, R. K., & Chaurasia, L. (2005). Synthesis and antifungal activity studies of some pyrazolopyrimidine derivatives. *JOURNAL-INDIAN CHEMICAL SOCIETY*, 82(7), 665.
45. Hewitt, J. T., & Fox, J. J. (1901). II.—The nitration of benzeneazosalicylic acid. *Journal of the Chemical Society, Transactions*, 79, 49-53.
46. Owuama, C. I. (2017). Determination of minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) using a novel dilution tube method. *African journal of microbiology research*, 11(23), 977-980.
47. Rodríguez-Melcón, C., Alonso-Calleja, C., García-Fernández, C., Carballo, J., & Capita, R. (2021). Minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) for twelve antimicrobials (biocides and antibiotics) in eight strains of *Listeria monocytogenes*. *Biology*, 11(1), 46.
48. Andrews, J. M. (2001). Determination of minimum inhibitory concentrations. *Journal of antimicrobial Chemotherapy*, 48(suppl\_1), 5-16.
49. Mouton, J. W., & Vinks, A. A. (2005). Relationship between minimum inhibitory concentration and stationary concentration revisited: growth rates and minimum bactericidal concentrations. *Clinical pharmacokinetics*, 44, 767-768.

50. Kowalska-Krochmal, B., & Dudek-Wicher, R. (2021). The minimum inhibitory concentration of antibiotics: Methods, interpretation, clinical relevance. *Pathogens*, *10*(2), 165.
51. Reimer, L. G., Stratton, C. W., & Reller, L. B. (1981). Minimum inhibitory and bactericidal concentrations of 44 antimicrobial agents against three standard control strains in broth with and without human serum. *Antimicrobial agents and chemotherapy*, *19*(6), 1050-1055.
52. Jacob, J. T., & DiazGranados, C. A. (2013). High vancomycin minimum inhibitory concentration and clinical outcomes in adults with methicillin-resistant *Staphylococcus aureus* infections: a meta-analysis. *International Journal of Infectious Diseases*, *17*(2), e93-e100.