Kumari Sonu / Afr.J.Bio.Sc. 6(9) (2024)

ISSN: 2663-2187

https://doi.org/10.33472/AFJBS.6.9.2024.5199-5209



Article History

Volume 6, Issue 9, 2024

Received: 26-04-2024

Accepted : 29-05-2024

doi: 10.33472/AFJBS.6.9.2024.5199-5209

A very accurate, precise, and reliable isocratic HPLC technique has been developed and verified for the simultaneous quantification of Rifampicin and Ofloxacin. The chromatographic separation was performed using a Phenomenex C18 column with a mobile phase consisting of a combination of 0.05M KH₂PO₄ buffer at pH 4.0 and methanol in a ratio of 55:35. The separation was monitored at a wavelength of 230 nm. Ofloxacin had a retention time of 4.920 min, whereas Rifampicin had a retention time of 11.710 min. The calibration plots for Rifampicin and Ofloxacin were linear between the concentration ranges of 2-10 µg/ml and 4–20 µg/ml, respectively. The correlation coefficient for Rifampicin was 0.9999, while for Ofloxacin it was 0.9996. The Rifampicin had a limit of detection (LOD) of 0.2345 µg/ml, whereas the Ofloxacin had a LOD of 0.4625 µg/ml.The approach underwent validation to assess its linearity, accuracy, precision, and robustness. The percentage of recoveries was determined to be about 100% with little fluctuation. This approach can be used for regular analysis. Keywords: Rifampicin, Ofloxacin and RP-HPLC.

Introduction:

TB treatment has grown more complicated due to the rise of multidrug-resistant (MDR) and extensively drug-resistant Mycobacterium tuberculosis strains, a worldwide challenge. In 2015, 10.4 million new cases of tuberculosis (TB) and 480,000 MDR-TB were recorded globally. TB killed nearly 1.4 million people globally in 2015 [1]. The prolonged period of therapy, hepatotoxicity, many side effects of conventional drugs, and early treatment termination make current tuberculosis treatment ineffective [2]. Thus, combination therapy reduces resistance by lowering treatment time, highlighting the need for new drug combinations. Streptomyces mediterranei produces rifampicin (RIF), a rifamycin antibiotic [3]. Complex semisynthetic macrocyclic chemical used to treat TB and other infectious disorders [4–7]. It is a key antituberculous agent. RNA production decreases and cell death occurs when RIF inhibits DNA-dependent RNA polymerase. This medicine only treats mycobacterial infections owing to the fast spread of drug-resistant microorganisms. Combining therapies delays resistance development in some circumstances. The drug also treats asymptomatic meningococcal carriers. Fluoroquinolones like ofloxacin (OFL) are now a common and promising drug-resistant therapy [9]. Clinical data suggests that fluoroquinolones may shorten treatment for drug-susceptible M. tuberculosis [10]. OfL is a human-made fluoroquinolone antibiotic. It prevents DNA replication by preventing bacterial DNA twisting gyrase [11].

Numerous studies have shown that adding OFL to the first rifampicin drug combination has a synergistic impact, improving therapy. This combination may work for drug-resistant and drug-susceptible isolates [12, 13]. A comprehensive literature review has documented several methods for measuring RIF [14–21] and OFX [22, 24] alone and in combination with other drugs. A high-performance liquid chromatographic (HPLC) method that concurrently measures RIF and OFX is unknown. This work aimed to create a selective, precise, and accurate reverse phase HPLC assay technique for parallel measurement of RIF and OFL in synthetic combinations. The suggested approach is validated under ICH Q2 (R1). To determine Ofloxacin and Rifampicin simultaneously using reverse phase high-performance liquid chromatography (HPLC), this study developed accurate and targeted methodologies.

Materials and Methods:

The drug substances used in this investigation include Rifampicin and Ofloxacin. The chemicals used for the analysis are Potassium dihydrogen orthophosphate (HPLC grade), Orthophosphoric acid (HPLC grade), Water (HPLC quality), and methanol (HPLC grade). The HPLC system used for the study is the Shimadzu-LC2010-CHT, manufactured in Kyoto, Japan.

Instrumentation:

An HPLC system, namely the Shimadzu-LC2010-CHT model from Kyoto, Japan, was used for the study. The system included an LC solutions data processing system, an SPD-M20A PDA detector, and an auto sampler. The data was collected using LC 2010 solutions software, specifically version 1.25. The investigation used an analytical balance (Shimadzu

AUW220 balance, Japan), a vacuum filtration assembly (TID 15, Mumbai, India), and an ultrasonic bath sonicator.

Selection of Wavelength:

The chromatographic conditions were optimized in order to establish a high-performance liquid chromatography (HPLC) approach for simultaneously determining RIF and OFL in both bulk and medicinal dose forms. To determine the appropriate wavelength, 10 μ g/ml standard solutions of RIF and OFX were analyzed using methanol as a reference in the spectrum mode, scanning between 200 and 400 nm. Both medicines exhibited significant absorption at a wavelength of 230 nm, which was chosen as the wavelength for detection (Figure 1).



Fig.No. 01: Zero order overlay spectra of RIF and OFL.

Preparation of mobile phase:

The mobile phase was prepared by mixing Potassium dihydrogen orthophosphate, acetonitrile, and methanol at a ratio of 65:35. Subsequently, the pH of the buffer was adjusted to 4.0, and the resultant solution was filtered using a 0.45μ membrane.

Preparation of standard stock solution:

10.0 mg of Rifampicin and 20.0 mg of Ofloxacin were quantified using a digital microbalance and then transferred into a 10 millilitre volumetric flask. After adding seven millilitres of diluent, the mixture underwent sonication to aid in dissolving. Afterwards, the solution was mixed with the diluent until it reached its maximum volume, and eventually, it was further diluted to get the necessary final volume by adding additional diluent.

Chromatographic conditions:

High Performance Liquid Chromatography equipped with PDA detector.

For Rifampicin and Ofloxacin (isocratic)

Column	:	Phenomenex C18 150 x 4.6 mm, 5m analytical column
Wavelength	:	230 nm
Injection Volume	:	20µ1
Column Temperature	:	Ambient

Flow rate : 1.0 ml/min

The RIF peak was seen at a retention time of 4.920 minutes. The displayed area measured 847,645 units, with a tailing factor of 1.23. Figure 2 and Table 1 indicate that the OFL peak was seen during a retention time of 11.710 minutes. The peak area was measured to be 6748754, with a tailing factor of 1.16 and a resolution of 5.35. This trial was deemed optimal due to its excellent outcomes and shorter retention period. The retention period of RIF is around 4.920 minutes, whereas the retention length of OFL is 11.710 minutes.



Fig.No. 02 : HPLC chromatogram for linearity of RIF and OFL at 230 nm Table 1: System suitability parameters

S.No.	Name of the Peak	Retention Time (Mins)	Peak Area	Tailing Factor	Resolution	Plate Count
01	Rifampicin	4.920	847645	1.23		6978
02	Ofloxacin	11.710	6748754	1.16	5.35	7547

Preparation of sample solution:

A total of 10.0 mg of Rifampicin and 20.0 mg of Ofloxacin were accurately measured and transferred into a 10 milliliter volumetric flask. Afterward, 7 milliliters of diluent were added. Afterwards, the combination underwent sonication to dissolve the material, and was subsequently diluted with a diluent until the necessary volume was achieved. The solution was further decreased to a volume of 10 ml by introducing the diluent and then filtered using a 0.45μ Nylon syringe filter.

Procedure:

Six injections, each with a volume of 20 μ l, were delivered using active standard solutions of RIF and OFL. Chromatograms were obtained and the peak responses were evaluated. The suitability of the system was evaluated by examining its parameters. The quantification of the quantities of RIF and OFL in the sample was achieved by examining the peak responses.

Method Validation:

The present analysis evaluated many criteria to verify the precision of the HPLC method in quantifying the amounts of RIF and OFL, in accordance with the authorized procedure. This demonstrates that the methodology is suitable for its designated objective. The validation criteria were implemented in compliance with the requirements set by the International Council for Harmonisation of Technical Requirements for Pharmaceuticals for Human Use (ICH).

Linearity and Range:

The concentrations of RIF and OFL that showed a direct link with peak area were between 2 and 10 μ g/ml for RIF and between 4 and 20 μ g/ml for OFL. The findings are shown in Figures 3 and 4, Tables 2 and 3, and the accuracy of the calibration curve is confirmed by the high correlation coefficient of the regression equation.

S.No.	Concentration (µg/ml)	Peak Area						
1	0	0						
2	2	424822						
3	4	847645						
4	6	1282467						
5	8	1695290						
6	10	2119455						
Slope		37621						
Intercept		2501.1						
Regression		0.9997						

 Table 2: Linearity data of RIF



Fig.No. 03 : Linearity of Rifampicin

 Table 3: Linearity data of OFL

S.No.	Concentration (µg/ml)	Peak Area
1	0	0
2	4	3378378
3	8	6748754
4	12	10124156
5	16	13797811
6	20	16971885
Slope	·	855902
Intercept		-66625
Regression	0.9996	



Fig.No. 04 : Linearity of Ofloxacin

Accuracy and Precision:

To evaluate the accuracy of recovery, we added extra Standard medication at three distinct concentration levels to a previously analyzed test solution. Our data indicate that the proposed technique is very accurate in predicting both RIF and OFL concurrently. The RSD was below 2.0%. In addition, we successfully obtained a recovery rate of 100.49% for RIF and 100.10% for OFL. The dependability of the Method is shown by its high level of repeatability and low values of relative standard deviation (RSD). The tables designated as 4 and 5.

	RIF				OFL			
Injection	Retention	Peak	Plate	Peak	Retention	Peak	Plate	Peak
Number	Time	Area	Count	Symmetry	Time	Area	Count	Symmetry
1	4.92	847711	6978	1.12	11.71	6748754	7567	1.22
2	4.91	847645	6999	1.23	11.65	6746734	7678	1.22
3	4.92	847723	6912	1.34	11.7	6749234	7689	1.34
4	4.9	847812	6923	1.25	11.45	6749467	7356	1.56
5	4.91	848657	6956	1.34	11.34	6749278	7547	1.1
6	4.92	849675	6979	1.45	11.34	6749678	7898	1.11
Average	4.913	1403270			11.532	6748858		
Standard	0 008	813 50			0 176	109/ 73		
Deviation	0.000	013.39			0.170	1004.75		
% RSD	0.1662	0.06			1.52	0.02		

Table 4: Precision data of RIF and OFL

Sample Preparation No.	RIF Assay (%)	OFL Assay (%)
	400.40	00.70
1	100.12	99.78
2	100.23	99.9
3	101.34	99.89
4	99.99	100.55
5	100.83	100.23
6	100.34	100.25
Mean	100.48	100.10
SD	0.5126	0.2927
RSD (%)	0.5102	0.2924

Table 5: Accuracy data of RIF and OFL

Robustness:

The outcomes of the robustness assessment are shown in Table 6. Both components had similar tailing factors, elution orders, resolutions, relative standard deviations, and recoveries. The research indicated that the relative standard deviation (RSD) of the peak locations was much less than 2.0%.

		Rifampio	cin	Ofloxacin				
Condition	%	Tailing	%	%	Tailing	%		
	RSD	Factor	Recovery	RSD	Factor	Recovery		
1) Change in Flow rate								
Normal Condition	0.18	1 1 1	100.01	0.11	1 21	100.21		
(1.0 ml per minute)	0.18	1.11	100.01	0.11	1.21	100.21		
Flow rate (0.8ml per	0.19	1 1 2	100.21	0.12	1 22	100.12		
minute)	0.17	1.12	100.21	0.12	1.22	100.12		
Flow rate (1.2 ml per	0.20	1 21	100.21	0.13	1 25	100.23		
minute)	0.20	1.21	100.21	0.15	1.25	100.25		
2) Change in minor compo	nent in t	he mobile	phase					
Normal Condition								
(Potassium dihydrogen								
orthophosphate,	0.18	1.23	100.21	0.18	1.31	100.23		
methanol in a ratio of								
65:35)								
(Potassium dihydrogen								
orthophosphate,	0.12	1 31	100.02	0.14	1 28	99 32		
methanol in a ratio of	0.12	1.51	100.02	0.14	1.28	99.32		
55:45)								
(Potassium dihydrogen	0.15	1 29	100.32	0.19	1.26	99 89		
orthophosphate,	0.15	1.27	100.52	0.19	1.20	77.07		

Table 6: Robustness data of RIF and OFL

methanol in a ratio of								
75:25)								
3) Change in Wave Length								
Normal:Wave Length	0.14	1 31	100.42	0.18	1.26	100 / 1		
230 nm	0.14	1.51	100.42	0.10	1.20	100.41		
Wave Length 235 nm	0.12	1.22	100.34	0.23	1.21	100.71		
Wave Length 225 nm	0.19	1.12	100.71	0.19	1.28	100.12		
4) Change in Ph								
Normal:pH 4.0	0.18	1.23	100.23	0.24	1.29	100.23		
pH 4.5	0.19	1.25	100.34	0.21	1.19	100.34		
рН 3.5	0.11	1.28	100.12	0.28	1.23	100.87		

Ruggudness:

The mean peak areas of Rifampicin and Ofloxacin were 845,682 and 6,748,934, respectively, with relative standard deviations (RSD) of 0.12% and 0.21%, respectively.

SUMMARY:

The study presents a newly designed and validated reverse-phase high-performance liquid chromatography (RP-HPLC) technique for precise quantification of RIF and OFL in both bulk and pharmaceutical samples. The literature analysis revealed a deficiency in approaches for precisely determining RIF and OFL in substantial quantities. Consequently, there is an urgent want for a direct, cost-effective, and precise solution to address this issue.

The concentrations of RIF and OFL were determined by injecting a solution consisting of Potassium dihydrogen orthophosphate and methanol (in a ratio of 65:35) with a pH of 4.0 onto a Phenomenex C18 column with dimensions of 150 x 4.6 mm and a particle size of 5m. The flow rate was set at 1.0 mL/min, while the injection volume was 20 μ L. The RIF peak had a retention duration of 4.920 minutes, whereas the OFL peak showed a retention period of 11.710 minutes.

After its enhancement, the technique underwent verification in accordance with ICH requirements to evaluate its compatibility with the system, linearity, sensitivity parameters, precision, accuracy, and durability. The findings of all validation parameters were within acceptable limits. The tests showed RSD values below 2, indicating a low level of variability. The recovery rate ranged from 99% to 101%.

CONCLUSION:

The recommended RP-HPLC technique offers a time-efficient and basic approach that is both simple and rapid. Additionally, it guarantees cost-efficiency. Therefore, it is a favored method for simultaneously measuring the amounts of Rifampicin and Ofloxacin. The implemented method was thoroughly verified to ensure adherence to ICH standards in every area.

REFERENCES:

- 1. World Health Organization. WHO global tuberculosis report. Geneva: WHO; 2016.
- 2. Hall RG, Leff RD, Gumbo T. Treatment of active pulmonary tuberculosis in adults: current standards and recent advances. Pharmacother. 2009;29:1468–1481.
- 3. O'Neil MJ, editor. The merck index an encyclopedia of chemicals, drugs, and biologicals. 13th ed. Whitehouse Station (NJ): Merck and Co., Inc.; 2001. p. 1474.
- 4. Maggi N, Pasqualucci CR, Ballota R, et al. Rifampicin: a new orally active rifamycin. Chemother. 1966;11: 285–292.
- 5. Rees RJW, Pearson JMH, Waters MFR. Experimental and clinical studies on rifampicin in treatment of leprosy. Br Med J. 1970;1:89–92.
- 6. Binda G, Domenichini E, Gottardi A. Rifampicin, a general review. Arzneim Forsch. 1971;21:1907–1977.
- 7. Pähkla R, Lambert J, Ansko P, et al. Comparative bioavailability of three different preparations of rifampicin. J Clin Pharm Ther. 1999;24:219–225.
- 8. Tsankov N, Angelova I. Rifampin in dermatology. Clin Dermatol. 2003;21:50–55.
- 9. Rainbow J, Cebelinski E, Bartkus J, et al. Rifampinresistant meningococcal disease. Emerg Infect Dis. 2005; 11:977–979.
- 10. Ginsburg AS, Grosset JH, Bishai WR. Fluoroquinolones, tuberculosis, and resistance. Lancet Infect Dis. 2003;3: 432–442. 154 P. SHAH ET AL.
- 11. Simpson KL, Markham A. Ofloxacin otic solution: a review of its use in the management of ear infections. Drugs. 1999;58:509–531.
- 12. Jurado ER, Tudó G, Bellacasac JP, et al. In vitro effect of three-drug combinations of antituberculous agents against multidrug-resistant mycobacterium tuberculosis isolates. Int J Antimicrob Agents. 2013;41:278–280.
- Jurado ER, Tudó G, Martinez JA, et al. Synergistic effect of two combinations of antituberculous drugs against mycobacterium tuberculosis. Tuberc. 2012;92: 260– 263.
- 14. Jain P, Pathak VM. Development and validation of UVvisible spectrophotometric method for estimation of rifapentine in bulk and dosage form. Der Pharma Chemica. 2013;5:251–255.
- 15. Tella ED, Sunitha S, Garikipati DK, et al. Assay of rifampicin in bulk and its dosage forms by visible spectrophotometry using chloranilic acid. IJCEE. 2012;3:64–67.
- 16. Sriram ST, Prasanthi B, Tata S, et al. Development and validation of high performance liquid chromatographic method for the determination of rifampicin in human plasma. Int J Pharm Pharm Sci. 2012;4:362–367.
- 17. Liua J, Suna J, Zhanga W, et al. HPLC determination of rifampicin and related compounds in pharmaceuticals using monolithic column. J Pharm Biomed Anal. 2008;46:405–409.
- 18. Ali J, Ali N, Sultana Y, et al. Development and validation of a stability- indicating HPTLC method for analysis of antitubercular drugs. Acta Chromatographica. 2007;18:168.

- 19. Kapuriya KG, Parmar PM, Topiya HR, et al. Method development and validation of rifampicine and piperine in their combined dosage form. Int Bull Drug Res. 2012;1:71–80.
- 20. Bhusari SS, Bhat V, Koul M, et al. Development and validation of a RP-HPLC method for the simultaneous determination of rifampicin and a flavonoid glycoside a novel bioavailability enhancer of rifampicin. Trop J Pharm Res. 2009;8:531–537.
- 21. Yan H, Zhou Y, Xie Q, et al. Simultaneous analysis of isoniazid and rifampicin by high performance liquid chromatography with gradient elution and wall-jet/thin-layer electrochemical detection. Anal Methods. 2014;6:1530–1537.
- 22. Wankhede SB, Prakash A, Chitlange SS. Simultaneous reverse phase Hplc estimation of ofloxacin and satranidazole in tablet dosage form. Int J PharmTech Res. 2009;1:1136–1138.
- Sireesha KR, Prakash K. HPLC-UV method for simultaneous determination of ofloxacin and dexamethasone sodium phosphate. Int J Pharm Pharm Sci. 2012;4: 415–418.
- 24. Deekonda P, Reddy MS. Method development and validation for the quantitative estimation of cefixime and ofloxacin in pharmaceutical preparation by RP- HPLC. Der Pharma Chemica. 2014;6:31–37.