



## ***In Vitro* Evaluation of the Antidiabetic and Antimitotic Potential of *Dracaena trifasciata* Leaves Extract**

**Dhawal Dorwal<sup>1</sup>, Sachin Sisodiya<sup>2</sup>, Sonam Chaturvedi<sup>3</sup>, Henika Ghadge<sup>2</sup>, Juliyas Shrotriya<sup>4\*</sup>, Pradeep Tomar<sup>5</sup>, Ishan Dubey<sup>4</sup>**

<sup>1</sup> University Institute of Pharmacy, Oriental University, Indore, M.P., India

<sup>2</sup> Oxford International College, Indore, M.P., India

<sup>3</sup> Sri Aurobindo Institute of Pharmacy, Indore, M.P., India

<sup>4</sup> Laksmi Narain College of Pharmacy, Indore, M.P., India

<sup>5</sup> Chameli Devi Institute of Pharmacy, Indore, M.P., India

**\*Author for correspondence:** Mr. Juliyas Shrotriya, Asso. Prof., LNCP, Indore, M.P., India

**Mail Id:** [juliyas.shrotriya@gmail.com](mailto:juliyas.shrotriya@gmail.com)

### Article History

Volume 6, Issue 10, Feb 2024

Received: 17 Feb 2024

Accepted: 01 Apr 2024

doi: 10.33472/AFJBS.6.10.2024.387-392

### **Background**

The present study aimed to investigate the *in vitro* anti-diabetic and antimitotic activity of leaves extract of *Dracaena trifasciata*. As it was reported that *Dracaena trifasciata* contain flavonoids and other phytoconstituents, an attempt was made to determine its anti-diabetic and antimitotic potential.

### **Methods**

The *in-vitro*  $\alpha$ -amylase inhibition and antimitotic activity of the leaves extract of *Dracaena trifasciata* were evaluated by using 3,5-dinitrosalicylic acid (DNSA) and *Allium cepa* root meristematic cells.

### **Results**

The result of  $\alpha$ -amylase enzyme inhibition activity was found in a dose-dependent way, the strongest activity was shown by 25mg/ml hydroalcoholic extract 32.48 $\pm$ 0.76%. The antimitotic activity was screened by using *Allium cepa* root meristematic cells. Methotrexate (0.1 mg/mL) was used as a standard. It is found that all concentration of extracts showed antimitotic activity, but 100mg/ml possess maximum antimitotic potential. Extract shows 61.88% mitotic index at 100mg/ml concentration.

**Keywords:** *Dracaena trifasciata*, Flavonoids, polyphenols, Antidiabetic, Antimitotic

**Introduction:**

Diabetes mellitus presents as a severe and enduring collection of metabolic irregularities, marked by continual high levels of glucose in the blood. These anomalies stem from disruptions in carbohydrate, fat, and protein processing, which can result from both genetic predispositions and external factors. These disruptions may manifest as either insufficient insulin production or inadequate response to the insulin generated (Okur ME *et al.*, 2017, Wadkar K *et al.*, 2008). The antidiabetic activity of medicinal plants is due to the presence phenolic compounds Flavonoids, Terpenoids, Alkaloids, Glycosides, Steroid, Peptides, Lipids and other constituents (Demissew S *et al.*, 2011, Grover J *et al.*, 2002, Vuksan V *et al.*, 2005). Flavonoids interfere with cyclin-dependent cell cycle regulation and interact with drug transport (Halliwell B. 2007). Ahmed *et al.* (2015) stated that flavonoids inhibit the proliferation of cell lines and demonstrated strong cytotoxicity towards colon cancer cells.

*Dracaena trifasciata* is a species of flowering plant in the family Asparagaceae, native to tropical West Africa and Asia. It is most commonly known as the snake plant (Takawira-Nyenya R. 2021). Thu ZM *et al.* (2020) reported presence of homoisoflavonoids, named trifasciatine A and (-)-(3R)-trifasciatine B from extract of *D. trifasciata*. The present study aimed to investigate the *in-vitro* anti-diabetic activity and anti-mitotic activity of leaves extract of *Dracaena trifasciata*.

**Materials and method:**

*Dracaena trifasciata* leaves was collected from Ashtang Ayurvedic College, Indore. The botanical identification and authentication of the plant materials were performed by Mrs. S. Mishra (botanist) and the voucher specimen was deposited in Herbarium of Biology Department, Govt. SN College, Knw (M.P.). The leaves were air dried at room temperature and milled with the aid of grinding machine.

**Preparation of leaves extract:**

Plant materials (100 g) were soaked in 450 mL of 50% hydroalcoholic solvent (Ethanol: water). This solvent system placed for 48 h at room temperature. After 48 h, solvents were filtered through Whatman filter paper no.1 separately in a beaker. The filtrate was dried in a rotary evaporator at 55 °C to obtain the concentrated yield of extracts (De Mesquita *et al.* 2007).

**Determination of total phenolic compounds and flavonoids content:**

The Folin-Ciocalteu method was employed to ascertain the total phenolic compound contents. In this process, 0.5 ml of various dilutions of the extract samples were combined with 2.5 ml of 0.2 N Folin-Ciocalteu reagent (obtained from Sigma–Aldrich) for a duration of 5 minutes. Subsequently, 2.0 ml of 75 g/l sodium carbonate solution was added. After an incubation period of 2 hours at room temperature, the absorbance of the reaction was measured at 760 nm. The outcomes were quantified in terms of gallic acid equivalents. Additionally, total flavonoids were assessed utilizing the method developed by Ordonez *et al.*, 2006. To 0.5 ml of the sample, 0.5 ml of 2% AlCl<sub>3</sub> ethanol solution was introduced.

Following a 1-hour incubation at room temperature, the absorbance was recorded at 420 nm. Total flavonoid contents were determined by referencing a calibration curve constructed with quercetin

### **$\alpha$ -Amylase inhibition activity**

The  $\alpha$ -amylase inhibition assay was conducted following the 3,5-dinitrosalicylic acid (DNSA) method as outlined by Wickramaratne *et al.* (2016). The hydroalcoholic extract, ranging in concentrations from 5, 10, 15, 20 and 25mg/mL, derived from *Dracaena trifasciata* leaves, was dissolved in a buffer solution (0.02 M Na<sub>2</sub>HPO<sub>4</sub>/NaH<sub>2</sub>PO<sub>4</sub>, 0.006 M NaCl) adjusted to pH 6.9. To initiate the assay, 200  $\mu$ L of  $\alpha$ -amylase solution (2 units/mL) was combined with 200  $\mu$ L of the extract and incubated for 10 minutes at 30°C. Subsequently, 200  $\mu$ L of starch solution (1% in water, w/v) was added to each tube and incubated for 3 minutes. The reaction was halted by adding 200  $\mu$ L of DNSA reagent (comprising 12 g of sodium potassium tartrate tetrahydrate, 8.0 mL of 2 M NaOH, and 20 mL of 96 mM 3,5-dinitrosalicylic acid solution) followed by boiling for 10 minutes in a water bath set at 85°C. After cooling to room temperature, the mixture was diluted with 5 mL of distilled water, and the absorbance was measured at 540 nm using a UV-Visible spectrophotometer. A blank with 100% enzyme activity was prepared by substituting the plant extract with 200  $\mu$ L of the buffer. Additionally, a control reaction was set up using acarbose, following the same procedure as with the plant extract. The inhibition of  $\alpha$ -amylase was expressed as percentage of inhibition and was calculated by the following equation:

$$\text{Inhibition (\%)} = \frac{(A_c - A_{cb}) - (A_s - A_{sb})}{(A_c - A_{cb})} \times 100,$$

Where,

$A_c$ =absorbance of control;  $A_{cb}$ =absorbance of control blank;  $A_s$ =absorbance of sample;

$A_{sb}$ =absorbance of sample blank. The %  $\alpha$ -amylase inhibition was plotted against the extract concentration and the IC<sub>50</sub> values were obtained from the graph.

### **Antimitotic assay**

A modified technique outlined by Fiskesjo (1985) was employed to assess the antimitotic activity using *Allium cepa* roots. *Allium cepa* bulbs weighing approximately 40  $\pm$  10 grams were allowed to germinate in water for 72 hours at room temperature under dark conditions. Bulbs exhibiting roots grown to approximately 3 cm were chosen for subsequent experimentation. These onion roots were placed in beakers filled with water, methotrexate (0.1 mg/mL), and various concentrations of plant extracts 10, 20, 30, 40, 50 and 100mg/ml for duration of 24 hours. Water was utilized for dilution purposes, serving as the control, while methotrexate was employed as the standard for the study. Following the 24-hour exposure period, the number of dividing and non-dividing cells was recorded, and the mitotic index was calculated using the following formula:

$$\text{Mitotic Index} = \frac{\text{No. of dividing cell}}{\text{Total no. of cells}} \times 100$$

## Results and discussion

### Total phenol and flavonoids contents:

The concentration of total phenol compounds in the leaves was determined using the Folin-Ciocalteu method and expressed in terms of gallic acid equivalents, utilizing a standard curve equation ( $y = 0.0064x$ ) with a correlation coefficient ( $r^2$ ) of 0.989. The total phenolic content was measured to be  $138.3 \pm 5.5$  mg gallic acid equivalent per gram of extract powder. Similarly, the total flavonoid content in the leaves was quantified as  $29.1 \pm 0.8$  mg quercetin equivalent per gram of extract powder, employing a standard curve equation ( $y = 0.0066x + 0.0131$ ) with an  $r^2$  value of 0.996.

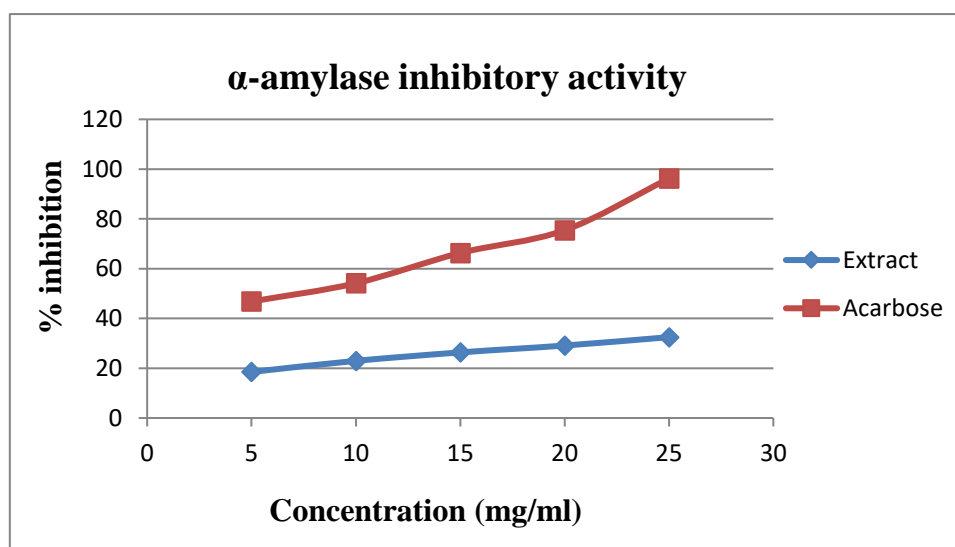
### *In-vitro* $\alpha$ -Amylase Inhibition Activity

In this study, leaves extract were evaluated for their possible  $\alpha$ -amylase inhibitory activities alongside acarbose as a positive control. The  $\alpha$ -amylase inhibitory activities and IC<sub>50</sub> values of the acarbose and leaves extract are summarized in Table 1. Results showed the inhibitory activities of extracts and standard, IC<sub>50</sub> values were found to be 50.42 and 7.5 for leaves extract and acarbose respectively. Concentration dependent inhibition was observed for various concentrations of the tested extracts and the standard. Among the extracts, the hydroalcoholic extract of 25mg/mL concentration showed the highest alpha-amylase enzyme inhibition activity of  $32.48 \pm 0.76\%$ . The standard positive control Acarbose showed an IC<sub>50</sub> value of  $96.22 \pm 0.78\%$  inhibition at 25mg/mL)

**Table 1:**  $\alpha$ -Amylase inhibitory activities of the extracts and standard

Concentration	Percentage inhibition	
	Extract	Acarbose
5mg/ml	18.52 $\pm$ 0.84	46.82 $\pm$ 0.68
10mg/ml	22.96 $\pm$ 0.78	54.16 $\pm$ 0.64
15mg/ml	26.36 $\pm$ 0.82	66.32 $\pm$ 0.58
20mg/ml	29.12 $\pm$ 0.88	75.45 $\pm$ 0.66
25mg/ml	32.48 $\pm$ 0.76	96.22 $\pm$ 0.78

Values are expressed as Mean  $\pm$  SD for triplicates



**Figure 1:**  $\alpha$ -Amylase inhibitory activities of the extracts and standard

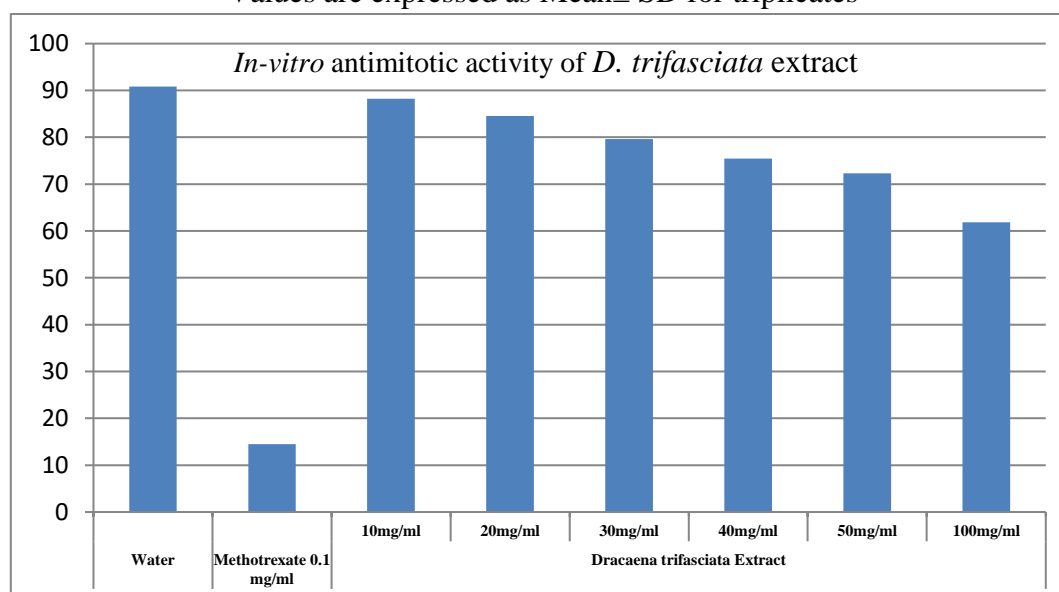
**Antimitotic assay:**

The results of antimitotic assay effect of *Dracaena trifasciata* hydroalcoholic extract on mitotic index (MI) of *Allium cepa* root tip cells are given in Table and Figure 2. Thus, all plants extracts displayed significant antimitotic activity which indicates its use as a potent antimitotic agent.

**Table 2** Effect of *D. trifasciata* extract on mean root length mitotic index (MI) of *Allium cepa* root tip cells

Treatments		Mean Root length (in mm)	Mitotic Index (MI) %
Water		34.12±1.26	90.85
Methotrexate 0.1 mg/ml		16.44±1.28	14.52
Dracaena trifasciata Extract	10mg/ml	29.22±1.32	88.22
	20mg/ml	28.56±1.34	84.56
	30mg/ml	26.08±1.3\	79.64
	40mg/ml	22.32±1.48	75.45
	50mg/ml	20.18±1.44	72.34
	100mg/ml	19.86±1.32	61.88

Values are expressed as Mean± SD for triplicates



**Figure 2:** Effect of *D. trifasciata* extract on mitotic index (MI) of *Allium cepa* root tip cells

**Conclusion**

According to the result of the study on the leaf extracts of *D. trifasciata* exhibit  $\alpha$ -amylase inhibitory activity with remarkable activity in the hydroalcoholic extract. Results for antimitotic activity for extract were comparable to the activity of methotrexate. Maximum numbers of non-dividing cells were observed. As a result of this cells arrest in mitosis and eventually die by apoptosis. Furthermore, this study has opened opportunities for future

research in searching for novel effective drugs for diabetics that possess anti-diabetic activity and for antimetabolic or anticancer agent.

### References

1. Ahmad A, Kaleem M, Ahmed Z, Shafiq H. Therapeutic potential of flavonoids and their mechanism of action against microbial and viral infections—A review. *Food Research International*. 2015 Nov 1;77:221-35.
2. De Mesquita ML, Grellier P, Mambu L, De Paula JE, Espindola LS. In vitro antiplasmodial activity of Brazilian Cerrado plants used as traditional remedies. *Journal of Ethnopharmacology*. 2007 Mar 1;110(1):165-70.
3. Demissew S, Friis I, Awas T, et al. Four new species of Aloe (Aloaceae) from Ethiopia, with notes on the ethics of describing new taxa from foreign countries. *Kew Bull*. 2011;66(1):111–121.
4. Fiskesjö G. The Allium test as a standard in environmental monitoring. *Hereditas*. 1985 Mar;102(1):99-112.
5. Grover J, Yadav S, Vats V. Medicinal plants of India with anti-diabetic potential. *J Ethnopharmacol*. 2002;81(1):81–100.
6. Halliwell B. Flavonoids: a Re-Run of the Carotenoids Story?. In *Dietary Supplements and Health: Novartis Foundation Symposium 282* 2007 Jul 27 (Vol. 282, pp. 93-104). Chichester, UK: John Wiley & Sons, Ltd.
7. Okur ME, Karantas ID, Siafaka PI. Diabetes mellitus: A review on pathophysiology, current status of oral medications and future perspectives. 2017; 55(1):61.
8. Ordonez AA, Gomez JD, Vattuone MA. Antioxidant activities of *Sechium edule* (Jacq.) Swartz extracts. *Food chemistry*. 2006 Aug 1;97(3):452-8.
9. Takawira-Nyenya R, Thiede J, Mucina L. New nomenclatural and taxonomic adjustments in *Dracaena* (Asparagaceae). *Phytotaxa*. 2021 Nov 9;524(4):293-300.
10. Thu ZM, Myo KK, Aung HT, Armijos C, Vidari G. Flavonoids and stilbenoids of the genera *Dracaena* and *Sansevieria*: Structures and bioactivities. *Molecules*. 2020 Jun 3;25(11):2608.
11. Vuksan V, Sievenpiper JL. Herbal remedies in the management of diabetes: lessons learned from the study of ginseng. *Nutr Metab Cardiovasc Dis*. 2005;15(3):149–160.
12. Wadkar K, Magdum C, Patil S, Naikwade N. Anti-diabetic potential and Indian medicinal plants. *J Herb Med Toxicol*. 2008;2(1):45–50.
13. Wickramaratne MN, Punchihewa JC, Wickramaratne DB. In-vitro alpha amylase inhibitory activity of the leaf extracts of *Adenanthera pavonina*. *BMC complementary and alternative medicine*. 2016 Dec;16:1-5.