



## In vitro Anti-biofilm Activity of Probiotic-mediated Biosynthesized Silver Nanoparticles

Hajar T. Mahdi, Zainab N. Al-Saadi

Department of Microbiology, College of Science, Wasit University, Iraq

Corresponding author (\*): Hajar T. Mahdi

### Article Info

Volume 6, Issue 8, April 2024

Received: 07 Feb 2024

Accepted: 03 March 2024

Published: 06 April 2024

### Abstract

**Background:** Seventy samples were collected from the feces of healthy newborn babies aged (2-4) weeks and from the arrivals to vaccination centers feed by breast milk in Wasit province, Iraq. Babies were assessed clinically, placed directly in the sterile liquid MRS medium in test tubes and transferred to the laboratory to complete the isolation processes. **Methods:** Diagnosis of lactic acid bacteria isolates included: catalase test, gelatin liquefaction test and carbohydrate fermentation test for diagnosis of anaerobic bacteria in order to contain the number of confirmatory biochemical tests. Characteristics of isolates were compared with what exists in the antagonism activity assay against *Klebsiella pneumoniae*, determination of minimum inhibitory concentration, determination of minimum bacteriocidal concentration and detection of biofilm production using microtiter plate method was done. **Results:** By tissue microtiter plate method indicated that 1(3.3%) were strong for biofilms formation, while 6(20%) were moderate and 23 (76.7%) were demonstrated as a weak biofilm formation, (0%) were reported as nonbiofilm producing isolates. Microtiter plate method Antibiofilm effect of silver nanoparticles on 30 isolates of *K.pneumoniae* showed that the biofilm inhibition was tested, in which mean of control (biofilm formation without silver nanoparticles) was 0.21 compared to 0.07 using Bn1, while it was 0.21 compared to 0.08 as a results for antibiofilm using sub-MIC, (p value= 0.001, 0.004) respectively. **Conclusion:** Antibiofilm effect of silver nanoparticles on *K.pneumoniae* isolates of *Bifidobacterium* isolates showed that the biofilm inhibition has a significant effect.

**Key words:** Silver Nanoparticles, Biofilm and Probiotic

© 2024 Hajar T. Mahdi, This is an open access article under the CC BY license (<https://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made

### Introduction

Today, the term “probiotic” refers to “live microorganisms which when administered in adequate amounts, confer a beneficial physiological effect on the host,” according to the Food and Agriculture Organization and World Health Organization (1, 2). The normal flora of the human gastrointestinal tract contains many diverse populations of bacteria which play an essential role in the development and well-being of the host. In the intestinal microflora exerts a protective role against pathogens (3, 4). Most commonly used probiotic supplements contain the species of *Lactobacillus* and *Bifidobacterium* and they are the part of normal human intestinal microbiota

(5, 6). The antagonism of lactic acid bacteria is exerted by competition for nutrients and for physical location, but also through the production of antimicrobial substances (7). These compounds are able to inhibit the growth of harmful microorganisms and the most important LAB are lactobacillus spp and Bifidobacterium spp (1, 8).

As a global public health issue, antimicrobial resistance (AMR) is a growing problem that occurs when microorganisms such as bacteria, viruses, fungi, and parasites become resistant to antimicrobial drugs that have previously served as effective treatments for infections (9). When bacteria become resistant to antibiotics, they can spread infections which are difficult to treat, which can result in prolonged illness, disability and even death as a result (10). Multi-drug resistance (MDR) is a form of AMR in which microorganisms become resistant to multiple drugs, making it more difficult to treat infections (11). This can happen when antibiotics are overused or misused, as well as when there is poor infection prevention and control in healthcare settings (12).

Recently, a lot of attention has been paid in finding ways to produce and use the nanomaterials, and the interest is growing every day. In terms of manufacturing nanoparticles, one of the methods to be considered is the bio-approach (13). Using microorganisms for synthesis of nanoparticles, for example, is referred to as nanoparticles synthesis by biological means. Nanoparticles are produced both by living and dead microorganisms, contribute greatly to nanoparticle production (14). An array of microorganisms such as *k. pneumoniae*, *E. coli*, *S. aureus*, *P. aeruginosa* and *S. typhus* may be susceptible to AgNPs as antimicrobial agents (15). Hence, as an emerging method for discovering antibacterial involves using green synthesized nanoparticles to target bacterial biofilm and QS. Utilization of silver nanoparticles as an alternative antimicrobial agent has been suggested (16). Nano-scaled materials appear to exhibit better biological effects than their bulk counterparts because their chemical and physical properties are different at this scale, which is mainly why the nano-scaled materials shown improved biological activity (17).

Probiotic bacterium that has gained considerable attention in the field of medicine due to its potential health benefits. As a naturally occurring bacterium in the human gastrointestinal tract (18). The synthesized AgNPs-LR were investigated for their broad-spectrum effect on inhibiting the virulence factors of resistance bacterial species (19, 20).

## **Materials and Methods**

### **Samples Collection**

Seventy samples were collected from the feces of healthy newborn babies at the age of (2-4) weeks, samples were collected from Wasit province, from 2022 to February 2023. The patients were first assessed clinically by the doctors in the hospital and then referred for sample collection (4).

### **Diagnosis of Lactic Acid Bacteria Isolates**

The isolates underwent biochemical tests that included: Catalase test, Gelatin liquefaction test and Carbohydrate fermentation test for diagnosis of anaerobic bacteria in order to contain the number of confirmatory biochemical tests. Characteristics of isolates were compared with what exists in the (1).

### ***Klebsiella pneumoniae* isolates**

Thirty isolates of *K. pneumoniae* were obtained from (21).

### **Antagonism Activity Assay**

#### **Cell-Free Extract (CFE) Preparation**

Cell-free extract of all lactic acid bacteria used in this study was prepared according to (22) as follows: Bifidobacterium spp. include (6) strains which were inoculated separately broth as 2 % of broth volume and incubated under anaerobic condition at 37°C for 72 h. The culture was then centrifuged at 5000 rpm for 30 min. The Supernatants were sterilized by filtration through (0.22µm) membranes (Millipore filter paper-Swinnex-25).

### **Determination of Minimum Inhibitory Concentration**

The MIC of CFEs for the test strains was determined according to (23). One isolate of *Bifidobacterium* fresh culture was inoculated in 10 ml nutrient broth containing filter concentrates of the culture supernatant (50,100,150,200)  $\mu$ l and incubated aerobically at 37°C for 24 h for MIC determination (6).

### **Determination of Minimum Bacteriocidal Concentration**

After the serial dilution for every treatment was done, then the bacteriocidal activity of CFE (*Bifidobacterium*) was determined by plating (0.1 ml) for each treatment into Mueller Hinton sterilized petri dishes and then incubated at 37°C for 24 h, after incubation, results were recorded and compared with the control treatment (0% of CFE) (1, 6).

### **Detection of Biofilm production**

#### **Microtiter plate method**

According to (21), biofilm formation test was detected by microtiter plate method.

### **Synthesis silver nanoparticles by biological method**

#### **Preparation of stock solution of AgNO<sub>3</sub>**

Preparation of stock solution of AgNO<sub>3</sub> by dissolving 0.085g of solid AgNO<sub>3</sub> with 10ml of distilled water and stirred for 30 min by a magnetic stirrer and kept until used for preparation concentration 10mM (24).

#### **Preparation of bacterial supernatant**

To produce the biomass for biosynthesis, *Bifidobacterium* is culture in nutrient broth or LB medium incubated rotary shaker (200 rpm) for 37°C at 24 hr then the supernatant is collected by centrifugation at 10,000 rpm for 10 min. The supernatant was used for synthesis of AgNPs (25).

#### **Synthesis of AgNPs**

Ten milliliters (10ml) of supernatant is mixed with 10ml of AgNO<sub>3</sub> solution to form 10mM concentration and other tube without silver nitrate serve as control supernatant without silver nitrate, pH of reaction was 9.0 and the prepared solution were incubated at 37°C for 48 hr. All solutions are kept in dark to avoid any reaction during testing after incubation the solution changed from yellow to brown solution the AgNPs are collected by centrifugation at 10,000rpm, for 5 min twice (24).

### **Characterization nanoparticles**

Identification of silver nanoparticle (AgNPs) in Nano-center (Technology University) by using:

#### **Scanning Electron Microscopy (SEM)**

A scanning electron microscope has resolution 3nm at 30kV take AgNPs images. The assembly involved with a computer software programming to analyze the mean size of the particles in sample (1).

#### **X-Ray Diffraction (XRD)**

According to Ramalingam *et al.*, (2014), "X-ray diffraction, 40 KV voltage and with current (20Ma), is used to identify the crystalline phases and to evaluation the crystalline size and the XRD patterns recorded with 2s in the range of (10 $\theta$  - 60 $\theta$ ) by step scanning, employing Cu tube with wavelength of Cu 1.54 Å".

#### **Fourier Transform Infra-red Spectroscopy (FTIR)**

FTIR analysis of the AgNPs carried out on FTIR 8400S, (SHIMADZU-FTIR) spectrophotometer in the range 400–4000 cm<sup>-1</sup> (26).

### **Effect of silver nanoparticles on biofilm production**

#### **Microtiter plate method**

According to (27), effect of silver nanoparticles on biofilm formation by microtiter plate method.

The inhibition rate (%) was read as following

$$\text{Inhibition rate (\%)} = \frac{\text{OD of control} - \text{OD of treated}}{\text{OD of control}} \times 100$$

The microtiter plate antibiofilm assay estimations the percentage of bacterial biofilm reduction. In comparative to the control wells, which were fixed at 100% to indicate the absence of silver nanoparticles. In compare, negative percentage results indicate no inhibition activity of AgNPs on biofilm association.

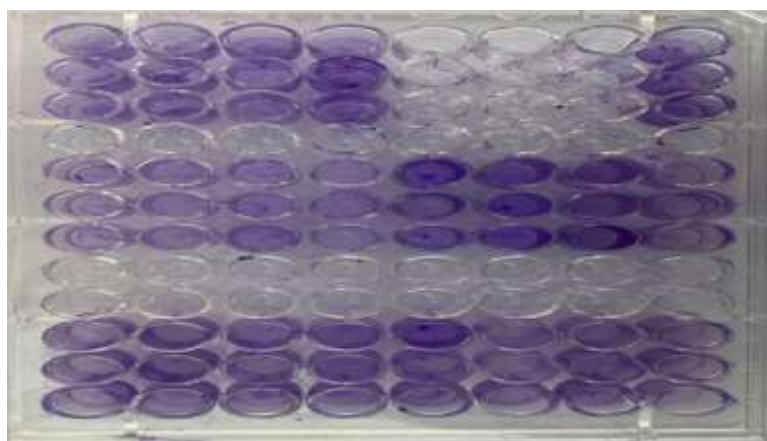
## Results and Discussion

### Biofilm Production Assay

The results of biofilm formation to *Klebsiella pneumoniae* isolate (30 isolates) obtained from (21). by tissue microtiter plate method indicated that 1(3.3%) were strong for biofilms formation, while 6(20%) were moderate and 23(76.7%) were demonstrated as a weak biofilm formation, (0%) were reported as nonbiofilm producing isolates, table (1) and figure (1).

**Table (1) Biofilm formation result of *Klebsiella pneumoniae* using microtiter plate method**

Biofilm formation	OD630 limits number of isolates	Number of isolates	percentage
Non-adherent	<0.06-0.12	0	0%
weak	0.12-0.24	23	76.7%
moderate	0.132-0.264	6	20%
strong	$\geq 0.24$	1	3.3%



**Figure (1). Biofilm production in *Klebsiella pneumoniae***

### Biosynthesis of silver nanoparticles

#### Using supernatant of *Bifidobacterium* in Synthesis of Silver nanoparticle

Nanoparticle synthesis in the used medium has been noticed by color change of *Bifidobacterium* supernatant with an aqueous solutions of silver nitrate (concentration of 10 mM) from yellow to brown after forty-eight hours after incubation.

### Characterization of synthesized silver nanoparticles

#### Scanning Electron Microscopy (SEM)

Figure (2) illustrates the morphological characteristics of produced silver nanoparticles examined by SEM apparatus. Ag-NPs had reduced aggregation, and 35 nm was the average size (28).

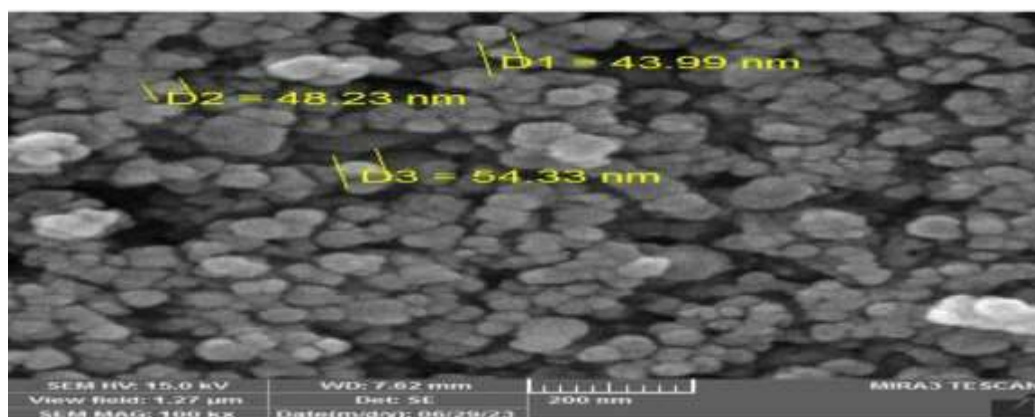


Figure (2): SEM image of Ag-NPs synthesized by *Bifidobacterium*

**X-ray Diffraction (XRD)**

Crystal structure of AgNPs was examined by XRD the, figure (3) exhibited one high peak is visible in the XRD spectrum of produced silver nanoparticles at  $2\theta$  (32.5°) at which corresponds to the (101) plane of conventional XRD data of nano silver Crystals (29).

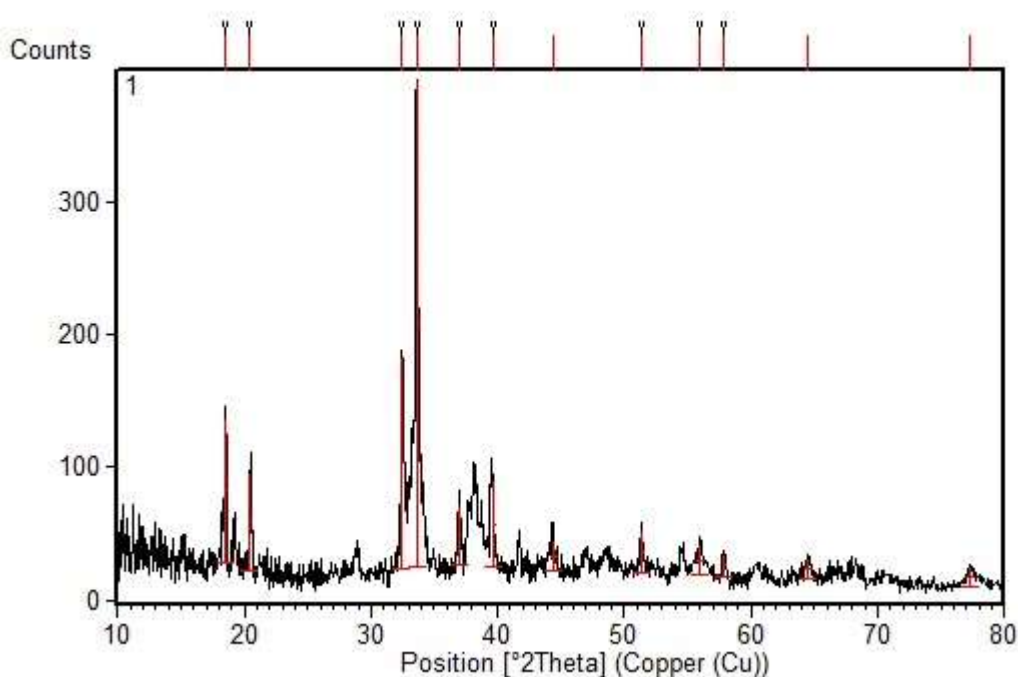


Figure (3): XRD range of synthesize Ag-NPs

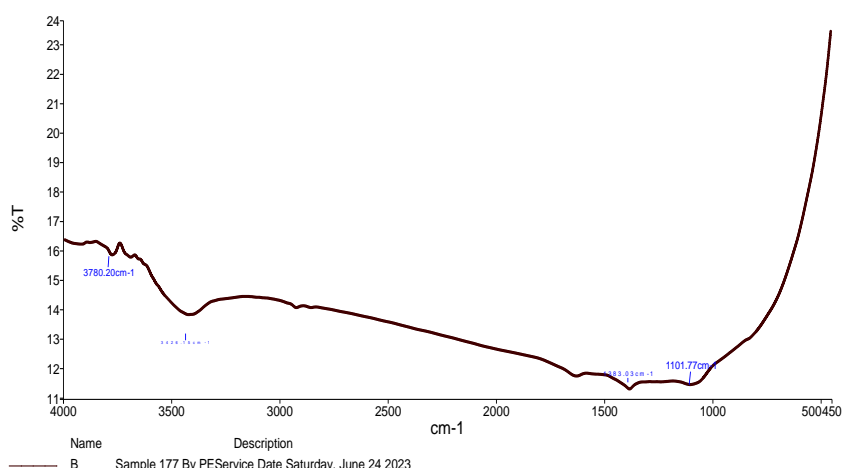
**Fourier Transform-Infrared spectroscopy (FT-IR)**

FT-IR analysis of AgNPs synthesized by *Bifidobacterium* noted in the range (450 -4000) cm-1 as shown in table (2) and figure (4) ,it is useful for determining the chemical composition of reactants involved in the synthesis and coating of AgNPs.

Table (2) FT-IR analysis of AgNPs synthesized

Spectrum Name	Peak Number	X (cm-1)	Y (%T)
B	1	3780.20	15.86
	2	3426.15	13.81
	3	1383.03	11.27

	4	1101.77	11.42
--	---	---------	-------



**Figure (4) : FTIR analysis of silver nanoparticles**

**Determination MIC and Sub-MIC of Silver nanoparticles**

In this study, Ag-NPs were tested against 30 isolates of *K.pneumoniae* by using a Resazurin-mediated microtiter plate and well diffusion method, the color change of the Resazurin indicator was used to visually reflect the inhibitory action. The results have shown that silver nanoparticles has inhibitory action (MIC) against *K.pneumoniae* with concentration 10Mm and sub-MIC with concentration 5mM . Determination of MIC is important for bacteria to determine the lowest concentration of AgNPs that is necessary for inhibition of visible bacterial growth after incubation period at 37°C for18-24 hr (30).

**Testing the effect of silver nanoparticles on biofilm formation**

**Microtiter plate method**

Antibiofilm effect of silver nanoparticles on 30 isolates of *K.pneumoniae* showed that the biofilm inhibition was tested, outcomes were illustrated in table (3) and (4), in which mean of control (biofilm formation without silver nanoparticles) was 0.21 compared to 0.07 using Bn1, while it was 0.21 compared to 0.08 as a results for antibiofilm using sub-MIC, (p value= 0.001, 0.004) respectively.

Table (3) and (4) Determination of biofilm and anti-biofilm in *Klebsiella pneumoniae* using silver nanoparticle

Sub-MIC	Biofilm in <i>K. pneumoniae</i>	Anti-biofilm using sub-MIC		Mean
		Bn1		
16	0.35	0.18		0.26
64	0.21	0.05		0.13
8	0.22	0.11		0.11
16	0.21	0.09		0.15
4	0.20	0.03		0.11
16	0.22	0.09		0.15
64	0.10	0.01		0.05
64	0.21	0.02		0.11
Mean	0.21	0.07		0.13
P value	<b>0.001</b>			

Sub-MIC	Biofilm in <i>K. pneumoniae</i>	Anti-biofilm using sub-MIC	Mean
---------	---------------------------------	----------------------------	------

		Bn2	
64	0.35	0.20	0.27
128	0.21	0.10	0.15
32	0.22	0.18	0.20
32	0.21	0.15	0.18
16	0.20	0.05	0.12
64	0.22	0.10	0.16
128	0.10	0.04	0.07
128	0.21	0.06	0.13
Mean	0.21	0.08	0.16
P value	<b>0.004</b>		

## Discussion

In a previous study (31) it has been proposed the using of medium contain MRS added to l-vancomycin, bromocresol purple and cysteine, this will provide optimum condition in isolation and counting probiotic. It was explored the probability to improving "LcS" select medium by (31) used all MRS component.

In biofilm formation, the structure and regulatory genes play important role that effect on colony aggregation by different mechanisms ,as well as alteration of synthesis of transcriptional factors and regulation of extracellular polysaccharide production (32). In agreement with some local studies, (33) indicated that 100% of clinical *K. pneumoniae* isolates were able to produce biofilm. Also, (34, 35), observed that of 30 *K. pneumoniae* isolates; 23 (46%) were weak-biofilm producers.

Findings are in line with those of several earlier research like that by (36) study the results show that the MTP method was more sensitive in detecting biofilm production compared to the CRA. (37) (38) and (39) studies showed that there was no relationship between the two methods, and the MTP method is the most sensitive and realistic.

Strong bands at (1101.77, 1383.03, 3426.15 and 3780.20  $\text{cm}^{-1}$ ) were discovered by FT-IR analysis of Ag-NPs. The stretch for Ag-NPs discovered around 1101.77  $\text{cm}^{-1}$ , and the other band at (3780.20) corresponds to -OH- free, (1383.03) shows the H-C-H Asymmetric, while band at (3426.15) for O-H stretching corresponds to carboxylic acid, corresponds OH-bend resembling to phenolic compounds which can potentially affect the synthesis and stability of AgNPs, these results are in agreement with previous reports (40, 41). (42) illustrated how silver weakens biofilm formation, when AgNPs transported into the cell and interact with proteins and enzymes, which are essential for microbial development quorum sensing or adherences that resulted in the decline in biofilm action. AgNPs inhibited biofilm formation by blocking the formation of exopolysaccharides also the silver nanoparticles move during the water channels used for nutrient moving and spread through exopolysaccharides layer (43).

Results of (34) study has revealed that mutation occurred in the gene after treatment with AgNPs, which may lead to the effect of the phenotype because they lead to change in amino acids and then protein that transition mutation was repeated 2 time at adnine and guanine bases that change amino acid.

In current study antibiofilm effect of silver nanoparticles on 30 isolates of *K.pneumoniae* showed that the biofilm mean of control (biofilm formation without silver nanoparticles) was 0.07 compared to 0.008 using Bn1 as inhibitor, while it was 0.93 compared to 0.30 as a results for antibiofilm using sub-MIC, (p value= 0.004 and 0.001) in microtiter plate method. These

outcomes were agreed with (44). Probiotics are the live micro-organisms having host beneficial effects by enhancing microbial balance in intestine, whereas, prebiotics are indigestible food components having beneficial effects by enhancing the activity and growth of one or more colonic bacteria (45).

According to (46, 47) studies the rate of biofilm formation increased, the uptake of silver nanoparticles would be significantly reduced. Bacterial growth was suppressed by more than 90% when nanoparticles were present at a given concentration. The organism is unable to develop biofilm when exopolysaccharide synthesis is halted.

Characteristic or synergist effect of AgNPs, the release of Ag<sup>+</sup>, and the mode of action of AgNPs and bacterial extracts against bacteria all contributed to the increased antibacterial activity of AgNPs. The results of bacterial growth Bacteriostatic, bactericidal, and inhibitory of bacteria have all been shown to be effective against bacteria. The bacteriostatic, bactericidal, and biofilm inhibitory properties of AgNPs are all dependent on the concentration of AgNPs, according to (48).

Present findings are consistent with (49, 50) found that using antibiotics like ampicillin and ceftriaxone in combination with silver nanoparticles increased the efficiency (percentage) of antibiotics like ampicillin and ceftriaxone against bacteria.

Synthesized and characterized a [probiotic \*Bacillus licheniformis\*](#) cell free extract (BLCFE) coated [silver nanoparticles](#) (BLCFE-AgNPs). These BLCFE-AgNPs were characterized by UV-visible spectrophotometer, XRD, [EDX](#), FTIR, [TEM](#) and AFM. A strong [surface plasmon resonance](#) centered at 422 nm in UV-visible spectrum indicates the formation of AgNP, these results suggests that BLCFE-AgNps may be used for the control of biofilm forming [bacterial populations](#) in the biomedical field (51). (52) reported a different visible light irradiation effects on the formation of silver nanoparticles from silver nitrate using the culture supernatant of *Klebsiella pneumonia*.

## Conclusion

(53) and (54) indicated that antibiotic resistance against present antibiotics is rising at an alarming rate with need for discovery of advanced methods to treat infections caused by resistant pathogens. Silver nanoparticles are known to exhibit satisfactory antibacterial and antibiofilm activity against different pathogens, the percentage biofilm inhibition was evaluated to be 64% for *K. pneumoniae* strain [MF953600](#) and 86% for [MF953599](#) at AgNP concentration of 100 µg/ml. AgNPs were evaluated to be minimally cytotoxic and safe at concentrations of 15-120 µg/ml. The data evaluated provided evidence of AgNPs being safe antibacterial and antibiofilm compounds against MDR *K. pneumonia*. Contrary to our results, one of the previous studies by (55) demonstrated an increase in biofilm biomass of *Pseudomonas aeruginosa* after treatment of cells with superparamagnetic iron oxide nanoparticles at a concentration of 0.2 mg/ml. This study suggested that iron nanoparticles could be used as a source of elemental iron by the cell that is why they observed an increase in biofilm biomass with corresponding increase in cell density.

## Competition Interest

Authors of this research showed there is no conflicts of interest.

## References

1. AL-Saadi Z. In vitro Antagonistic Activity of Bifidobacterium breve Isolated from Breast-fed Infants Human Gastrointestinal Microflora against Two Clinical Strains of Staphylococcus aureus (MRSA and MSSA). Journal of Advances in Biology & Biotechnology. 2016;10(2):1-9.
2. Joint F. WHO Working Group on Drafting Guidelines for the Evaluation of Probiotics in Food: London. Ontario, Canada. 2002.
3. Guinane CM, Cotter PD. Role of the gut microbiota in health and chronic gastrointestinal disease: understanding a hidden metabolic organ. Therapeutic advances in gastroenterology. 2013;6(4):295-308.
4. Rahi AA, Nashaat Z. Leaky cells and the use of Lactobacillus casei in the treatment of laboratory animals infected with amoebic dysentery. Euphrates Journal of Agriculture Science. 2013;5(1):15-22.
5. Senok A, Ismaeel A, Botta G. Probiotics: facts and myths. Clinical Microbiology and Infection. 2005;11(12):958-66.



6. AL-Saadi ZN. Estimation of minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) of cell-free extracts of Bifidobacterium species against methicillin-resistant Staphylococcus aureus in vitro. American Journal of Biomedical and Life Sciences. 2016;4(5):75-80.
7. Makras L, Triantafyllou V, Fayol-Messaoudi D, Adriany T, Zoumpopoulou G, Tsakalidou E, et al. Kinetic analysis of the antibacterial activity of probiotic lactobacilli towards Salmonella enterica serovar Typhimurium reveals a role for lactic acid and other inhibitory compounds. Research in Microbiology. 2006;157(3):241-7.
8. Al-Saadi ZN. BACTERICIDAL ACTIVITY OF NEW SILVER NANOPARTICLES BIOSYNTHESIZED FROM METHICILLIN-RESISTANT STAPHYLOCOCCUS AUREUS AGAINST CLINICAL MRSA AND MSSA ISOLATES. Biochemical & Cellular Archives. 2020;20(2).
9. Aslam B, Wang W, Arshad MI, Khurshid M, Muzammil S, Rasool MH, et al. Antibiotic resistance: a rundown of a global crisis. Infection and drug resistance. 2018;1645-58.
10. Murray CJ, Ikuta KS, Sharara F, Swetschinski L, Aguilar GR, Gray A, et al. Global burden of bacterial antimicrobial resistance in 2019: a systematic analysis. The Lancet. 2022;399(10325):629-55.
11. Dadgostar P. Antimicrobial resistance: implications and costs. Infection and drug resistance. 2019;3903-10.
12. Giurazza R, Mazza MC, Andini R, Sansone P, Pace MC, Durante-Mangoni E. Emerging treatment options for multi-drug-resistant bacterial infections. Life. 2021;11(6):519.
13. Qais FA, Khan MS, Ahmad I. Nanoparticles as quorum sensing inhibitor: Prospects and limitations. Biotechnological applications of quorum sensing inhibitors. 2018;227-44.
14. Nagajyothi PC, Cha SJ, Yang IJ, Sreekanth T, Kim KJ, Shin HM. Antioxidant and anti-inflammatory activities of zinc oxide nanoparticles synthesized using Polygala tenuifolia root extract. Journal of Photochemistry and Photobiology B: Biology. 2015;146:10-7.
15. Manikandan R, Manikandan B, Raman T, Arunagirinathan K, Prabhu NM, Basu MJ, et al. Biosynthesis of silver nanoparticles using ethanolic petals extract of Rosa indica and characterization of its antibacterial, anticancer and anti-inflammatory activities. Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy. 2015;138:120-9.
16. Schachter B. Slimy business—the biotechnology of biofilms. Nature biotechnology. 2003;21(4):361-5.
17. Mohanpuria P, Rana NK, Yadav SK. Biosynthesis of nanoparticles: technological concepts and future applications. Journal of nanoparticle research. 2008;10:507-17.
18. Nishiyama K, Sugiyama M, Mukai T. Adhesion properties of lactic acid bacteria on intestinal mucin. Microorganisms. 2016;4(3):34.
19. Awadelkareem AM, Siddiqui AJ, Noumi E, Ashraf SA, Hadi S, Snoussi M, et al. Biosynthesized Silver Nanoparticles Derived from Probiotic Lactobacillus rhamnosus (AgNPs-LR) Targeting Biofilm Formation and Quorum Sensing-Mediated Virulence Factors. Antibiotics. 2023;12(6):986.
20. Issa RA, Al-Maamori JA, Al-Saadi ZN. OJVRTM.
21. Alquraishi FE, AL-Saadi ZN, Al-Azzawi JA. Detection of Biofilm Formation Among the Clinical Isolates of Klebsiella pneumoniae: Phenotypic and Genotypic Methods.
22. AL-Saadi Z. Study of ability of Lactobacillus acidophilus and Bifidobacterium bifidum in the prevention and treatment for enteritis induced by Salmonella typhimurium in Rats. College of Education Tikrit University. 2010.
23. Wayne P. Clinical and laboratory standards institute; 2007. Performance standards for antimicrobial susceptibility testing CLSI document M100-S17. 2005.
24. Al-Saady OMF, Zaki NH. THE EFFECT OF BIOSYNTHESIZED AG-NANOPARTICLES ON KLEBSIELLA PNEUMONIAE BIOFILM AND SOME VIRULENCE GENES. CHINESE JOURNAL OF MEDICAL GENETICS.32(4):2022.
25. Hashim A. Cytotoxic effects of silver nanoparticles prepared from Escherichia coli culture filtrates on vero cell line: M. Sc. Thesis. Mustansiriyah University; 2020.
26. Hussain Z. Inhibition of Biofilm using biological synthesis of silver nanoparticles from supernatants of E. coli: M. Sc. Thesis. Mustansiriyah University; 2016.

27. Namasivayam SKR, Preethi M, Bharani A, Robin G, Latha B. Biofilm inhibitory effect of silver nanoparticles coated catheter against *Staphylococcus aureus* and evaluation of its synergistic effects with antibiotics. *Int J Biol Pharm Res.* 2012;3(2):259-65.
28. Mohan AC, Renjanadevi B. Preparation of zinc oxide nanoparticles and its characterization using scanning electron microscopy (SEM) and X-ray diffraction (XRD). *Procedia Technology.* 2016;24:761-6.
29. Thakar MA, Jha SS, Phasinam K, Manne R, Qureshi Y, Babu VH. X ray diffraction (XRD) analysis and evaluation of antioxidant activity of copper oxide nanoparticles synthesized from leaf extract of *Cissus vitiginea*. *Materials Today: Proceedings.* 2022;51:319-24.
30. Clinical, Institute LS. Performance standards for antimicrobial susceptibility testing. *Clinical and Laboratory Standards Institute Wayne, PA;* 2017. p. 106-12.
31. Sutula J, Coulthwaite L, Verran J. Culture media for differential isolation of *Lactobacillus casei* Shirota from oral samples. *Journal of microbiological methods.* 2012;90(1):65-71.
32. Khodair ZT, Alzubaidy MWM, Almohaidi AMS, Sultan AA, AL-Shimmary SM, Albusultan SS, editors. Synthesis of copper oxide nanoparticles (CuO-NPs) and its evaluation of antibacterial activity against *P. aeruginosa* biofilm gene's. *AIP Conference Proceedings;* 2019: AIP Publishing.
33. Muhsin EA, Said LA, Al-Jubori SS. Correlation of type 1 and type 3 Fimbrial genes with the type of specimen and the antibiotic resistance profile of clinically isolated *Klebsiella pneumoniae* in Baghdad. *Al-Mustansiriyah Journal of Science.* 2022;33(3):1-11.
34. Alzubaidy M, editor Almohaidi. AM S, Sultan A. A, AL-Shimmary, SMH (2019). Virulence Gene of *Pseudomonas aeruginosa* with Nanoparticle. *AIP Conference Proceedings.*
35. Hassan NS, Al-marjani MF, Hussain NH. Detection of Antiseptic Resistant Genes in Colistin-Resistant *Pseudomonas aeruginosa* and MDR *Klebsiella pneumoniae*. *Indian Journal of Forensic Medicine & Toxicology.* 2021;15(1).
36. Iliadis I, Daskalopoulou A, Simões M, Giaouris E. Integrated combined effects of temperature, pH and sodium chloride concentration on biofilm formation by *Salmonella enterica* ser. Enteritidis and Typhimurium under low nutrient food-related conditions. *Food Research International.* 2018;107:10-8.
37. Mathur T, Singhal S, Khan S, Upadhyay D, Fatma T, Rattan A. Detection of biofilm formation among the clinical isolates of staphylococci: an evaluation of three different screening methods. *Indian journal of medical microbiology.* 2006;24(1):25-9.
38. Samie A, Nkgau T. Biofilm production and antibiotic susceptibility profile of *Escherichia coli* isolates from HIV and AIDS patients in the Limpopo Province. *African Journal of Biotechnology.* 2012;11(34):8560-70.
39. Bose S, Khodke M, Basak S, Mallick S. Detection of biofilm producing staphylococci: need of the hour. *J Clin Diagn Res.* 2009;3(6):1915-20.
40. de Oliveira Silva BS, Seabra AB. Characterization of iron nanoparticles produced with green tea extract: a promising material for nitric oxide delivery. *Biointerface Research in Applied Chemistry.* 2016;6(3).
41. Kumar D, Kumar G, Agrawal V. Green synthesis of silver nanoparticles using *Holarrhena antidysenterica* (L.) Wall. bark extract and their larvicidal activity against dengue and filariasis vectors. *Parasitology research.* 2018;117:377-89.
42. Sharma S, Mudgal N, Sharma P, Shrngi B. Comparison of phenotypic characteristics and virulence traits of *Klebsiella pneumoniae* obtained from pneumonic and healthy camels (*Camelus dromedarius*). *Adv Anim Vet Sci.* 2015;3(2):116-22.
43. Ansari M, Khan H, Khan A, Cameotra S, Alzohairy M. Anti-biofilm efficacy of silver nanoparticles against MRSA and MRSE isolated from wounds in a tertiary care hospital. *Indian journal of medical microbiology.* 2015;33(1):101-9.
44. Abdel-Daim A, Hassouna N, Hafez M, Ashor MSA, Aboulwafa MM. Antagonistic activity of *Lactobacillus* isolates against *Salmonella typhi* in vitro. *BioMed Research International.* 2013;2013.
45. Karimi R, Azizi MH, Ghasemlou M, Vaziri M. Application of inulin in cheese as prebiotic, fat replacer and texturizer: A review. *Carbohydrate polymers.* 2015;119:85-100.
46. Bakht M, Alizadeh SA, Rahimi S, Kazemzadeh Anari R, Rostamani M, Javadi A, et al. Phenotype and genetic determination of resistance to common disinfectants among biofilm-

- producing and non-producing *Pseudomonas aeruginosa* strains from clinical specimens in Iran. *BMC microbiology*. 2022;22(1):124.
47. Martinez-Gutierrez F, Boegli L, Agostinho A, Sánchez EM, Bach H, Ruiz F, et al. Anti-biofilm activity of silver nanoparticles against different microorganisms. *Biofouling*. 2013;29(6):651-60.
  48. Keshari A, Srivastava R, Yadav S, Nath G, Gond S. Synergistic activity of green silver nanoparticles with antibiotics. *Nanomedicine Research Journal*. 2020;5(1):44-54.
  49. Krishna G, Kumar SS, Pranitha V, Alha M, Charaya S. Biogenic synthesis of silver nanoparticles and their synergistic effect with antibiotics: a study against *Salmonella* sp. *Int J Pharm Pharm Sci*. 2015;7(11):84-8.
  50. Kapadia C, Alhazmi A, Patel N, Elesawy BH, Sayyed R, Lokhandwala F, et al. Nanoparticles combined with cefixime as an effective synergistic strategy against *Salmonella enterica* typhi. *Saudi Journal of Biological Sciences*. 2021;28(8):4164-72.
  51. Shanthi S, Jayaseelan BD, Velusamy P, Vijayakumar S, Chih CT, Vaseeharan B. Biosynthesis of silver nanoparticles using a probiotic *Bacillus licheniformis* Dahb1 and their antibiofilm activity and toxicity effects in *Ceriodaphnia cornuta*. *Microbial Pathogenesis*. 2016;93:70-7.
  52. Mokhtari N, Daneshpajouh S, Seyedbagheri S, Atashdehghan R, Abdi K, Sarkar S, et al. Biological synthesis of very small silver nanoparticles by culture supernatant of *Klebsiella pneumoniae*: The effects of visible-light irradiation and the liquid mixing process. *Materials research bulletin*. 2009;44(6):1415-21.
  53. Siddique MH, Aslam B, Imran M, Ashraf A, Nadeem H, Hayat S, et al. Effect of silver nanoparticles on biofilm formation and EPS production of multidrug-resistant *Klebsiella pneumoniae*. *Biomed research international*. 2020;2020:1-9.
  54. Suriati G, Mariatti M, Azizan A. Synthesis of silver nanoparticles by chemical reduction method: Effect of reducing agent and surfactant concentration. *International journal of automotive and mechanical engineering*. 2014;10:1920-7.
  55. Haney C, Rowe JJ, Robinson JB. Spions increase biofilm formation by *Pseudomonas aeruginosa*. *Journal of Biomaterials and Nanobiotechnology*. 2012;3(04):508

### Authors contribution

Hajar T. Mahdi: Material preparing, questionnaire designer, data collection, explaining of outcomes and writing on article.

Prof Dr. Zainab N. Al-Saadi: Article proposal designer, statistical analysis, editing and writing.