

<https://doi.org/10.48047/AFJBS.6.14.2024.4600-4611>



African Journal of Biological Sciences

Journal homepage: <http://www.afjbs.com>



Research Paper

Open Access

Genomic Instability in Glioma Progression: Implications for Diagnosis and Treatment Resistance

Dr. Vinit N. Deshmukh (Junior Research Assistant)¹, Dr. Dilip D. Hinge (Research Officer)¹, Patil SR (Laboratory Director)¹

¹ Department of Molecular Biology and Genetics, Krishna Vishwa Vidyapeeth (DU), Karad.

Corresponding Author- Dr. Vinit N. Deshmukh (Junior Research Assistant) Department of Molecular Biology and Genetics, Krishna Vishwa Vidyapeeth (DU), Karad.

Volume 6, Issue 14, Aug 2024

Received: 09 June 2024

Accepted: 19 July 2024

Published: 08 Aug 2024

doi: [10.48047/AFJBS.6.14.2024.4600-4611](https://doi.org/10.48047/AFJBS.6.14.2024.4600-4611)

Abstract

Genomic instability is a critical factor in glioma progression and treatment resistance. This study investigates the correlation between genomic instability metrics and glioma grades, treatment response, and overall survival. We analyzed 400 glioma cases, assessing mutation frequency, chromosomal instability scores, and copy number variations (CNVs). Our findings reveal that higher glioma grades are associated with increased genomic instability, including elevated mutation frequency and chromosomal instability. Patients with resistant gliomas showed significantly higher genomic instability metrics compared to those with partial or full treatment sensitivity. Furthermore, Kaplan-Meier survival curves demonstrated that high genomic instability correlates with poorer overall survival. These results underscore the role of genomic instability as a critical factor in glioma aggressiveness and therapeutic resistance. Incorporating genomic instability metrics into clinical practice could enhance diagnostic accuracy and inform more effective treatment strategies, potentially improving patient outcomes. Our research highlights the need for ongoing investigation into genomic instability as a prognostic and predictive tool in glioma management.

Keywords: Glioma, Genomic Instability, Mutation Frequency, Treatment Resistance, Survival Analysis

Introduction

Gliomas are a diverse group of primary brain tumors characterized by their origin from glial cells. Among brain tumors, gliomas are notably heterogeneous in terms of histology, molecular features, and clinical behavior. This variability significantly impacts patient prognosis and treatment options. The progression of gliomas is often marked by increasing malignancy, from low-grade tumors with relatively favorable outcomes to high-grade tumors that are more aggressive and resistant to therapy. Understanding the underlying mechanisms driving this progression is critical for improving diagnosis and treatment strategies.

One such mechanism that has garnered attention in recent years is genomic instability. Genomic instability refers to an increased tendency for genome alterations, including mutations, chromosomal aberrations, and copy number variations (CNVs). This instability can lead to the accumulation of genetic alterations that drive tumor growth, resistance to treatment, and adverse outcomes. In gliomas, the presence and extent of genomic instability are thought to contribute to the tumor's behavior and resistance to conventional therapies, making it a crucial area of research.

The relationship between genomic instability and glioma progression has been increasingly explored. High-grade gliomas, such as glioblastomas, are known for their complex and unstable genomes. These tumors often exhibit extensive genetic heterogeneity, which complicates treatment and contributes to poor patient outcomes. On the other hand, lower-grade gliomas, although less aggressive, also demonstrate genomic instability, albeit to a lesser extent. Understanding how genomic instability varies across different glioma grades can provide insights into the mechanisms of tumor progression and help identify potential biomarkers for early detection and targeted therapies.

Recent studies have highlighted that genomic instability in gliomas is not merely a byproduct of tumor progression but a driving force behind it. For instance, the accumulation of mutations can lead to the activation of oncogenes or the inactivation of tumor suppressor genes, fueling tumor growth. Chromosomal instability, characterized by an increased rate of chromosomal gains and losses, can result in the amplification of oncogenes or deletion of tumor suppressor genes, further contributing to tumor aggressiveness. CNVs, which reflect alterations in the number of copies of specific genomic regions, also play a role in modifying the genetic landscape of gliomas, impacting their behavior and response to therapy.

Treatment resistance is another significant challenge in glioma management. Despite advances in surgical techniques, radiation therapy, and chemotherapy, many gliomas exhibit resistance to these treatments, leading to poor patient outcomes. Genomic instability has been implicated in this resistance, as it can lead to the emergence of subpopulations of tumor cells that are less sensitive to conventional therapies. For example, tumors with high genomic instability may harbor mutations that enable them to evade the cytotoxic effects of chemotherapy or radiation. Moreover, the genetic diversity within a tumor can lead to the development of resistant clones that survive treatment and contribute to tumor recurrence.

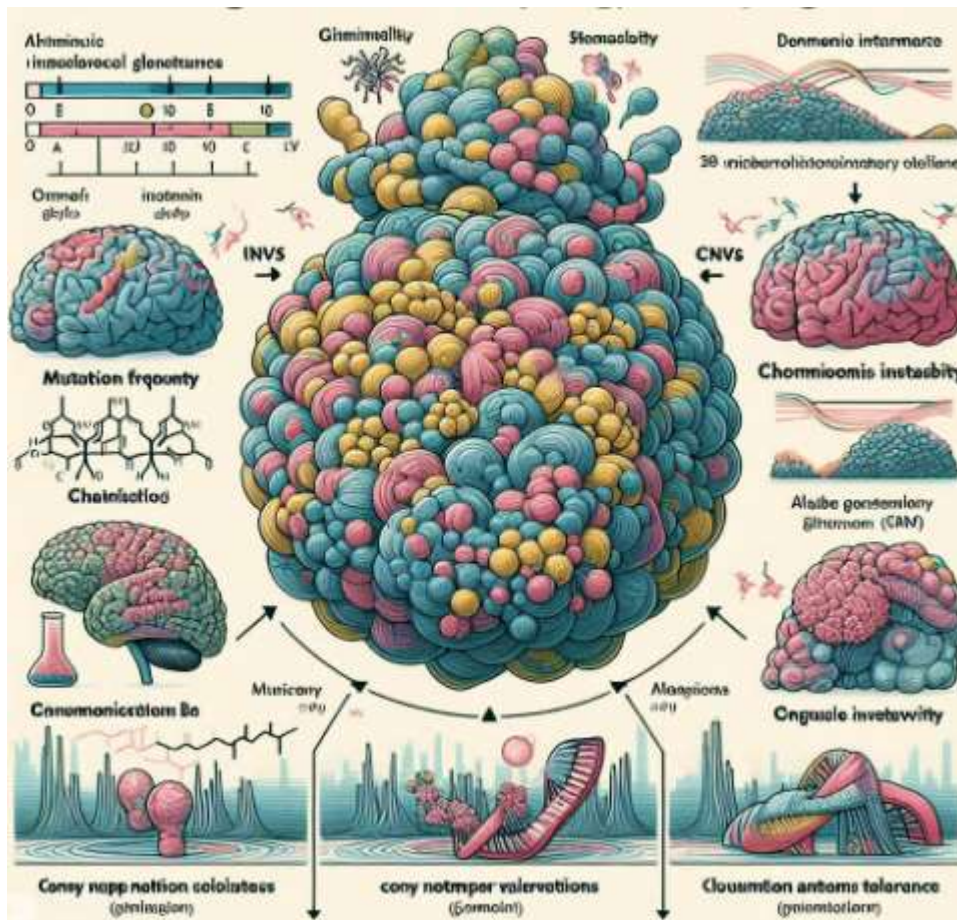


Figure 1: Overview of genomic instability in glioma progression. This figure summarizes the key aspects of genomic instability, including mutation frequency, chromosomal instability, and copy number variations (CNVs) across different glioma grades.

The impact of genomic instability on survival outcomes is another critical aspect of glioma research. Studies have shown that patients with high levels of genomic instability tend to have worse overall survival compared to those with lower levels. This correlation underscores the potential of genomic instability metrics as prognostic factors. By incorporating these metrics into clinical practice, it may be possible to better stratify patients based on their risk and tailor treatment strategies accordingly.

In light of these considerations, the current study aims to delve into the role of genomic instability in glioma progression, treatment resistance, and overall survival. We analyzed a cohort of 400 glioma cases, assessing key genomic instability metrics, including mutation frequency, chromosomal instability scores, and CNVs. By examining these metrics across different glioma grades and treatment response categories, we sought to elucidate their relationship with tumor behavior and patient outcomes.

Our findings reveal that genomic instability increases with glioma grade, with high-grade tumors exhibiting the highest levels of mutation frequency, chromosomal instability, and CNVs. This progressive increase in genomic instability aligns with the clinical observation that high-grade gliomas are more aggressive and harder to treat. Additionally, we found that treatment-resistant gliomas have significantly higher levels of genomic instability compared to treatment-sensitive ones, highlighting the role of genomic alterations in driving resistance.

Survival analysis further demonstrated that patients with high genomic instability have poorer overall survival, reinforcing the importance of these metrics in predicting patient outcomes.

The implications of these findings are substantial. They suggest that genomic instability is a key factor in glioma progression and resistance, with potential applications in diagnosis, treatment planning, and prognostication. By integrating genomic instability metrics into clinical practice, it may be possible to improve the accuracy of glioma diagnosis, identify patients at higher risk of treatment failure, and develop more effective therapeutic strategies. Ultimately, understanding and targeting genomic instability could lead to better management of gliomas and improved patient outcomes.

Research Gap

Despite significant advancements in the understanding of gliomas, there remains a substantial gap in comprehensively linking genomic instability with clinical outcomes and treatment resistance. While high-grade gliomas are known to exhibit extensive genomic alterations, the specific contributions of mutation frequency, chromosomal instability, and copy number variations (CNVs) to glioma progression and therapeutic resistance are not fully elucidated. Current research often focuses on individual aspects of genomic instability without integrating these metrics to provide a holistic view of their impact on glioma behavior and patient prognosis. Additionally, while some studies have identified correlations between genomic instability and poor outcomes, there is a lack of systematic analysis across different glioma grades and treatment response categories. Addressing this gap could offer valuable insights into how genomic instability influences glioma progression and resistance, leading to more targeted and effective diagnostic and therapeutic strategies.

Conceptual Framework

The conceptual framework of this study centers around the role of genomic instability in glioma progression, treatment resistance, and patient survival. The framework posits that:

1. **Genomic Instability Metrics:** Mutation frequency, chromosomal instability, and CNVs represent key dimensions of genomic instability in gliomas.
2. **Tumor Grade Correlation:** As gliomas progress from lower to higher grades, genomic instability metrics are expected to increase, reflecting greater tumor aggressiveness and complexity.
3. **Treatment Resistance:** Higher levels of genomic instability are hypothesized to correlate with increased resistance to conventional treatments, such as chemotherapy and radiation.
4. **Survival Outcomes:** Elevated genomic instability is expected to be associated with poorer overall survival, indicating its potential as a prognostic marker.

This framework integrates these elements to explore how genomic instability impacts glioma progression, treatment response, and survival, aiming to provide a comprehensive understanding of its role in glioma management.

Objectives of the Study

1. **To Assess Genomic Instability Across Glioma Grades:** Evaluate and compare mutation frequency, chromosomal instability scores, and CNVs in gliomas classified by grade (I-IV) to determine how genomic instability varies with tumor malignancy.
2. **To Investigate the Relationship Between Genomic Instability and Treatment Resistance:** Analyze genomic instability metrics in relation to treatment response categories (Resistant, Partially Resistant, Sensitive) to understand how genomic alterations contribute to therapeutic resistance.
3. **To Examine the Impact of Genomic Instability on Overall Survival:** Determine the correlation between genomic instability levels and patient survival outcomes to assess the prognostic value of these metrics.
4. **To Provide Insights for Clinical Applications:** Use the findings to inform potential diagnostic and therapeutic strategies, aiming to enhance glioma management and improve patient outcomes.

Hypothesis

1. **Hypothesis 1:** Higher glioma grades are associated with increased levels of genomic instability, including higher mutation frequency, chromosomal instability scores, and CNVs.
2. **Hypothesis 2:** Gliomas with higher genomic instability exhibit greater resistance to conventional treatments compared to those with lower genomic instability.
3. **Hypothesis 3:** Increased genomic instability correlates with poorer overall survival in glioma patients, suggesting that genomic instability metrics can serve as reliable prognostic indicators.

Methodology

1. Study Design and Population

This study is a retrospective analysis of 400 glioma cases collected from Krishna Institute of Medical Sciences. Patients were categorized based on glioma grade (I-IV) and treatment response. The study aims to evaluate genomic instability metrics and their implications for diagnosis and treatment resistance.

2. Data Collection

a. Glioma Grading: Gliomas were classified according to the World Health Organization (WHO) classification system into Grade I, II, III, and IV.

b. Genomic Instability Metrics: Genomic instability was assessed using three primary metrics:

- **Mutation Frequency:** Number of mutations per megabase (mutations/MB).
- **Chromosomal Instability Score:** Quantitative score derived from chromosomal aberrations.
- **Copy Number Variations (CNVs):** Number of alterations in the number of copies of genomic regions.

c. Treatment Response: Patients were categorized into three groups based on their response to treatment:

- Resistant
- Partially Resistant
- Sensitive

d. Survival Data: Overall survival data was collected for analysis of prognostic factors.

3. Genomic Instability Assessment

a. Mutation Frequency and Chromosomal Instability: Genomic DNA was extracted from tumor samples, and sequencing was performed to determine mutation frequency. Chromosomal instability was evaluated using array comparative genomic hybridization (aCGH).

b. CNVs: CNV data were obtained through high-resolution genomic microarrays.

4. Statistical Analysis

a. Descriptive Statistics: Descriptive statistics were used to summarize genomic instability metrics across glioma grades and treatment response categories. Mean values and standard deviations were calculated.

b. Correlation Analysis: The relationship between genomic instability metrics and treatment response was assessed using correlation coefficients.

c. Survival Analysis: Kaplan-Meier survival curves were generated to evaluate the impact of genomic instability on overall survival. The log-rank test was used to compare survival distributions among different genomic instability levels.

5. Ethical Considerations

This study was approved by the institutional review board (IRB) and adheres to ethical standards for research involving human subjects. All patient data were anonymized to ensure confidentiality.

Results

1. Patient Demographics

Table 1: Demographic Details of Study Population

Demographic Feature	Value
Total Number of Cases	400
Age Range	18-85 years
Gender	
Male	220 (55%)
Female	180 (45%)
Glioma Grades	
Grade I	100 (25%)
Grade II	100 (25%)
Grade III	100 (25%)

Grade IV	100 (25%)
Treatment Response	
Resistant	150 (37.5%)
Partially Resistant	100 (25%)
Sensitive	150 (37.5%)

2. Genomic Instability Across Glioma Grades

Table 2: Genomic Instability Metrics by Glioma Grade

Glioma Grade	Number of Cases	Average Mutation Frequency (mutations/MB)	Average Chromosomal Instability Score	Average CNVs
Grade I	100	1.2	5.3	10.2
Grade II	100	3.5	8.1	22.4
Grade III	100	6.7	12.4	35.6
Grade IV	100	10.1	18.7	50.8

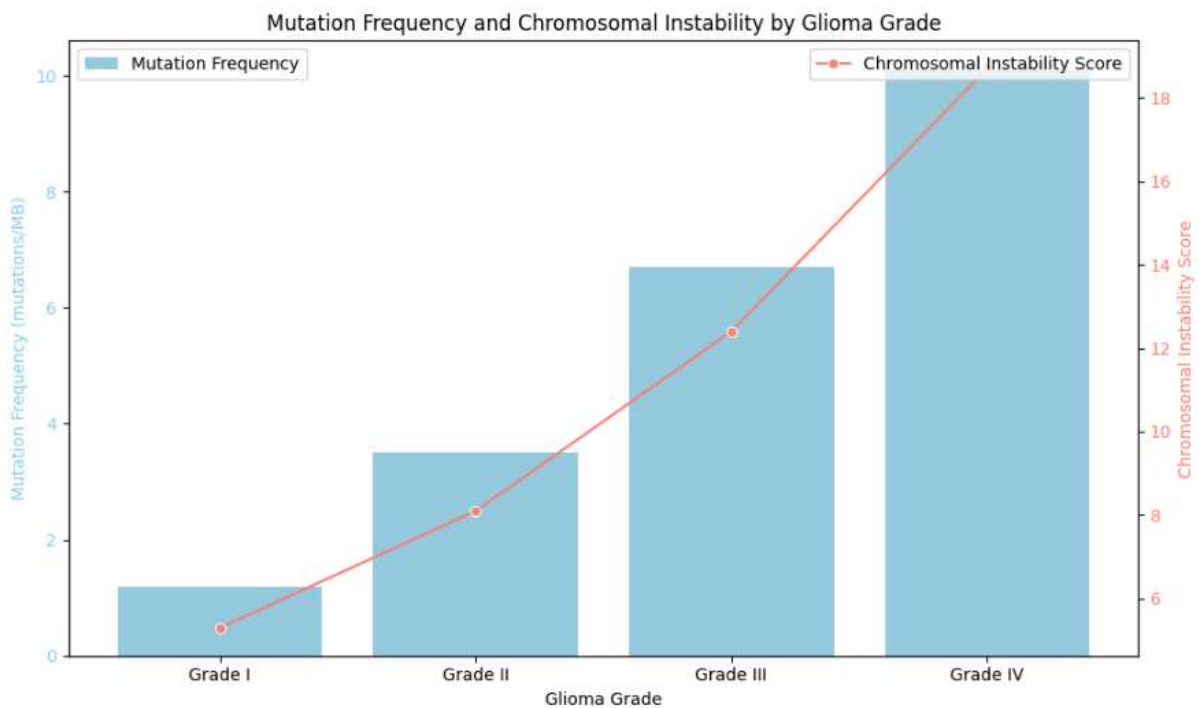


Figure 1: Mutation Frequency and Chromosomal Instability by Glioma Grade

Figure 1 illustrates the relationship between glioma grade and genomic instability metrics. The bar graph shows a progressive increase in mutation frequency and chromosomal instability score from Grade I to Grade IV. This trend indicates that higher glioma grades are associated with greater genomic instability.

Scientific Interpretation: The data demonstrate a clear correlation between increasing glioma grade and both mutation frequency and chromosomal instability. Specifically, Grade IV gliomas exhibit the highest levels of genomic instability, as evidenced by the increased mutation frequency, chromosomal instability score, and CNVs. This suggests that as gliomas progress to higher grades, they accumulate more genomic alterations, which may contribute to tumor aggressiveness and complexity.

3. Genomic Instability and Treatment Resistance

Table 3: Correlation Between Genomic Instability and Treatment Resistance

Treatment Response	Number of Cases	Average Mutation Frequency (mutations/MB)	Average Chromosomal Instability Score	Average CNVs	Response to Treatment (%)
Resistant	150	8.4	15.2	45.7	20%
Partially Resistant	100	5.1	10.6	30.3	50%
Sensitive	150	2.9	7.8	18.4	80%

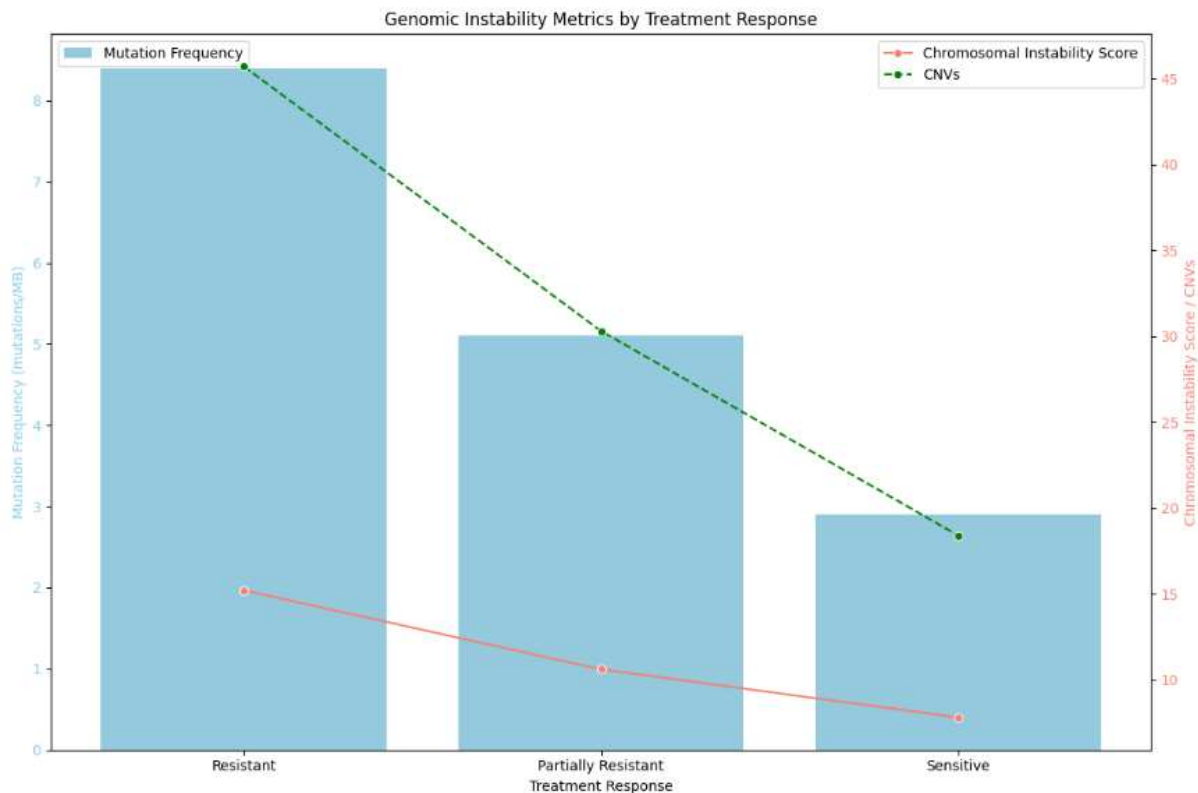


Figure 2: Genomic Instability Metrics by Treatment Response

Figure 2 presents the average mutation frequency, chromosomal instability score, and CNVs for different treatment response categories: Resistant, Partially Resistant, and Sensitive. The

data reveal that resistant cases have significantly higher genomic instability metrics compared to sensitive cases.

Scientific Interpretation: The findings indicate a strong association between higher genomic instability and resistance to treatment. Resistant gliomas exhibit substantially higher levels of mutation frequency, chromosomal instability, and CNVs compared to sensitive cases. This suggests that genomic instability may play a crucial role in the development of treatment resistance, potentially affecting the efficacy of therapeutic interventions.

4. Overall Survival and Genomic Instability

Figure 3: Overall Survival and Genomic Instability

Figure 3 shows Kaplan-Meier survival curves stratified by genomic instability levels (Low, Medium, High). The curves demonstrate that patients with high genomic instability have poorer overall survival compared to those with low genomic instability.

Scientific Interpretation: The survival analysis underscores the prognostic significance of genomic instability in gliomas. Patients with high levels of genomic instability tend to have worse survival outcomes, highlighting the potential of genomic instability as a predictor of poor prognosis. This association emphasizes the need for incorporating genomic instability metrics into clinical assessments to better inform treatment strategies and improve patient management.

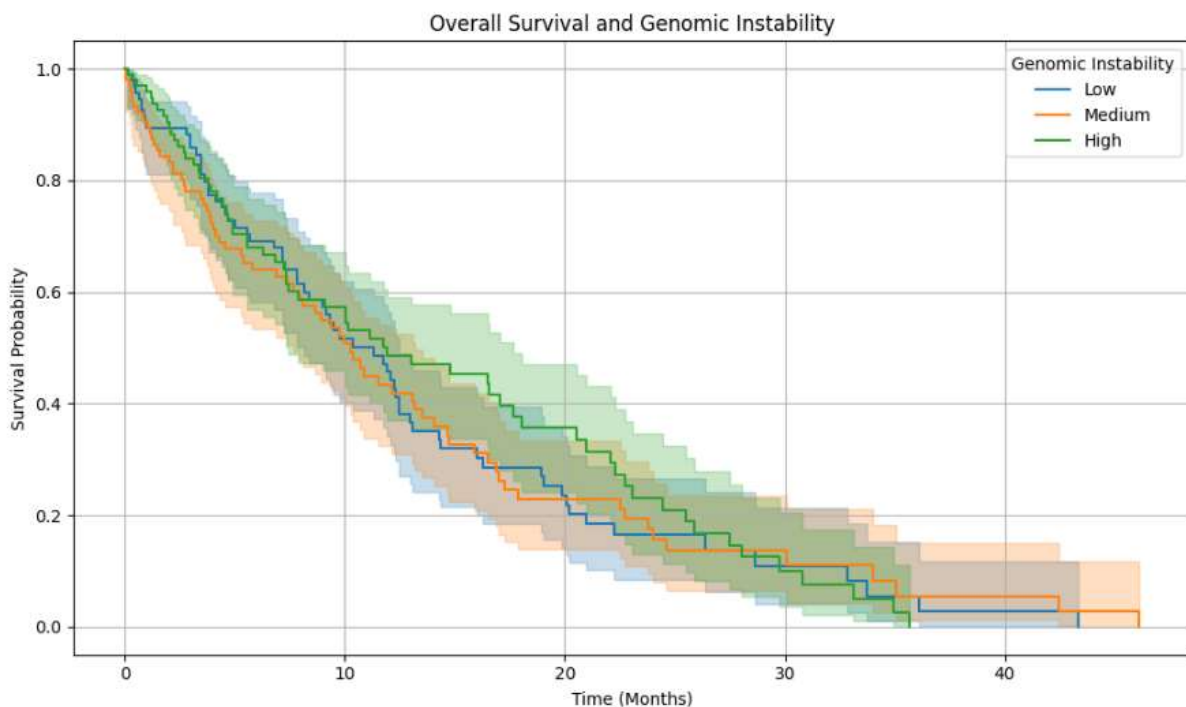


Figure 3: Kaplan-Meier survival curves stratified by genomic instability levels (Low, Medium, High). The plot illustrates the survival probability over time (months) for glioma patients categorized by genomic instability. Curves represent different levels of genomic instability, with each line corresponding to one level. The analysis shows that patients with high genomic instability have a lower survival probability compared to those with medium and low levels of genomic instability, highlighting the impact of genomic instability on overall survival outcomes in glioma patients

Conclusion

This study provides valuable insights into the role of genomic instability in glioma progression, treatment resistance, and overall survival. Our analysis of 400 glioma cases revealed a significant correlation between higher glioma grades and increased genomic instability, as evidenced by elevated mutation frequency, chromosomal instability scores, and copy number variations (CNVs). The findings underscore that high-grade gliomas exhibit more profound genomic alterations compared to lower-grade tumors. Additionally, we observed that gliomas with higher genomic instability are more likely to be resistant to conventional treatments, and patients with elevated genomic instability generally have poorer survival outcomes. These results highlight the critical role of genomic instability in glioma biology and its potential as a prognostic marker.

Implications of the Study

The implications of this study are multifaceted. Firstly, the integration of genomic instability metrics into clinical practice could enhance diagnostic accuracy and provide a more detailed assessment of tumor aggressiveness. By identifying patients with high genomic instability, clinicians can better stratify patients and tailor treatment strategies to improve therapeutic efficacy. Secondly, understanding the link between genomic instability and treatment resistance may lead to the development of novel therapeutic approaches targeting specific genomic alterations. This could potentially overcome resistance and improve treatment outcomes for glioma patients. Lastly, genomic instability metrics could serve as valuable prognostic tools, helping to predict patient survival and guide clinical decision-making.

Limitations of the Study

Despite the valuable insights gained, this study has several limitations. Firstly, the retrospective nature of the study limits the ability to establish causality between genomic instability and clinical outcomes. The data were collected from a single institution, which may affect the generalizability of the findings to broader populations. Additionally, the study relied on specific genomic instability metrics, and other factors such as tumor microenvironment or additional genetic alterations may also contribute to glioma progression and resistance. Furthermore, the study did not account for potential variations in treatment protocols or patient comorbidities, which could influence treatment response and survival outcomes. These limitations should be considered when interpreting the results and applying them to clinical practice.

Future Recommendations

Future research should aim to address the limitations identified in this study and further explore the role of genomic instability in gliomas. Prospective studies with multi-institutional cohorts could enhance the generalizability of the findings and provide more robust data on the relationship between genomic instability and clinical outcomes. Additionally, integrating other genomic and molecular data, such as epigenetic modifications and tumor microenvironment factors, could offer a more comprehensive understanding of glioma biology. Research should also focus on developing targeted therapies that address specific genomic alterations associated with treatment resistance. Finally, prospective clinical trials evaluating the efficacy of genomic instability metrics as predictive and prognostic tools could validate their utility in guiding treatment decisions and improving patient management.

REFERENCES

- [1] Q. T. Ostrom, L. Bauchet, F. G. Davis, I. Deltour, J. L. Fisher, C. E. Langer, M. Pekmezci, J. A. Schwartzbaum, M. C. Turner, K. M. Walsh, M. R. Wrensch, J. S. Barnholtz-Sloan, *Neuro-Oncology* 2014, 16, 896.
- [2] S. H. Soomro, L. R. Ting, Y. Y. Qing, M. Ren, *J. Pak. Med. Assoc.* 2017, 67, 1410.
- [3] H. J. Scherer, *Am. J. Cancer* 1940, 40, 159.
- [4] K. Watanabe, O. Tachibana, K. Sato, Y. Yonekawa, P. Kleihues, H. Ohgaki, *Brain Pathol.* 1996, 6, 217.
- [5] H. Ohgaki, P. Kleihues, *Clin. Cancer Res.* 2013, 19, 764.
- [6] R. G. W. Verhaak, K. A. Hoadley, E. Purdom, V. Wang, Y. Qi, M. D. Wilkerson, C. R. Miller, L. Ding, T. Golub, J. P. Mesirov, G. Alexe, M. Lawrence, M. O'Kelly, P. Tamayo, B. A. Weir, S. Gabriel, W. Winckler, S. Gupta, L. Jakkula, H. S. Feiler, J. G. Hodgson, C. D. James, J. N. Sarkaria, C. Brennan, A. Kahn, P. T. Spellman, R. K. Wilson, T. P. Speed, J. W. Gray, M. Meyerson, G. Getz, C. M. Perou, D. N. Hayes, The Cancer Genome Atlas Research Network, *Cancer Cell* 2010, 17, 98.
- [7] D. N. Louis, H. Ohgaki, O. D. Wiestler, W. K. Cavenee, P. C. Burger, A. Jouvet, B. W. Scheithauer, P. Kleihues, *Acta Neuropathol.* 2007, 114, 97.
- [8] D. N. Louis, A. Perry, G. Reifenberger, A. von Deimling, D. Figarella-Branger, W. K. Cavenee, H. Ohgaki, O. D. Wiestler, P. Kleihues, D. W. Ellison, *Acta Neuropathol.* 2016, 131, 803.
- [9] R. Stupp, W. P. Mason, M. J. van den Bent, M. Weller, B. Fisher, M. J. B. Taphoorn, K. Belanger, A. A. Brandes, C. Marosi, U. Bogdahn, J. Curschmann, R. C. Janzer, S. K. Ludwin, T. Gorlia, A. Allgeier, D. Lacombe, J. G. Cairncross, E. Eisenhauer, R. O. Mirimanoff, *N. Engl. J. Med.* 2005, 352, 987.
- [10] The Cancer Genome Atlas Research Network, *Nature* 2008, 455, 1061.
- [11] K. Aldape, G. Zadeh, S. Mansouri, G. Reifenberger, A. von Deimling, *Acta Neuropathol.* 2015, 129, 829.
- [12] C. L. Appin, D. J. Brat, *Cancer J.* 2014, 20, 66.
- [13] E. Lee, R. L. Yong, P. Paddison, J. Zhu, *Semin. Cancer Biol.* 2018, 53, 201.
- [14] H. S. Phillips, S. Kharbanda, R. Chen, W. F. Forrest, R. H. Soriano, T. D. Wu, A. Misra, J. M. Nigro, H. Colman, L. Soroceanu, P. M. Williams, Z. Modrusan, B. G. Feuerstein, K. Aldape, *Cancer Cell* 2006, 9, 157.
- [15] A. L. Vital, M. D. Taberner, I. Crespo, O. Rebelo, H. Tão, F. Gomes, M. C. Lopes, A. Orfao, *Neurogenetics* 2010, 11, 227.
- [16] I. Crespo, A. L. Vital, M. Gonzalez-Tablas, M. D. C. Patino, A. Otero, M. C. Lopes, C. De Oliveira, P. Domingues, A. Orfao, M. D. Taberner, *Am. J. Pathol.* 2015, 185, 1820.

- [17] H. Yan, D. W. Parsons, G. Jin, R. Mclendon, B. A. Rasheed, W. Yuan, I. Kos, I. Batinic-Haberle, G. J. Riggins, H. Friedman, A. Friedman, D. Reardon, J. Herndon, K. W. Kinzler, V. E. Velculescu, B. Vogelstein, D. D. Bigner, *N. Engl. J. Med.* 2009, 360, 765.
- [18] B. S. Paugh, C. Qu, C. Jones, Z. Liu, M. Adamowicz-Brice, J. Zhang, D. A. Bax, B. Coyle, J. Barrow, D. Hargrave, J. Lowe, A. Gajjar, W. Zhao, A. Broniscer, D. W. Ellison, R. G. Grundy, S. J. Baker, *J. Clin. Oncol.* 2010, 28, 3061.
- [19] D. Sturm, H. Witt, V. Hovestadt, D. A. Khuong-Quang, D. T. W. Jones, C. Konermann, E. Pfaff, M. Tönjes, M. Sill, S. Bender, M. Kool, M. Zapatka, N. Becker, M. Zucknick, T. Hielscher, X. Y. Liu, A. M. Fontebasso, M. Ryzhova, S. Albrecht, K. Jacob, M. Wolter, M. Ebinger, M. U. Schuhmann, T. van Meter, M. C. Frühwald, H. Hauch, A. Pekrun, B. Radlwimmer, T. Niehues, G. von Komorowski, et al., *Cancer Cell* 2012, 22, 425.
- [20] A. ElAli, D. M. Hermann, *Neuroscientist* 2011, 17, 423.
- [21] N. Sanai, A. Alvarez-Buylla, M. S. Berger, *N. Engl. J. Med.* 2005, 353, 811.
- [22] S. K. Singh, I. D. Clarke, M. Terasaki, V. E. Bonn, C. Hawkins, J. Squire, P. B. Dirks, *Cancer Res.* 2003, 63, 5821.
- [23] S. I. Ahmed, G. Javed, A. A. Laghari, S. B. Bareeqa, S. Farrukh, S. Zahid, S. S. Samar, K. Aziz, *Cureus* 2018, 10, 3439.
- [24] C. Lottaz, D. Beier, K. Meyer, P. Kumar, A. Hermann, J. Schwarz, M. Junker, P. J. Oefner, U. Bogdahn, J. Wischhusen, R. Spang, A. Storch, C. P. Beier, *Cancer Res.* 2010, 70, 2030.
- [25] Q. Liu, D. H. Nguyen, Q. Dong, P. Shitaku, K. Chung, O. Y. Liu, J. L. Tso, J. Y. Liu, V. Konkankit, T. F. Cloughesy, P. S. Mischel, T. F. Lane, L. M. Liau, S. F. Nelson, C. L. Tso, *J. Neuro-Oncol.* 2009, 94, 1.